Check for updates

RESEARCH ARTICLE

REVISED Evaluation of immunologic response of salivary sIg-A in pediatric tuberculosis patients to antigen Ag38-rec of *Mycobacterium tuberculosis* Indonesian strain [version 5; peer review: 2 not approved]

Tri Yudani Mardining Raras¹, Diah Erma Pritta Santi², HMS Chandra Kusuma³

¹Department of Biochemistry and Molecular Biology, Faculty of Medicine, Brawijaya University, Malang, Indonesia

²Master Program in Biomedical Science, Faculty of Medicine, Brawijaya University, Malang, Indonesia

³Department of Pediatrics, Dr. Saiful Anwar Public Hospital, Malang, Indonesia

v5 First published: 16 Nov 2015, 4:1267
<https://doi.org/10.12688/f1000research.7234.1>

Second version: 01 Feb 2016, 4:1267
<https://doi.org/10.12688/f1000research.7234.2>

Third version: 18 Feb 2016, 4:1267
<https://doi.org/10.12688/f1000research.7234.3>

Fourth version: 30 Jun 2016, 4:1267
<https://doi.org/10.12688/f1000research.7234.4>

Latest published: 24 Oct 2016, 4:1267
<https://doi.org/10.12688/f1000research.7234.5>

Abstract

Objective: We studied the immune response of salivary secretory immunoglobulin A (sIg-A) from a pediatric tuberculosis (TB) group (scoring ≥ 6) and non-TB group (scoring < 6) against recombinant Ag38 (Ag38-rec) from *Mycobacterium tuberculosis* Indonesian strain.

Materials and Methods: Seventy-eight children were divided into three groups; those with TB (n=26), those with suspected TB (n=26), and healthy children (n=26), their saliva was collected, and salivary sIg-A was challenged with purified Ag38-rec using the dot blot method. A change of color from white to dark blue indicated a positive reaction.

Results: The immune response of sIg-A of children with TB and those with suspected TB to Ag38-rec was not significantly different. In the TB group, Ag38-rec showed a higher sensitivity than protein purified derivative (PPD) (70.8% vs. 62.5%), but a lower specificity (26.9% vs. 34.62%). However, within both groups (scoring ≥ 6) as well as non-TB group (scoring < 6) Ag38-rec was able to identify children with a positive TST (tuberculin skin test) better than PPD.

Conclusion: The antigen Ag38-rec could not distinguish between children with TB (scores ≥ 6 and < 6). However, it demonstrated the potential of Ag38-rec for use in screening for TB infection among children with suspect TB (scores < 6).

Open Peer Review

Approval Status **XX**

	1	2
version 5		
(revision)		
24 Oct 2016		
version 4		
(revision)	X	
30 Jun 2016	view	
version 3		
(revision)	X	X
18 Feb 2016	view	view
version 2		
(revision)	X	
01 Feb 2016	view	
version 1		
16 Nov 2015	X	
	view	

1. **Dorothy Yuan**, UT Southwestern Medical Center at Dallas, Dallas, USA

2. **Philippa M Musoke**, The Makerere University - Johns Hopkins University Research Collaboration, Kampala, Uganda

Keywords

salivary sIg-A , children , tuberculosis , antigen Ag38-rec

This article is included in the **World TB Day** collection.

Willy Ssengooba , Makerere University,
Kampala, Uganda

Any reports and responses or comments on the article can be found at the end of the article.

Corresponding author: Tri Yudani Mardining Raras (daniraras@yahoo.com)

Competing interests: No competing interests were disclosed.

Grant information: The author(s) declared that no grants were involved in supporting this work.

Copyright: © 2016 Raras TYM *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. Data associated with the article are available under the terms of the [Creative Commons Zero "No rights reserved" data waiver](#) (CC0 1.0 Public domain dedication).

How to cite this article: Raras TYM, Pritta Santi DE and Kusuma HC. **Evaluation of immunologic response of salivary sIg-A in pediatric tuberculosis patients to antigen Ag38-rec of *Mycobacterium tuberculosis* Indonesian strain [version 5; peer review: 2 not approved]** F1000Research 2016, 4:1267 <https://doi.org/10.12688/f1000research.7234.5>

First published: 16 Nov 2015, 4:1267 <https://doi.org/10.12688/f1000research.7234.1>

REVISED Amendments from Version 4

In this version, a small explanation concerning Figure 1 was added, as well as Figure 1B. A conclusion has also been added.

See referee reports

Introduction

Tuberculosis (TB) among children has been a serious threat in Indonesia, primarily due to a lack of accurate methods to diagnose the disease. Clinical examination, the acid-fast bacillus (AFB) test, and culture are the gold standard methods for the diagnosis of TB in adults. However, these methods have limited use for diagnosing TB in children in a clinical setting because collecting sputum samples from children is difficult. Moreover, children with TB do not exhibit the typical symptoms as TB observed in adults. The diagnosis of pediatric TB in Indonesia is determined according to the TB Scoring System¹. This system is based on the clinical signs and symptoms found in suspected TB in children¹. Children with a total score of ≥ 6 are considered to have TB and are treated for the disease. However, the accuracy of this scoring method is constrained due to the non-specificity of TB symptoms in children, and may result in over or under-diagnosis². In consequence, a rapid, simple, and inexpensive method to confirm *Mycobacterium tuberculosis* (Mtb) infection is required to reduce the spread of the disease. One such promising method, a serodiagnostic approach focusing on the detection of specific TB antibodies in patients, has been extensively studied³⁻⁵. A non-invasive diagnostic method like this would be highly valued, particularly with regards to pediatric TB patients.

Secretory immunoglobulin A (sIg-A) antibodies and presentations by antigen presenting cells (APC) are the first line of defense against bacterial invasion. These antibodies are a major class of immunoglobulin in external secretions and provide specific immunological protection in all mucosal surfaces that prevent the entry of bacteria⁶. Saliva contains a significant amount (85%) of sIg-A. This antibody is produced by B lymphocytes found near the salivary glands⁷. The 38-kDa protein from Mtb contains B-cell epitopes and has been shown to have high specificity for the *Mycobacterium* complex⁸. Previous study by Raras *et al.* demonstrated an immune response of salivary sIgA against recombinant Ag38 (Ag38-rec) Indonesian strain in adult pulmonary TB patients with a sensitivity of up to 80%, but with a low specificity (38%)⁹. Another study investigating Mtb specific antibodies, including sIgA, in saliva from children of the Warao Amerindian tribes in Venezuela, reported significantly greater reactivity to the purified protein derivative (PPD) antigen with a sensitivity of 26.5% and specificity of 97%¹⁰. In this study, the saliva from children diagnosed with TB (those scoring ≥ 6 in the TB Scoring System) and from children with suspected TB (those scoring < 6) was tested against Ag38-rec produced in our laboratory. The term 'Indonesian strain' is used throughout this article to refer to *M. tuberculosis* that was isolated from an Indonesian patient who was suffering from severe pulmonary TB, from which the *pab* gene was amplified. We evaluated whether Ag38-rec from the Mtb Indonesian strain could be used to differentiate between

children with pulmonary TB with scores of ≥ 6 and children with suspected TB with scores of < 6 .

Methods

Purification of Ag38-rec

Production of Ag38-rec was conducted according to a previous study with slight modifications¹¹. The *pab* gene coding for Ag38 was amplified via a PCR method using chromosomal DNA from *M. tuberculosis* which was isolated from a severe pulmonary TB patient in Malang, Indonesia. The fragment was inserted into a plasmid producing pMBhis. We do not know the strain of this Mtb, but alignment of the nucleotide sequence of the *pab* gene showed 95% homology to the *pab* gene from *M. tuberculosis* H37Rv. *Escherichia coli* BL21-(DE3) containing plasmid pMBhis was grown in Luria broth medium until OD₆₀₀ of 0.5 and induced with IPTG. After 3 h the cells were then harvested by centrifugation. The pellet was mixed with phosphate buffer containing protease inhibitor phenyl methyl sulphonyl fluoride (Sigma, USA) and was broken using a sonicator. Protein purification was performed using a Protino®Ni-TED column according to the manufacturer's protocol (Protino, Dueren, Germany). Following centrifugation for 15 min at 10,000 $\times$ g the supernatant was loaded onto an Ni-TED 1000 column and washed twice with phosphate buffer containing 10 mM imidazole. The antigen Ag38-rec and other proteins that bound to the nickel column were eluted using phosphate buffer containing 250 mM imidazole. The eluted proteins were collected and loaded once more on the new Ni-TED column. The eluent was then dialyzed against the elution buffer in order to remove imidazole. The purity of Ag38-rec was analyzed using 12.5% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE).

Clinical specimens

A total of 78 children (n=78) were included in this study: 26 patients with recently diagnosed pulmonary TB (total scores of ≥ 6), 26 patients with suspected TB (scores of < 6) from primary health care clinics in Malang City, Indonesia, and 26 healthy children to serve as negative controls. All of the participants were between the ages of 6 and 15 years, and their parents signed informed consent forms. The inclusion criteria for the healthy controls included absence of illness in the three weeks prior to the study and a lack of contact with adult TB patients. The study was initiated after being approved by the Ethical Commission from the Faculty of Medicine, Universitas Brawijaya, Malang Indonesia (No.341/EC/KEPK-OPDS/05/2014). Saliva samples were collected from June to October 2014. The diagnosis of TB was conducted by a pediatrician based on the eight parameters of the scoring system¹ i.e., contact with TB patients; fever; cough; enlarged lymph nodes of the neck, groin, and armpits; nutritional status; bone and joint swelling; chest X-ray (CXR); and Mantoux tuberculin skin test (TST).

Serological test

The immunodot blot method was applied according to a previous experiment⁹. Before mounting onto the dot blot apparatus, a nitrocellulose membrane was pre-wetted using sterile H₂O. A 20 μ L volume containing 1 g of antigen Ag38-rec in Tris-Cl buffer [pH 7.4] was dropped onto the membrane following overnight

incubation at 4°C with blocking buffer. PPD (protein purified derivative) from Mtb (Serum Staten Institute, Denmark) served as a positive control. The next day, the blocking buffer was removed and replaced with TBS and gently shaken for 10 min at 4°C. After the blocking agent was removed, 50 µL of primary antibody was applied to the membrane and incubated for 12 h at room temperature with gentle shaking. The solution was then discharged and the membrane was washed three times with 0.05% TBS-Tween-20 and subsequently shaken in Tris-Cl buffer containing secondary antibody (1:500) (Anti-human IgA (alpha-chain specific) alkaline phosphatase conjugate, SIGMA, USA.Product.no. A.9669) at room temperature for 1 h, followed by three washing steps. Finally, a chromogenic substrate nitro-blue tetrazolium chloride and (5-bromo-4-chloro-3'-indolylphosphate p-toluidine salt (NBT-BCIP) (KPL.USA.Product.no.50-81-18)) was applied to the membrane in the darkroom at room temperature for 30 min. The reaction was stopped by the addition of H₂O. In order to optimize which concentration of antigen that produced the strongest color, we performed checker board using different concentration of antigen (500 ng, 250 ng and 125 ng) against salivary sIgA from TB and non TB patients. The Corel Draw graphic suite X4 (Corel, USA) program was used to interpret the color range of the blot(s). The gradation of color was quantified to numerical value using Corel Photopaint 11. Positive and negative value was based on the cut-off of the median value of the positive control (PPD) after reaction with saliva of the TB group. A dot was regarded as positive when the value was below the cut-off point and as negative when the value was higher than the cut-off point. A positive result was defined as a dark blue or dark purple spot (>50%) on the blot.

Saliva collection

The procedure of saliva collection was conducted according to Chiappin *et al.*⁷. For patients with cough, saliva collection was preceded by induction with nebulized 3% NaCl to thin phlegm. Subjects were asked to transfer the collected saliva (a minimum amount of 5 ml) from the mouth to a falcon tube. Saliva samples were immediately deposited into a thermos of ice at 4°C for up to 2 h to protect the unstable analyte and prevent the growth of bacteria prior to long-term storage in a refrigerator at -40°C until use.

To avoid the possibility of contamination from substances in the saliva that may interfere with the immunoassay, the following precautions for participants were applied: avoid consumption of a meal 1 h before sample collection; avoid dairy products 20 min before sample collection; avoid foods with high acidity, high sugar, or caffeine immediately before sample collection; and rinse mouth with water to remove food residue before sample collection.

Statistical analysis

The sensitivity and specificity, of an immune response to Ag38-rec in children with a TB score of ≥ 6 and those with a score < 6 were determined and compared to those values for PPD. The statistical significance of the differences between the sensitivity and specificity of the immune response to Ag38-rec and to PPD was analyzed using the Student's t-test. A p-value of less than 0.05 was considered statistically significant. All data were analyzed using SPSS version 15.

Results

Dataset 1. Raw data for Raras *et al.* 2015, 'Evaluation of immunologic response of salivary sIg-A in pediatric tuberculosis patients to antigen Ag38-rec of *Mycobacterium tuberculosis* Indonesian strain'

<http://dx.doi.org/10.5256/f1000research.7234.d106955>

A detailed description of the raw data can be found in the text file provided ('Raw data legend').

Patient characteristics

In all three groups, the number of male patients was slightly higher than the number of female patients. The mean age of the subjects of all three groups was 8 years old. In the case of the TB group (score ≥ 6), most have had contact with an adult TB patient with a positive smear test. A positive TST result had been obtained from the majority of children from the TB group (22/26). In contrast, only a single positive TST result was found in the group of children with a suspected TB (score of < 6). Persistent cough for more than 3 weeks dominated the patients in both groups (score of ≥ 6 and < 6). All subjects in the group with pulmonary TB had X-rays suggestive of pulmonary TB, while only 6 subjects (6/28) in the suspected TB group had X-rays suggestive of pulmonary TB.

Purification of protein Ag38-rec

The antigen Ag38-rec was purified using purification kit Protino® NI-TED. It is shown that from the first purification step some copurified protein still appeared (Figure 1). However after dialysis the bands that corresponds to proteins with MW < 20 kDa could be removed. The purity of Ag38-rec reached 80%. The purified Ag38-rec was confirmed using Western blot against anti-Ag38. A single band could be detected in the range between 32 kDa and 52 kDa after incubation with chromogenic substrate (Figure 1.B).

Immune response of sIgA in saliva of children with TB (scoring ≥ 6) against Ag38-rec

The result of checker board showed that there is no significant difference between three concentration of Ag38-rec tested (Figure 2). A dark blue color indicates a reaction between Ag38-rec Mtb antigens with sIgA, while a pale color or white indicates no reaction. Therefore a minimal amount of 250 ng protein was used for the subsequent experiment.

To test the accuracy of anti Ag38-rec -sIgA in the diagnosis of active TB we compared the saliva of children with three different groups (TB. scores ≥ 6) and non TB (scores < 6) and healthy group. All samples from the three different groups were tested and the results of the dot blot are shown in Figure 3.

Comparison between response of sIg-A from TB group to Ag38-rec and the standard antigen PPD showed that sIg-A recognized PPD better than Ag38-rec (Table 1). Seventeen of 29 patients (65%) with active TB were positive for anti-Ag38-rec sIgA, which is higher than PPD (15/24,62%). However among non-TB group as well as healthy group, Ag38-rec could also recognize sIgA (19/26,73%) and (18/26,69%) respectively.

Figure 1. **A.** Purification process of Ag38-rec using Protino® NI-TED kit: (1) protein marker; (2) flowthrough; (3) washing I; (4) washing II; (5) elution. **B.** The result of Western Blot of Ag38-rec against anti Ag38: Protein marker (1): Elution (2–4). Arrow points to the position of Ag38-rec.

Figure 2. Checker board to test best crossreaction between antigen Ag38-rec and sIgA using different concentration of Ag38-rec: 500ng, 250ng and 125ng on nitrocellulose discs of dot-blot assay. The positive tests for anti Ag38-rec – sIgA in saliva from TB group (1–3), and negative test with sIgA in saliva of healthy group (5 to 7) and PPD served as positive control (9).

The strength of Ag38-rec as an immunodiagnostic agent to detect sIgA was determined based on the sensitivity and specificity. It was found that the sensitivity of sIg-A antibody response in saliva against Ag38-rec Mtb is significantly higher than PPD (70.8% vs 62.5%) ($p < 0.05$), although with lower specificity (26.9% vs 34.62%) ($p < 0.001$).

Comparison of sIgA responses to Ag38-rec between the confirmed TB group (score ≥ 6) and the suspected TB group (score < 6) (70.8% vs 73%), suggested that Ag38-rec could be recognized

among TB suspected children, although this difference was not statistically significant ($p = 0.861$) (Table 2).

Immune response of salivary sIg-A from TB patients with positive Tuberculin Skin Test result against Ag38-rec

We then compared the response of sIgA towards Ag38-rec in both groups based on the TST result. It was clear that patients with a positive TST responded significantly better to Ag38-rec than did patients with a negative TST result (77.2% vs. 41%, $p = 0.003$) (Table 3).

Figure 3. The response of salivary slg-A of TB group scores ≥ 6 (A) non TB group scores < 6 (B) healthy children (C) against Ag 38-rek and PPD.

Table 1. Comparison of positive response of salivary slg-A to Ag38-rec and PPD in TB patients group (scores ≥ 6) and non TB group (scores < 6).

Group	Ag38-rec			PPD		
	TB	Non TB	Healthy	TB	Non TB	Healthy
Positive Response	17(65%)	19(73%)	18(69%)	15(62%)	11(42%)	17(70%)
Negative Response	9(34%)	7(26%)	8(30%)	9(37%)	15(57%)	7(29%)
Total Tested	26	26	26	24	26	24

Table 2. Comparison of salivary slg-A response from TB patient with score ≥ 6 and non TB with score < 6 against Ag38-rec.

Type of Response	Patient's Group		Total
	Scoring ≥ 6	Scoring < 6	
Positive	17	19	36
Negative	7	7	14
Percentage(+)	70.8%	73%	

Table 3. Immune response of saliva slg-A against Ag38-rec in children with tuberculin skin test.

TST	Patient's Group		Total	Response(+)	Percentage
	Scoring ≥ 6	Scoring < 6			
Positive	21	1	22	17	77.2%
Negative	3	14	17	7	41%
Total Tested	24	15	39	24	

Discussion

The diagnosis of TB in children in Indonesia is conducted using a scoring system¹. This system is not a gold standard to diagnose TB in children, but rather a consensus by the WHO for areas that do not have the facilities for complete TB screening⁷. Therefore, it may be beneficial to add another complementary parameter to support the scoring determination.

The objective of the current study was to observe whether the Mtb Ag38-rec antigen could be used to discriminate children that are diagnosed with TB (scoring ≥ 6) from the suspected TB (scoring < 6) based on their immune response in salivary sIgA. However, we found that the positive response of salivary sIgA against Ag38-rec was lower for children with pulmonary TB than for children with suspected TB, albeit this difference was not statistically significant (73% vs 70.8%, $p = 0.861$). It could be that because the children with scores < 6 were recruited from Malang, Indonesia where TB is endemic; therefore these children may be infected with TB but still in the incubation stage of the disease, resulting in negative TST results¹². When we consider the potential of the Ag38-rec antigen as a serodiagnostic agent in pediatric TB compared to the PPD antigen, Ag38-rec has a significantly higher sensitivity than PPD (70.83% vs. 62.5%, $p < 0.001$).

Although the 38-kDa Mtb antigen is the most widely studied antigen for serological purposes and is a major component in commercial TB tests¹³, the serum antibodies showed high heterogeneity against Mtb antigens so that specific antibody response requires high purity of antigen¹⁴. In this study, the purity of Ag38-rec was not very high (80%). Nevertheless, the sensitivity of Ag38-rec compared to PPD may indicate that the immunogenicity value of Ag38-rec is slightly better than that of PPD. A study conducted in children of the Warao Amerindian tribes in Venezuela against Mtb-specific antibodies, including sIgA in saliva, demonstrated that PPD and Ag38-rec had a higher reactivity than other antigens¹⁰. This immunodominant antigen in patients with smear-positive TB is apparently specific for Mtb bacteria. A humoral immune response against this protein is often associated with active TB disease. Since our research participants were children that could not produce sputum, we could not confirm whether the low sensitivity of our Ag38-rec antigen was a result of negative sputum smears from the children. A previous study using the same antigen demonstrated that the response of sIgA from AFB-positive adult TB patients had a sensitivity of up to 80%, although, with a lower specificity (38%)⁹. This suggests that the immunogenicity of Ag38-rec is better than that of PPD. The explanation for this may be the fact that the salivary sIgA antibody is part of the mucosal immune response that belongs to the naive immune response. These immune responses are produced earlier in infection than the cellular immune response (the immune response of TST). When the naive immune response alone is sufficient to

eradicate a pathogen, consequently the cellular immune response will not be developed. The route of TB infection is initiated via the mucosal immune response, followed by an adaptive immune response. If the Mtb bacteria have been eliminated by the mucosal immune response, there would be no cellular immune response produced¹⁴.

With respect to the specificity of the sIgA response to Ag38-rec, it showed that it was significantly lower than that of PPD (26.9% vs. 34.6%, $p < 0.001$). Considering that all subjects in this study were recruited from TB endemic areas therefore it is possible that the healthy subjects may have had contact with TB patients without being infected and produced antibodies against Mtb. Interestingly the variation in specificity was also observed even when the study subjects came from non-endemic areas¹⁵.

The positive control PPD also produced a low specificity despite PPD being a mixture of antigens secreted by the bacteria Mtb. This suggests that the problem of low specificity is not simply a property of Ag38-rec, but also was affected by sample population and variations in antibody levels. These results are consistent with a previous study using serum samples in which the largest proportion of positive antibody responses was detected from TB endemic areas¹⁰.

The reactivity of antibodies against Mtb antigens in children can vary slightly due to an immature immune system, and severity of the disease¹⁶. Therefore, their antibody response has a widely ranging sensitivity and specificity, from 14% to 85% and from 86% to 100%, respectively^{12,16}. Moreover, several factors such as the purity of the protein and its immunogenicity play an important role in antibody response. Patient characteristics, the severity of disease, the presence or absence of Mtb bacteria in sputum, and the type and origin of the antibody should be considered¹³. Finally, last but not least is the process of taking the saliva sample. In previous experiments, whole saliva exhibited a greater antibody response than the supernatant or pellet⁹. Although, Hagewald *et al.* found that centrifugation of saliva resulted in a greater sIgA antibody response in saliva supernatant¹⁷. In this study, due to the limited volume of saliva in children, we used whole saliva. It is perhaps worthwhile to test saliva supernatants for comparison.

An interesting phenomenon occurred as we connected a positive immune response to the Ag38-rec antigen in children with suspected TB (score < 6) and a positive TST result. Considering that a positive TST indicates an infection with Mtb bacteria¹⁸, the fact that sIgA in saliva showed a greater reactivity against Ag38-rec in children with a positive TST (77.2%) compared to children with a negative TST (41%) indicates that the Ag38-rec antigen is able to differentiate infected children within the group of suspected TB patients (score < 6). Similar results showed a significant difference between the levels of anti-Ag38-rec sIgA in individuals

with a positive TST and a negative TST within the control group¹⁹. However Araujo *et al.*, found that there was no significant difference in the antibody response between patients with a positive TST and a negative TST¹⁹.

The specificity of Ag38-rec among healthy children compared to that of PPD suggests that Ag38 may potentially be used to identify TB infected children among children with suspected TB (scores < 6). Most of these children had a negative TST result (90%); however, 30% had a positive response to Ag38-rec.

This study has several limitations. The ideal method for saliva collection is passive drooling; however, we were unable to use this method because the children refused it. The criteria used for the determination of healthy child controls are also a limitation of this study. The control group was determined based on the criteria laid out in the scoring parameters for TB. They were children who did not meet the criteria for suspected TB and were very healthy.

Conclusion

The salivary sIgA from TB (scores ≥ 6) and non TB (scores < 6) patients demonstrated no significantly different immune response towards antigen Ag38-rec.

Data availability

F1000Research: Dataset 1. Raw data for Raras *et al.* 2015, 'Evaluation of immunologic response of salivary sIg-A in pediatric

tuberculosis patients to antigen Ag38-rec of *Mycobacterium tuberculosis* Indonesian strain', [10.5256/f1000research.7234.d106955](https://doi.org/10.5256/f1000research.7234.d106955)²⁰

Consent

Written informed consent for publication of their clinical details was obtained from the parents of the patients.

Author contributions

TMR, DE and CK conceived the study. TMR and DE designed the experiments. DE and TMR carried out the research. CK provided expertise in clinical analysis. DE and TMR prepared the first draft of the manuscript. All authors were involved in the revision of the draft manuscript and have agreed to the final content.

Competing interests

No competing interests were disclosed.

Grant information

The author(s) declared that no grants were involved in supporting this work.

Acknowledgments

We thank Suci Megasari, MS from Central Laboratory of Biomedical Science, Faculty of Medicine, Brawijaya University for technical assistance.

References

1. "Indonesian Pediatric Association": *National Guidance in TB Diagnosis in Children*, 2008.
2. Triasih R, Graham SM: **Limitations of the Indonesian Pediatric Tuberculosis Scoring System in the context of child contact investigation.** *Paediatr Indones*. 2011; **51**: 332–7. [Reference Source](#)
3. Chiang IH, Suo J, Bai KJ, *et al.*: **Serodiagnosis of tuberculosis. A study comparing three specific mycobacterial antigens.** *Am J Respir Crit Care Med*. 1997; **156**(3 Pt 1): 906–911. [PubMed Abstract](#) | [Publisher Full Text](#)
4. Pottumarthy S, Wells VC, Morris AJ: **A comparison of seven tests for serological diagnosis of tuberculosis.** *J Clin Microbiol*. 2000; **38**(6): 2227–2231. [PubMed Abstract](#) | [Free Full Text](#)
5. Araujo Z, Giampietro F, Cançado LC, *et al.*: **Comparison of serological responses in two different populations with pulmonary tuberculosis.** *Mem Inst Oswaldo Cruz*. 2008; **103**(7): 661–667. [PubMed Abstract](#) | [Publisher Full Text](#)
6. Alvarez N, Otero O, Camacho F, *et al.*: **Passive administration of purified secretory IgA from human colostrum induces protection against *Mycobacterium tuberculosis* in a murine model of progressive pulmonary infection.** *BMC Immunol*. 2013; **14**(Suppl 1): S3. [PubMed Abstract](#) | [Free Full Text](#)
7. Chiappin S, Antonelli G, Gatti R, *et al.*: **Saliva specimen: a new laboratory tool for diagnostic and basic investigation.** *Clin Chim Acta*. 2007; **383**(1–2): 30–40. [PubMed Abstract](#) | [Publisher Full Text](#)
8. Jung SB, Yang CS, Lee JS, *et al.*: **The mycobacterial 38-kilodalton glycolipoprotein antigen activates the mitogen-activated protein kinase pathway and release of proinflammatory cytokines through Toll-like receptors 2 and 4 in human monocytes.** *Infect Immun*. 2006; **74**(5): 2686–2696. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
9. Raras TY, Sholeh G, Lyrawati D: **Salivary sIg-A response against the recombinant Ag38 antigen of *Mycobacterium tuberculosis* Indonesian strain.** *Int J Clin Exp Med*. 2014; **7**(1): 129–35. [PubMed Abstract](#) | [Free Full Text](#)
10. Araujo Z, Waard JH, Fernández de Larrea C, *et al.*: **Study of the antibody response against *Mycobacterium tuberculosis* antigens in Warao Amerindian children in Venezuela.** *Mem Inst Oswaldo Cruz*. 2004; **99**(5): 517–524. [PubMed Abstract](#) | [Publisher Full Text](#)
11. Raras TY, Lyrawati D: **Cloning and expression of *pab* gene of *M. tuberculosis* isolated from pulmonary TB patient in *E. coli* DH5 [alpha].** *Med J Indones*. 2011; **20**(4): 247. [Reference Source](#)
12. Abbas AK, Lichtman AH, Pillai S: **Cellular and molecular immunology**. 2010. [Reference Source](#)
13. Achkar JM, Ziegenbalg A: **Antibody responses to mycobacterial antigens in children with tuberculosis: challenges and potential diagnostic value.** *Clin Vaccine Immunol*. 2012; **19**(12): 1898–1906. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
14. Uma Devi KR, Ramalingam B, Brennan PJ, *et al.*: **Specific and early detection of IgG, IgA and IgM antibodies to *Mycobacterium tuberculosis* 38kDa antigen in pulmonary tuberculosis.** *Tuberculosis (Edinb)*. 2001; **81**(3): 249–253. [PubMed Abstract](#) | [Publisher Full Text](#)
15. Raviglione MC: **Tuberculosis: the essentials.** CRC Press, 2009. [Reference Source](#)
16. Abebe F, Holm-Hansen C, Wiker HG, *et al.*: **Progress in serodiagnosis of *Mycobacterium tuberculosis* infection.** *Scand J Immunol*. 2007; **66**(2–3): 176–191. [PubMed Abstract](#) | [Publisher Full Text](#)
17. Hägewald SJ, Fishel DL, Christan CE, *et al.*: **Salivary IgA in response to periodontal treatment.** *Eur J Oral Sci*. 2003; **111**(3): 203–208. [PubMed Abstract](#) | [Publisher Full Text](#)
18. Coulter JB: **Diagnosis of pulmonary tuberculosis in young children.** *Ann Trop Paediatr*. 2008; **28**(1): 3–12. [PubMed Abstract](#) | [Publisher Full Text](#)
19. Sitienei JK, Kipruto H, Borus P, *et al.*: **Predictors of Low Body Mass Index Among Patients with Smear Positive Pulmonary Tuberculosis in Kenya.** *Int J Trop Dis Health*. 2014; **4**(4): 427–436. [Publisher Full Text](#)
20. Raras TYM, Pritta Santi DE, Kusuma HC: **Dataset 1 in: Evaluation of immunologic response of salivary sIg-A in pediatric tuberculosis patients to antigen Ag38-rec of *Mycobacterium tuberculosis* Indonesian strain.** *F1000Research*. 2015. [Data Source](#)

Open Peer Review

Current Peer Review Status:

Version 4

Reviewer Report 01 July 2016

<https://doi.org/10.5256/f1000research.9761.r14705>

© 2016 Yuan D. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Dorothy Yuan

Department of Pathology, UT Southwestern Medical Center at Dallas, Dallas, TX, 75390, USA

The authors have not adequately explained the results presented in Fig. 1.

If A represents patients with scores greater than 6 why are they generally more negative than samples in C collected from healthy donors?

Are the samples shown vertically repeats of 1 patient or from individual patients? If so there are in total only 20 samples instead of 26.

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

Reader Comment 06 Aug 2016

Tri yudani Mradining Raras, Department of Biochemistry and Molecular Biology, Faculty of Medicine, Brawijaya University, Jl Veteran, Malang, Indonesia

Thank you for your comment.

I have put some more explanation for Fig 1.

Concerning the second question. Truly we do not know the exact answer, it also wonders us, because our preliminary experiment using checker board demonstrated that saliva from TB and healthy children produced a significantly different immune response against Ag38rec (Fig.2). Furthermore the PPD we used as positive control also showed the similar result compared to our antigen.

We exclude the possibility that the problem is due to the non-specific criteria during TB screening (such as lymphadenitis, fever and coughing) because we also performed Tuberculin Skin Test (TST) and X-rays that showed 100% positive for all TB suspected children. Unfortunately we did not perform TST test and X-ray for healthy children, since there was no indication that they were sick. So the most likely reason could be that due to impurity of our Ag38rec protein that consequently did not recognize salivary sIgA consistently.

Your third question concerning the number of sample we used. There were 26 six samples, it is just that, due to the limited number of hole in the apparatus (92), we can only load 20 samples in each run. The rest of sample (6) we loaded in the second run.

Competing Interests: No competing interests were disclosed.

Version 3

Reviewer Report 03 May 2016

<https://doi.org/10.5256/f1000research.8725.r12998>

© 2016 Musoke P et al. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Philippa M Musoke

The Makerere University - Johns Hopkins University Research Collaboration, Kampala, Uganda

Willy Ssengooba

Department of Medical Microbiology, College of Health Sciences, Makerere University, Kampala, Uganda

This manuscript covers an important area of novel diagnostic tests for paediatric TB. They highlight the well known challenge of obtaining appropriate specimens for identifying mycobacterium tuberculosis in children with PTB. The authors report on the use of an in-house sIgA assay for diagnosis of pulmonary TB in children. It is not clear if the sIgA assay has been standardized or validated, so I asked a microbiologist to also review the manuscript. The authors found no difference in the sIgA assay between the children with PTB and those without TB. If the sIgA assay was clearly described and validated, the findings may have provided useful information for future studies. However, the methodology of the assay has major limitations.

Co-reviewer: a microbiologist comments

1. The results are still not well presented to trust the conclusions.
2. The details on how the optimal concentrations of Ag38-rec, PPD and volume of saliva was determined are missing yet they are needed to yield confidence in the author's procedures.

3. Whether the second purification step of protein Ag38-*rek* was optimized after the failure of the purification kit remains unclear.
4. The use of PPD as a positive control and a comparator is not clear.
5. Results of the dot blot are very subjectively interpreted yet even the criteria is not clear to ensure reliability.

Generally, I find the methods and results are not well presented to enable critical understanding of the findings and relate them to the conclusion.

Competing Interests: No competing interests were disclosed.

We confirm that we have read this submission and believe that we have an appropriate level of expertise to state that we do not consider it to be of an acceptable scientific standard, for reasons outlined above.

Author Response 15 Jun 2016

Tri yudani Mradining Raras, Department of Biochemistry and Molecular Biology, Faculty of Medicine, Brawijaya University, Jl Veteran, Malang, Indonesia

Thank you for your review. I apologize for the late response. Concerning your comments, here are my answers:

1. We have made several changes in Results.
2. The optimization of the amount of Ag38-*rec* to produce the strongest immune response to sIgA was conducted using checker board containing several concentration of Ag38-*rec* i.e 500 ng, 250 ng and 125 ng. We provide figure 2. What we meant in the sentence ...“the best concentration of Ag38-*rec*with 25 ul saliva”, is that we used 25ul saliva in all concentration of Ag38-*rec* in checkerboard, but we did not make any optimization in the amount of saliva used in this experiment. Because it was already done in previous experiment (Tri Yudani et al, 2014). Actually the correct amount is 50 ul. But I have made correction.
3. The purification of protein did not fail. We used IMAC (Immobilized Metal Affinity Chromatography) kit. It is very straightforward method for protein purification. We did not do any improvement after second purification step, we have just followed with dialysis step after protein was eluted from the kit. Dialysis belong to the purification procedure, that is why we did not verify the result of dialysis process in SDS PAGE to avoid unnecessary wasting of Antigen 38.
4. PPD (purified protein derivative) used in Tuberculin sensitivity test, a screening tool for tuberculosis (TB) Tuberculin sensitivity test. That is why PPD is commonly used as a positive control in testing for new protein used for diagnostic agent. (see [Chan et al. , 2000](#))
5. The Corel Draw graphic suite X4 (Corel, USA) program was used to interpret the color range of the blot(s). The gradation of color was quantified to numerical value using Corel Photopaint 11. As standard we used PPD. Positive and negative value was based

on the cut-off of the median value of the positive control (PPD) after reaction with saliva of the TB group. A dot was regarded as positive when the value was below the cut-off point and as negative when the value was higher than the cut-off point. A positive result was defined as a dark blue or dark purple spot (>50%) on the blot.

Competing Interests: No competing interests were disclosed.

Reviewer Report 19 February 2016

<https://doi.org/10.5256/f1000research.8725.r12523>

© 2016 Yuan D. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Dorothy Yuan

Department of Pathology, UT Southwestern Medical Center at Dallas, Dallas, TX, 75390, USA

Upon my query as to why the authors did not utilize anti-human IgA for their assay the authors now responded by saying that they did indeed use this reagent and that the original description was in error.

Now that I can understand how the test was conducted I still find the raw data quite uninterpretable.

To be fair to the authors I do not feel I should contribute further to the peer review of this article.

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

Version 2

Reviewer Report 03 February 2016

<https://doi.org/10.5256/f1000research.8443.r12246>

© 2016 Yuan D. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Dorothy Yuan

Department of Pathology, UT Southwestern Medical Center at Dallas, Dallas, TX, 75390, USA

In this revision the author only addressed one of the two major problems. They added a figure to show the purity of the their preparation. However, they still failed to provide the rationale for the method used for determining titers. The method is not understandable as presented. Furthermore, anti-IgA was not used at any point so it is not clear that they are testing for IgA.

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

Reader Comment 12 Feb 2016

Tri yudani Mradining Raras, Department of Biochemistry and Molecular Biology, Faculty of Medicine, Brawijaya University, Jl Veteran, Malang, Indonesia

Thank you for your comment. After I checked the serological test, I found a mistype concerning the secondary antibody I used in DotBlot method. Truly, instead of mouse anti-*Mycobacterium tuberculosis* Ag38 monoclonal antibody (AbD, Serotec, England, cat.no.0100-0519)[T1] I used Anti-human IgA (alpha-chain specific) alkaline phosphatase conjugate (SIGMA, USA.Product.no. A.9669[T2]).

I hope that this correction could make a rationale explanation for your question.

Competing Interests: No competing interests were disclosed.

Version 1

Reviewer Report 15 December 2015

<https://doi.org/10.5256/f1000research.7794.r11488>

© 2015 Yuan D. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Dorothy Yuan

Department of Pathology, UT Southwestern Medical Center at Dallas, Dallas, TX, 75390, USA

In this submission the authors wish to evaluate the efficacy of using a protein (Ag38-rec) from *Mycobacterium tuberculosis* to determine whether it can be used to differentiate between pediatric patients who have TB from those suspected of disease. The use of salivary IgA is a promising non-

invasive approach for testing pediatric patients.

They conclude that the approach may be useful for identifying presumptive infected patients. Unfortunately the submission suffers from two major flaws that prevent any confirmation of these conclusions. Firstly, no data was presented to show the purity of the Ag38-rec produced in their laboratory. Secondly, the rationale and reliability of the method used to determine the titer of salivary IgA was not understandable to this reviewer such that it is impossible to evaluate their results. It is unclear why they did not utilize a more traditional method¹ of using anti-human IgA antibodies in the Elisa.

References

1. Uma Devi KR, Ramalingam B, Brennan PJ, Narayanan PR, et al.: Specific and early detection of IgG, IgA and IgM antibodies to Mycobacterium tuberculosis 38kDa antigen in pulmonary tuberculosis. *Tuberculosis (Edinb)*. 2001; **81** (3): 249-53 [PubMed Abstract](#) | [Publisher Full Text](#)

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

F1000Research