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RESEARCH ARTICLE

Ocimum basilicum (kemangi) intervention on powder and microencapsulated *Spirulina platensis* and its bioactive molecules [version 1; peer review: 2 approved with reservations]

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v1 First published: 18 Jun 2021, 10:485
<https://doi.org/10.12688/f1000research.52394.1>
 Second version: 05 Nov 2021, 10:485
<https://doi.org/10.12688/f1000research.52394.2>
 Latest published: 14 Mar 2022, 10:485
<https://doi.org/10.12688/f1000research.52394.3>

Abstract

Background: *Spirulina platensis* contains several bioactive molecules such as phenol, flavonoid and phycocyanin pigments. This study unveils total phenol, flavonoid, antioxidant activity, phycocyanin content and evaluated encapsulation efficiency from *Ocimum basilicum* intervention on *S. platensis*. *O. basilicum* intervention aims to reduce unpleasant odors from *S. platensis* that will increase consumption and increase bioactive compounds.

Methods: The intervention was carried out by soaking a *S. platensis* control sample (SP) in *O. basilicum* with a ratio of 1:4 (w/v) and it was then dried (DSB) and microencapsulated by freeze drying methods (MSB) using a combination of maltodextrin and gelatin. Total flavonoid and phenolic analysis with curve fitting analysis used a linear regression approach. Antioxidant activity of samples was analysed with the 2,2'-azino-bis-3-thylbenzthiazoline-6-sulphonic acid (ABTS) method. Data were analysed using ANOVA at significance level ($p < 0.05$) followed by Tukey test models using SPSS v.22.

Results: The result of this study indicated that *O. basilicum* intervention treatment (DSB) has the potential to increase bioactive compounds such as total phenol, antioxidant activity and phycocyanin, and flavonoid content. Intervention of *O. basilicum* on *S. platensis* (DSB) significantly increases total phenol by 48.7% and

Open Peer Review

Approval Status

	1	2
version 3		
(revision)		view
14 Mar 2022		
version 2		
(revision)	view	view
05 Nov 2021		
version 1		
18 Jun 2021	view	view

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phycocyanin by 40.7%. This is due to the phenol and azulene compounds in *O. basilicum* which have a synergistic effect on phenol and phycocyanin in *S. platensis*. Microencapsulation using a maltodextrin and gelatin coating is effective in phycocyanin protection with an encapsulation efficiency value of 71.58%.

Conclusion: The intervention of *O. basilicum* on *S. platensis* improved the total phenol and phycocyanin content and there is potential for a pharmaceutical product.

Keywords

Spirulina platensis, *Ocimum basilicum*, synergistic, bioactive compounds

This article is included in the **Plant Science** gateway.

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Author roles: **Yuliani Y:** Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Resources, Software, Validation, Visualization, Writing – Original Draft Preparation; **Riyadi PH:** Conceptualization, Resources, Software, Supervision, Validation, Writing – Review & Editing; **Dewi EN:** Conceptualization, Project Administration, Resources, Supervision, Validation, Writing – Review & Editing; **Jaswir I:** Supervision, Validation, Writing – Review & Editing; **Agustini TW:** Conceptualization, Project Administration, Resources, Supervision, Validation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: The research was funded by the PMDSU (Program of Master Degree Leading to Doctoral Degree for Excellent Graduates) for the Ministry of Research and Technology of the Republic of Indonesia in 2019 with grant number 642-07/UM7.6.1/PP/2020.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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How to cite this article: Yuliani Y, Riyadi PH, Dewi EN *et al.* ***Ocimum basilicum* (kemangi) intervention on powder and microencapsulated *Spirulina platensis* and its bioactive molecules [version 1; peer review: 2 approved with reservations]** F1000Research 2021, 10:485 <https://doi.org/10.12688/f1000research.52394.1>

First published: 18 Jun 2021, 10:485 <https://doi.org/10.12688/f1000research.52394.1>

Introduction

Spirulina platensis is a blue-green microalgae that thrives in alkaline water and it has a high potential as a source of bioactive compounds with commercial importance^{1,2}. High value compounds with interesting functional properties such as phycobiliproteins consisting of phycocyanins and allophycocyanins, carotenoids, phenolic acids, omega-3 and omega-6 polyunsaturated fatty acids, phenol and flavonoid have been identified in *S. platensis*³⁻⁵. Phenolic compounds are a source of bioactive molecules with several beneficial health effects⁶ due to their ability to act as antioxidants⁷, antibacterial⁸, and antidiabetes agents⁹. Phycobiliproteins, carotenoids and phenol present in *S. platensis* have anti-inflammatory activities¹⁰, thus making them a potential functional food product¹¹.

Ocimum basilicum, commonly known as sweet basil or *kemangi* in Indonesia and called *rehan* in Arabic¹² is a popular culinary herb. *O. basilicum* is added to a variety of foods to impart a specific aroma. *O. basilicum* contains essential oils such as chavicol, linalool and eugenol, which are widely used in the food and pharmaceuticals industries¹³. The essential oils are able to reduce unpleasant odors and replace antioxidants^{14,15}. Besides essential oils, basil also contains phenol and flavonoid compounds which have antioxidant properties¹⁶⁻¹⁸.

Microencapsulation is a technique used to coat a material to protect the material from outside factors, as well as ease handling of the material. The most important factor in encapsulation is the type of coating used. The encapsulated material is referred to as the core, internal phase-, or filler, whereas the walls are sometimes called shells, layers, material wall, or membranes. A microcapsule can be coated by several coatings, but only one core compound can be coated^{19,20}.

To predict the potential for bioactivity, absorption, distribution, metabolism, and excretion of a substance, our research was performed with bioinformatics and *in silico* approaches. If we do not have special apps, certain internet-based or online resources can be used. DOCK Blaster for molecular docking prediction²¹, MDWeb and MDMoby for molecular dynamics analysis²², ADMET and DrugBank for drug database creation²³, as well as PreADME for ADMET tools²⁴, are some of the tools available online.

Various studies have reported the presence of bioactive compounds such as phenols and flavonoid in *S. platensis*²⁵ and *O. basilicum*²⁶. The present research aims to evaluate bioactive compounds of *O. basilicum* intervention on *S. platensis*. Firstly, total phenol, flavonoid, antioxidant activity and phycocyanin contents were evaluated. Secondly, the success of encapsulation of phenol, flavonoid, antioxidant activity and phycocyanin compounds was evaluated. The addition of these compounds was expected to reduce the amount of volatiles in *S. platensis*, which cause unpleasant odors. The third was predicting absorption, distribution, metabolism, and excretion (ADME) of phenols, azulene, flavonoids, and phycocyanin.

Methods

S. platensis powder was obtained from brackish water Aquaculture Fisheries (BBPBAP) Jepara (Central Java, Indonesia), *O. basilicum* was bought from a traditional market (Semarang, Central Java). The water used was multilevel distilled water, aquabidest Otsu-WI (PT. Otsuka Indonesia, Lawang, Indonesia). The reagents and chemicals used in this study were of analytical grade (CV. Chemix Pratama, Special Region of Yogyakarta, Indonesia), maltodextrin (CV. Multi Kimia Raya, Semarang, Indonesia) and gelatin (Xian, Biof Bio-Technology, Cina). This research was conducted in the food chemistry laboratory of Diponegoro University, Semarang, Central Java, Indonesia) from January 06, 2020 up to May 29, 2020.

Preparation of *Ocimum basilicum* leaf extract

The *O. basilicum* was extracted using distilled water (aquabidest) following modified methods reported by Handiani *et al.*²⁷. 2000 g of fresh *O. basilicum* leaves were added to 400 ml aquabidest and ground. The slurry was then filtered by using filter fabric and the extract result was approximately 1200 ml.

Intervention of *Ocimum basilicum* on *S. platensis*

A freeze-dried sample (DSB) and microencapsulation sample (MSB) of *S. platensis* were soaked with *O. basilicum* extract at a ratio of 1:4, w/v. A *S. platensis* sample with no *O. basilicum* added was used as a control (SP).

Preparation of freeze-dried *Ocimum basilicum*

O. basilicum and *S. platensis* were freeze-dried using a freeze dryer (Heto Powerdry LL 1500, Germany) at a temperature of -100°C for 48 hours. The *O. basilicum* extracts were applied to *S. platensis* (DSB) in the intervention study below.

Microencapsulation of freeze-dried *Ocimum basilicum*

Microencapsulation of *O. basilicum* was performed following the methods reported by Castro-Munoz *et al.* and Dewi *et al.*^{28,29} Maltodextrin and gelatin at a ratio of 9:1, w/w were used as a coating. Homogenization was then performed with a homogenizer (15A HG-wiseTis, Germany). Microencapsulated *O. basilicum* (MSB) was used in the intervention studies below.

Determination of total phenol content

Total phenol content was measured using modified Folin-Ciocalteu methods³⁰. Samples were sonicated for 30 minutes prior to measurement. Gallic acid was used as standard and was read at $\lambda=739$ nm using a spectrophotometer. In the test solution, 0.5 ml of Folin-Ciocalteu reagents and 1 ml of NaCO₃ were added to 1 ml of sample and the solution was mixed. Samples were incubated for 10 minutes at room temperature, then diluted with aquabidest to 10 ml. The measurement results were reported in milligram (mg) and were calculated as gallic acids equivalent (GAE) per gram of sample.

Determination of flavonoid content

Measurement of total flavonoid was performed using the slightly modified aluminium chloride method³¹. Modification was

through ultrasonic treatment before measurement, the sample was sonicated for 30 minutes and quercetin was used as a standard. In the test solution, 1.0 ml of sample was mixed with 0.3 ml of NaNO_2 (5%, w/v) and the solution was left to stand 5 minutes before 0.5 ml of AlCl_3 (2%, w/v) was added to the test solution. Samples were neutralized with 0.5 ml of 1 M NaOH solution and the samples were incubated for 10 minutes at room temperature. Absorbance was measured at $\lambda=310$ nm. The results are presented in milligrams (mg) and calculated as quercetin equivalent (QE) per gram of sample.

Phycocyanin content

40 mg of sample was added to 10 ml phosphate buffer (pH 7); the solution was sonicated for 30 minutes and stored at 4°C overnight. Samples were centrifuged to separate the blue supernatant. Next, samples were measured for absorbance at 620 nm according to the methods described by Setyoningrum & Nur³². Phycocyanin content was determined using Equation 1:

$$PC(\%) = \frac{\text{Abs} \times v}{3.39 \times w \times w_{\text{dry}}} \times 100\% \quad (1)$$

Where PC is phycocyanin content, Abs is absorbance at 620 nm; v is volume of solvent (ml); 3.39 is the coefficient of C-Phycocyanin at 620 nm; w is weight of sample (mg); and w_{dry} is percentage dry weight of sample.

Determination of antioxidant activity

The antioxidant activity of the sample was measured by 2,2'-azinobis-3-ethylbenzo-thiazoline-6-sulfonic acid (ABTS) radical according to the methods of Shalaby & Shanab³³. ABTS was formed by reacting 7 mM ABTS aqueous solution with 2.45 mM phosphate per sulphate in the dark for 4–16 hours at room temperature. Dilute ABTS solution with ethanol absorbance of 0.700 ± 0.05 at 734 nm was used for measurement. The photometric test was carried out with 0.9 mL ABTS solution and 0.1 mL of the tested sample mixed for 45 seconds, measurements were made immediately at 734 nm after 15 minutes. Antioxidant activity was expressed as the inhibition percentage of free radicals by the sample and was determined using Equation 2:

$$\text{Inhibition (\%)} = \frac{\text{Ab} - \text{As}}{\text{Ab}} \times 100 \quad (2)$$

Where Ab is the absorbance of the control reaction and As is the absorbance in the presence of the extract sample.

Determination of encapsulation efficiency

Encapsulation efficiency (EE) was determined following the methods described by Ong *et al.*³⁴. Encapsulation efficiency was calculated based on total coated active compounds and free active compounds. Percent encapsulation efficiency was determined using Equation 3:

$$EE(\%) = \frac{\text{Total coated active compounds} - \text{Free active compounds}}{\text{Total coated active compounds}} \times 100 \quad (3)$$

ADME analysis

The research was performed in two phases, namely: the first stage of accessing the PubChem server (<https://pubchem.ncbi.nlm.nih.gov/>) to obtain canonical SMILE information; the next step is to use swissADME (<http://www.swissadme.ch/>) to predict absorption, distribution, metabolism, and excretion³⁵. The BOILED Egg (Brain Or IntestinaL EstimateD permeation predictive model) methods are used for the determination of the absorption of the inhibitors in the brain and gastrointestinal tract. BOILED Egg provides a threshold ($\text{TPSA} \leq 131.6$ and $\text{WLOGP} \leq 5.88$) and the best representation of how far molecular structure is for well- or poorly absorbed³⁶. ADME is based on the Lipinski rule of five³⁷. The Lipinski rule of five is generally employed in accessing the drug-likeness of active compounds to prioritize compounds with an increased likelihood of high oral absorption³⁸.

Statistical analysis

Data obtained was reported as the mean of triplicates ($n=3$) $\pm$ standard deviation. Parametric data was analyzed using SPSS version 22.0 (IBM, Armonk, NY, USA)³⁹. Statistical analysis was preceded by a normality test with One Sample Kolmogorov-Smirnov Test and a homogeneity test with the Levenes Test at significance level ($P > 0.05$). Parametric tests were carried out with One Way ANOVA at significance level ($P < 0.05$), followed by post hoc Tukey HSD.

Results

Total phenol, flavonoid and antioxidant activity were measured in *S. platensis* with no treatment (SP), *S. platensis* treated with freeze-dried *O. basilicum* (DSB), *S. platensis* treated with microencapsulation freeze-dried *O. basilicum* (MSB) and *O. basilicum* leaf extract (B), plot of standard calibration of gallic acid (left) quercetin (right) (Figure 1). Phycocyanin content was measured in SP, DSB and MSB, and then encapsulation efficiency was measured on total phenol, flavonoid, antioxidant activity and phycocyanin. The results of the One Sample Kolmogorov Smirnov Test and Levene's Test were more than ($P > 0.05$) thus, the data were normally or homogeneously distributed. Therefore the data analysed by the One Way ANOVA were less than ($P < 0.05$) and are shown in Table 1, followed by the post hoc Tukey HSD (Table 2–Table 6). The DSB sample can increase the total phenol 49.50% and antioxidant activity 12.66% of *S. platensis*. However, total flavonoid is not significantly different with *O. basilicum* intervention on *S. platensis* (Figure 2).

O. basilicum intervention can increase the levels of phycocyanin in *S. platensis* 40.72% shown in (Figure 3). *O. basilicum* intervention on *S. platensis* when extracted will make a blue ring on the surface, it is caused by compounds contained in *O. basilicum* called azulene.

This is encapsulation is less effective in microencapsulation of polyphenol compounds such as phenol and flavonoid

Figure 1. Plot of standard calibration of gallic acid (left) and quercetin (right).

Table 1. Normality test, homogeneity and ANOVA result of total phenol, flavonoid, phycocyanin and antioxidant activity *statistically significant ($p < 0.05$).

Asymptotic significance	Total Phenol	Flavonoid	Antioxidant Activity	Phycocyanin	Encapsulation Efficiency
Normality	0.062	0.061	0.130	0.200	0.200
Homogeneity	0.067	0.062	0.380	0.117	0.118
ANOVA	0.000	0.000	0.000	0.000	0.000

Table 2. Descriptive statistics and result of the Tukey HSD tests of total phenol.

SAMPLE	N	Subset for alpha = 0.05			Superscripts
		1	2	3	
MSB	3	19,6707			a
SP	3	24,0413			a
DSB	3		35,9413		b
B	3			117,2467	c
Significant.		,654	1.000	1.000	

SP = *S. platensis* with no treatment, DSB = *S. platensis* treated with freeze-dried *O. basilicum*, MSB = *S. platensis* treated with microencapsulation freeze-dried *O. basilicum*, B = *O. basilicum* leaf extract.

(Figure 4). Raw absorbance data for bioactive compounds assays are available as *underlying data*⁴⁰.

Discussion

Algae are a valuable source of proteins and phenol compounds⁴¹. *S. platensis* is a type of algae with a high total phenol content. Extraction methods and the solvent used are responsible for the type and yield of phenolic compounds from algae sources⁴². In *S. platensis*, distilled water has been reported as the best solvent for extraction of phenolic compounds with

total phenol content of 43.2 ± 1 GEA/g⁴³. *S. platensis* powder prepared via oven drying is reported to have a broad range phenolic profile that includes gallic acid, catechin, caffeic acid, P-hydroxybenzoic acid, P-cumaric acid, ferulic acid, quercetin, genistein and kaempferol⁴⁴. Variation in total phenol content between algae species is reportedly due to algal type, origin and growth condition of different microalgae⁴⁵.

Fresh *O. basilicum* leaf extract has been reported to have lower total phenol content than that which has been freeze-dried⁴⁶.

Table 3. Descriptive statistics and result of the Tukey HSD tests of flavonoid.

SAMPLE	N	Subset for alpha = 0.05			Superscripts
		1	2	3	
MSB	3	1,8370			a
DSB	3		6,4733		b
SP	3		6,6000		b
B	3			7,0433	c
Significant.		1.000	.600	1.000	

SP = *S. platensis* with no treatment, DSB = *S. platensis* treated with freeze-dried *O. basilicum*, MSB = *S. platensis* treated with microencapsulation freeze-dried *O. basilicum*, B = *O. basilicum* leaf extract.

Table 4. Descriptive statistics and result of the Tukey HSD tests of antioxidant activity.

SAMPLE	N	Subset for alpha = 0.05			Superscripts
		1	2	3	
MSB	3	69.2777			a
SP	3		87.1610		b
B	3			94.9330	c
DSB	3			98.1997	c
Significant.		1.000	1.000	.109	

SP = *S. platensis* with no treatment, DSB = *S. platensis* treated with freeze-dried *O. basilicum*, MSB = *S. platensis* treated with microencapsulation freeze-dried *O. basilicum*, B = *O. basilicum* leaf extract.

Table 5. Descriptive statistics and result of the Tukey HSD tests of phycocyanin.

SAMPLE	N	Subset for alpha = 0.05			Superscripts
		1	2	3	
MSB	3	1.7733			a
SP	3		3.0167		b
DSB	3			4.2567	c
Significant.		1.000	1.000	1.000	

SP = *S. platensis* with no treatment, DSB = *S. platensis* treated with freeze-dried *O. basilicum*, MSB = *S. platensis* treated with microencapsulation freeze-dried *O. basilicum*, B = *O. basilicum* leaf extract.

Previous studies have reported that dried *O. basilicum* leaf extracted with methanol gave high total phenol values⁴⁷. The phenol compounds present in *O. basilicum* include rosmarinic, caftaric, caffeic, chicoric, p-hydroxybenzoic, p-coumaric, and protocatechuic acids. The phenol compounds in *O. basilicum* play an important role in its antioxidant activity⁴⁸. *S. platensis*

microcapsule with the intervention of *O. basilicum* (MSB) gives low total phenol. Microencapsulation using maltodextrin and gelatin can protect polyphenol compounds⁴⁹.

O. basilicum intervention on *S. platensis* significantly increases bioactive compounds (Figure 3), except for total flavonoid.

Table 6. Descriptive statistics and result of the Tukey HSD tests of encapsulation efficiency.

SAMPLE	N	Subset for alpha = 0.05				Superscripts
		1	2	3	4	
FLAVONOID	3	30,7700				a
PHENOL	3		58,3467			b
PHYCOCYANIN	3			71,5833		c
ANTIOXIDANT	3				80,5933	d
Significant.		1.000	1.000	1.000	1.000

Figure 2. Total phenol, flavonoid and antioxidant activity (n=3, mean value ± standard deviation; different superscripts indicate a significant difference). SP = *S. platensis* with no treatment, DSB = *S. platensis* treated with freeze-dried *O. basilicum*, MSB = *S. platensis* treated with microencapsulation freeze-dried *O. basilicum*, B = *O. basilicum* leaf extract.

The total flavonoid content of the *S. platensis* treated with freeze-dried *O. basilicum* (DSB) was not significantly different from the control sample (SP). Previous studies have reported that total flavonoid in *S. platensis* is less than the total phenols, phenolics (1.73%) and flavonoids (0.87%)⁵⁰. Another study reported that the powder of *S. platensis*, which was dried in an oven at a temperature of $\pm 50^{\circ}\text{C}$, did not effect the phenolic compound quercetin, where the compound was one of the active substances of the flavonoid class⁴⁴. The flavonoids are considered as indispensable in a variety of medicines, nutraceutical, pharmaceutical and cosmetic applications⁵¹. Flavonoids play an anti-inflammatory and anti-viral role⁵². The optimum for the extraction process are dry conditions compared to wet conditions. Extraction using ethanol had a higher total flavonoid content⁵³. The total flavonoid content of the *S. platensis* microencapsulated and freeze-dried tended to be

low. Microencapsulation can maintain the stability of flavonoid from processing effects that cause degradation^{54,55}.

Spirulina platensis could be considered as a valuable source of bioactive colored and phenolic compounds with potent antioxidant activity²⁵. The ABTS method was chosen because it has a high level of sensitivity (99.44%) compared to the 1,1-Diphenyl-2-Picrylhydrazyl (DPPH) method (95.3%)³³. Total phenol and flavonoid content showed positive correlation to the antioxidant activity of *S. platensis*. Phenolic components play an important role in the antioxidant activity⁵⁶. Phenolic compounds are good electron donors because the hydroxyl groups can contribute to antioxidant activity⁵⁷. The tocopherol and phycocyanin in microalgae have potential as antioxidants in food, so that it acts as a functional food⁵⁸. The *S. platensis* treated with freeze-dried *O. basilicum* (DSB) showed an

Figure 3. Phycocyanin content (n=3, mean value $\pm$ standard deviation; different superscripts indicate a significant difference). SP = *S. platensis* with no treatment, DSB = *S. platensis* treated with freeze-dried *O. basilicum*, MSB = *S. platensis* treated with microencapsulation freeze-dried *O. basilicum*.

Figure 4. Encapsulation efficiency. (n=3, mean value $\pm$ standard deviation; different superscripts indicate a significant difference).

increase in antioxidant activity compared to *S. platensis* with no treatment (SP). Previous research explained that *O. basilicum* contains essential oils which also have potential as antioxidants⁵⁹. According to Asghari *et al.*⁶⁰, the mixture of carotenoid pigments, chlorophyll and blue pigments such as phycocyanin of *S. platensis* produce strong antioxidants.

Ocimum basilicum contains 65 active compounds, and the compounds with the highest content are namely 31.6% linalool and 23.8% methylchavicol. Essential oils in *O. basilicum* have the potential as antioxidants¹³. The essential oil of linalool significantly prevents the formation of UVB-mediated 8-deoxy guanosine, which causes oxidative damage to DNA. This is because it has the ability to prevent reactive oxygen species (ROS) and restore the balance of oxidative cells⁶¹. This research indicates that there is a synergistic interaction between phycocyanin and total phenol in antioxidant activities. *S. platensis* treated with microencapsulation freeze-dried *O. basilicum* (MSB) impart smaller values on total phenol, flavonoid, phycocyanin and antioxidant activity. This is in correlation with previous research which showed that the *S. platensis* microcapsule has antioxidant activity of 49.05%⁶². Essential oils that play a role as an antioxidant can last for six months with a slight decrease in antioxidant activity and phenol content after microencapsulation⁶³. Treated microencapsulation can control antioxidant capacity and is a promising strategy in extending shelf life³⁵.

S. platensis cultivated with brackish water had a higher phycocyanin content (Figure 4), whereas *S. platensis* cultivated in freshwater only had a 1.74% phycocyanin content⁶⁴. *S. platensis* cultivated with seawater has a maximum phycocyanin content⁶⁵. Phycocyanin is a natural blue pigment that functions as an antioxidant, anti-inflammatory and anti-carcinogenic^{66,67}. The *S. platensis* treated with freeze-dried *O. basilicum* (DSB) impart higher levels of phycocyanin, where a combination of *S. platensis* and *O. basilicum* with a ratio of 1:5 detects the presence of azulene using gas chromatography-mass spectrometry (GC-MS)²⁷. Azulene is an aromatic compound from essential oils in *O. basilicum*⁶⁸, and it is a blue hydrocarbon compound that has a strong dipole moment^{27,69}. Azulene has a small gap between the highest energy molecular orbitals (HOMO) with the lowest energy molecular orbitals that do not have electrons (LUMO)⁷⁰. Therefore, the presence of azulene in *S. platensis* treated freeze-dried *O. basilicum* can increase phycocyanin levels.

Encapsulation efficiency is used to evaluate the success of a microencapsulation technique. Encapsulation using a combination of polyanion and polycation coatings such as maltodextrin and gelatin has a higher yield. This is due to the stability of the emulsion between maltodextrin and gelatin⁷¹. The amount of bioactive content on the surface will reduce the value of encapsulation efficiency. This will cause the amount of bioactive compounds that are wrapped to increasingly shrink because many are attached to the surface. So that this event

will damage the oxidative stability of microcapsules⁷². The encapsulation efficiency of phycocyanin was in accordance with the results of previous studies⁷³, which is encapsulation using an alginate coating has an encapsulation efficiency value of 71.75%. The value of encapsulation efficiency in total phenols and flavonoids in *S. platensis* is effected by using liposomes or nanoliposomes in encapsulation of bioactive compounds, this is because liposome is stable at low pH and is able to withstand the time of release in the stomach, but it is less consistent in the intestine^{74,75}. The encapsulation efficiency of antioxidant has been shown in previous research where antioxidant microencapsulation using the freeze drying method has an encapsulation efficiency value ranging from 73–86%⁷⁶.

Intestinal absorption and brain permeation set crucial parameters at their target site of action for any medication for its pharmacokinetics and bioavailability. Consequently, the BOILED-Egg study was used, as previously stated, to predict gastrointestinal (GI) absorption and brain access for phenol, azulene, flavonoid, and phycocyanin. The white region is the physicochemical space of the molecules most likely to be consumed by the gastrointestinal tract, whereas the yellow region (yolk) is the physicochemical space of the molecules most likely to reach the brain. The white and yolk regions are not mutually exclusive³⁶. Phenol, azulene, and phycocyanin were found to be among the well-absorbed molecules based on the study (Table 7).

Tables 8 and Table 9 demonstrate that phenol, azulene, and phycocyanin comply with Lipinski or drug-likeness laws. Drug-likeness is a term used to explain how *in vivo* molecular properties are influenced by compounds' physicochemical properties. This research indicates that the substance will spread well to all parts of the body to play an active role as a drug⁷⁷. The physicochemical properties obtained from molecular structures are used by most drug-likeness testing laws and compare such properties with the medicines that have been reported. The Lipinski rule is one of the most used rules⁷⁸. The rule of five was developed to set drugability guidelines for new molecular entities (NMEs)⁷⁹. Therefore, the rule suggests that molecules, whose properties fall outside of these boundaries, are unlikely to become orally bioavailable drugs⁸⁰. As drug candidates, phenol, azulene, and phycocyanin have excellent potential. This calculation is based on a molecular weight (MW) value of less than 500 g mol⁻¹, an acceptor of hydrogen bonds of less than 10, a donor of hydrogen bonds of less than five, a surface area of topology (TPSA) of less than 140 Å, and a LogP of less than five.

Conclusion

Ocimum basilicum intervention significantly increased total phenol, phycocyanin and antioxidant activity in *S. platensis*. However, total flavonoid content did not differ significantly in untreated *S. platensis* controls compared to treated. Bioactive compounds after microencapsulation showed the lowest values. Microencapsulation of phycocyanin with maltodextrin and

Table 7. Pharmacokinetics parameters of the analysis compound predicted by Swiss ADME.

Compound	GI absorption	BBB permanent	Bioavailability radar	Boiled egg
Phenol	High	Yes
Azulene	Low	Yes
Flavonoid	Low	No
Phycocyanin	High	No

Note: GI: gastrointestinal; BBB: blood-brain barrier; LIPO: lipophilicity; FLEX: flexibility; INSATU: unsaturation; INSOLU: insolubility; POLAR: polarity; SIZE: molecular weight.

Table 8. Physicochemical parameters of the analysis compound predicted by Swiss ADME.

Compound	MW (g.mol ⁻¹)	HA	AHA	RB	HBA	HBD	MR	TPSA	L
Phenol	94.11	7	6	0	1	1	28.46	20.23	1.24
Azulene	128.17	10	10	0	0	0	43.06	0.00	2.07
Flavonoid	594.52	42	16	6	15	10	138.73	260.20	2.23
Phycocyanin	526.71	39	5	8	3	3	174.47	86.35	4.24

Note: MW: molecular weight; HA: heavy atoms; AHA: aromatic heavy atoms; RB: rotatable bonds; HBA: hydrogen bond acceptor; HBD: hydrogen bond donor; MR: molar refractivity; TPSA: topology polar surface area (Å²); L: lipophilicity

Table 9. Druglikeness property using Lipinski Rule of Five.

Compound	Molecular mass less than 500 Dalton	High lipophilicity (expressed as LogP less than 5)	Less than 5 hydrogen bond donors	Less than 10 hydrogen bond acceptors	Molar refractivity should be between 40–130	Conclusion
Phenol	Yes	Yes	Yes	Yes	No	Yes
Azulene	Yes	Yes	Yes	Yes	Yes	Yes
Flavonoid	No	Yes	No	No	No	No
Phycocyanin	No	No	Yes	Yes	No	Yes

gelatin showed high encapsulation efficiency values. Hence, *S. platensis* treated freeze-dried *O. basilicum* has potential as a pharmaceutical product.

Data availability

Underlying data

Figshare: Underlying data for ‘*Ocimum basilicum* (kemangi) intervention on powder and microencapsulated *Spirulina platensis* and its bioactive molecules’, <https://doi.org/10.6084/m9.figshare.14291069>⁴⁰

This project contains the following underlying data:

- Data file 1. Flavonoid content from the intervention of *O. basilicum* on *S. platensis* with microencapsulation.
- Data file 2. Antioxidant activity from the intervention of *O. basilicum* on *S. platensis* with microencapsulation.
- Data file 3. Encapsulation efficiency of phenol, flavonoid, antioxidant and phycocyanin content from the intervention of *O. basilicum* on *S. platensis* with microencapsulation.
- Data file 4. Phycocyanin content from the intervention of *O. basilicum* on *S. platensis* with microencapsulation.
- Data file 5. Statistical analysis by SPSS v.22 on bioactive compounds.
- Data file 6. Total phenol content from the intervention of *O. basilicum* on *S. platensis* with microencapsulation.

Data are available under the terms of the [Creative Commons Attribution 4.0 International license](#) (CC BY 4.0).

Author contributions

Yuliani: Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Resources, Software, Validation, Writing-Original Draft Preparation, Visualization,

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Acknowledgements

The authors wish to thank Dr. Diana Nur Afifah, S.TP., M.Si, Department of Nutrition Research, Universitas of Diponegoro, Semarang, Central Java for his guidance with this study.

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Version 1

Reviewer Report 21 July 2021

<https://doi.org/10.5256/f1000research.55668.r87948>

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The manuscript by Yuliani *et al.* presents the effect of the intervention of *Ocimum basilicum* on freeze-dried and microencapsulated *Spirulina platensis*. In general, I find this manuscript provides an important contribution in developing palatable and higher bioactive compounds in *S. platensis*. However, I have outlined below a few issues that need to be addressed before publication, and some more minor comments and suggestions to further improve the quality of the manuscript.

Major:

1. The methods section is not clear. It is not clear whether *O. basilicum* alone was freeze-dried and/or microencapsulated, or the *S. platensis* alone, or is it after the intervention of *O. basilicum* to *S. platensis*? Example: In the sub-methods title "Intervention of *Ocimum basilicum* on *S. platensis*", the authors stated "...freeze-dried sample (DSB) and microencapsulation sample (MSB) of *S. platensis*...", while in the sub-methods title "Preparation of freeze-dried *Ocimum basilicum*", the authors described that both *O. basilicum* and *S. platensis* were freeze-dried using a freeze dryer, and followed by a description of the "Microencapsulation of freeze-dried *Ocimum basilicum*". Please clarify and make changes to the methods section as appropriate.
2. In the abstract and introduction, the authors stated that one of the aims of the studies is to reduce unpleasant odours of *S. platensis* by intervention with *O. basilicum*. However, the organoleptic properties were not presented and discussed.
3. The authors tried to predict the ADME of the bioactive compounds in the product (*Ocimum basilicum* - *Spirulina platensis*) by using the SwissADME. However, as this approach can only predict pharmacokinetics, drug-likeness of small molecules, the authors use four main compounds found in the product namely phenol, azulene, flavonoid (not clear on which flavonoids), and phycocyanin (the structure of each compound used for analysis need to be added to Table 7). While this information provides a prediction of an individual molecule of

the complex bioactive compounds in *Ocimum basilicum* – *Spirulina platensis*, it does not provide any additional insight to the manuscript as matrix-derived combination influences ADME. *In vivo* study is needed to determine the bioavailability of *Ocimum basilicum* intervention on freeze-dried and microencapsulated *Spirulina platensis*. If the authors wish to include this data in the current study, additional discussion in limitations and future studies include the need for *in vivo* study are needed.

4. In the method section, it is not clear how the encapsulation efficiency was determined. Do the authors analyze the active compounds in both the free and encapsulated fractions? If yes, please describe the procedure and parameter of separation of the free active compound from the encapsulation.
5. In the discussion section, it is not clear how this research indicates a synergistic interaction between phycocyanin and total phenols in antioxidant activities (page 9 of 16). No data support this statement.
6. Figure 4 is data on encapsulation efficiency. In the discussion (Page 9 of 16), by using the information in Figure 4, the authors stated the effect of different cultivation methods of *S. platensis* on phycocyanin content. No data support this statement.

Minor:

1. In the result section paragraph 2 (page 4 of 16), please add a description of the result of MSB.
2. In the method section, microencapsulation, please describe what is the total solid content of coating materials used in the study.
3. In the discussion section paragraph 3 (page 6 of 16), please be clear on what bioactive compounds are the authors referring to and for which treatment.
4. In the discussion section paragraph 4 (page 7 of 16), please describe "...bioactive colored ..."
5. In the introduction and abstract, *O. basilicum* intervention to *S. platensis* is to increase consumption and increase bioactive compounds and their potential for a functional food product. The conclusion state as a potential for a pharmaceutical product. Please be consistent.
6. In Figure 2, B = *O. basilicum* leaf extract was analyzed. Please describe B in the method section.

The paper would benefit from a more detailed discussion regarding the organoleptic properties of how the *Ocimum basilicum* intervention reduces unpleasant odours as outlined in the comments above.

An additional suggestion that will improve the presentation of the manuscript is listed below:

1. The result section should focus on the main finding and not the statistical analysis. Figure 1 and Table 1-6 can be added to the supplemental section.
2. The authors need to check the accuracy of the statement in the result section's first sentence (page 4 of 16). Figure 1 is a standard calibration curve use to calculate total phenol of samples as gallic acid equivalent (GAE) and total flavonoid of samples as quercetin

equivalent (QE) but not for antioxidant activity. Additionally, this information is more suitable to be in the method section and not the result section. Please also refer to my additional suggestion point 1.

3. Citation in the discussion should always be referred to the result of the study, either it is in accordance or opposition with the current finding. In the discussion section page 7, the authors cite that microencapsulation can maintain the stability of flavonoids from processing effects that cause degradation. Please elaborate more on how this statement is in the context of the current finding.

Is the work clearly and accurately presented and does it cite the current literature?

Partly

Is the study design appropriate and is the work technically sound?

Partly

Are sufficient details of methods and analysis provided to allow replication by others?

Partly

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Bioactive compounds, lipid metabolism, carotenoids/retinoids homeostasis, pharmacology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 26 Oct 2021

Yuliani Yuliani

We would like to thank the reviewer for the time and willingness to assess the quality of our manuscript. In the new version our manuscript, suggested changes in the all section has been added according to the reviewer's recommendation, except for the SwissADME analysis which will be carried out in next studies.

This research, SwissADME is used to predict chemical structure homology. SwissADME is used as the initial stage for further research with experimental animals. The proof will be

carried out in future research and publications on in vivo.

Competing Interests: No competing interests were disclosed

Reviewer Report 12 July 2021

<https://doi.org/10.5256/f1000research.55668.r87945>

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Joko Santoso

Department of Aquatic Products Technology, Faculty of Fisheries and Marine Sciences, IPB University, Bogor, Indonesia

This work presents to enrich *Spirulina platensis* with *Ocimum basilicum* by soaking method, and evaluated their characteristic after being freeze-dried and Microencapsulation of freeze-dried.

The manuscript is recommended to be revised as follows:

- Authors stated the intervention of *O. basilicum* on *S. platensis* (DSB) significantly increases total phenol by 48.7% (in Page 1); Page 4 (Results) states The DSB sample can increase the total phenol 49.50% (I calculated it, that 49.5% is true value); antioxidant activity 12.66% (page 4 results), I calculated 12.67% (Please check it).
- Fig 1 shows the calibration, this is the method of how to get the value of total phenol and flavonoid content. Fig 1 should be deleted.
- The results, Tukey HSD (Table 2 2-6) (Page 4, 5, 6, 7) statistic analysis, it is not necessary to written in the manuscript, please delete.
- Fig 2. In the analysis data process (statistic analysis), B (*Ocimum basilicum*) does not include the factors studied in accordance with the research objectives, B is excluded from the data analysis process (statistics) (See Fig 3, the ANOVA like this). The value of phenol, flavonoid and antioxidant activity of B can be written in the paragraph). Please re-analyse the statistics (ANOVA).
- (5) Fig 2, please separate into three figures and complete the description of the y-axis (the unit of y-axis such as %, mgQE/g).

Minor revision:

- "*Spirulina platensis* is a blue-green microalgae" (Page 2) - it should be microalga (single).
- "A freeze-dried sample (DSB) and microencapsulation sample (MSB) of *S. platensis* were soaked (page 4)" - add time for soaking (how long).

- "Gallic acid was used as standard and was read at $\lambda=739$ nm using a spectrophotometer" - add the type of spectrophotometer.
- "40 mg of sample was added" - sample of 40 g.
- Make all figures clear with same font, x-axis and y-axis description should be added in the each figure.
- "Algae are a valuable source of proteins and phenol compounds" - A algae means macroalgae, be careful with this statement.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Partly

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Partly

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Processing of aquatic organism

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 26 Oct 2021

Yuliani Yuliani

We would like to thank the reviewer for the time and willingness to assess the quality of our manuscript. In the new version our manuscript, suggested changes in the all section has been added according to the reviewer's recommendation.

Competing Interests: No competing interests were disclosed

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