REVIEW

Novel Insights and Therapeutics in Multiple Sclerosis [version 1; peer review: 3 approved]

Catriona A. Wagner, Joan M. Goverman

Department of Immunology, University of Washington, Seattle, WA, 98109-8509, USA

V1 First published: 07 Aug 2015, 4(F1000 Faculty Rev):517
<https://doi.org/10.12688/f1000research.6378.1>
Latest published: 07 Aug 2015, 4(F1000 Faculty Rev):517
<https://doi.org/10.12688/f1000research.6378.1>

Abstract

The last twelve years have witnessed the development of new therapies for relapsing-remitting multiple sclerosis that demonstrate increased efficacy relative to previous therapies. Many of these new drugs target the inflammatory phase of disease by manipulating different aspects of the immune system. While these new treatments are promising, the development of therapies for patients with progressive multiple sclerosis remains a significant challenge. We discuss the distinct mechanisms that may contribute to these two types of multiple sclerosis and the implications of these differences in the development of new therapeutic targets for this debilitating disease.

Keywords

multiple sclerosis, immune, therapeutics

Open Peer Review

Approval Status

	1	2	3
version 1 07 Aug 2015			
1. Lars Fugger , University of Oxford, Oxford, UK			
2. Melissa A. Brown , Northwestern University Feinberg School of Medicine, Chicago, USA			
3. Benjamin M. Segal , University of Michigan, Ann Arbor, USA			
Any comments on the article can be found at the end of the article.			

Corresponding author: Joan M. Goverman (goverman@uw.edu)

Competing interests: No competing interests were disclosed.

Grant information: This work was supported by grants to J.M. Goverman from NIAID (R37 AI107494-01) and the National Multiple Sclerosis Society (RG 4792A6/1), and by a grant from NIAID supporting C.A. Wagner (T32 AI106677). The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Copyright: © 2015 Wagner CA and Goverman JM. This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Wagner CA and Goverman JM. **Novel Insights and Therapeutics in Multiple Sclerosis [version 1; peer review: 3 approved]** F1000Research 2015, 4(F1000 Faculty Rev):517 <https://doi.org/10.12688/f1000research.6378.1>

First published: 07 Aug 2015, 4(F1000 Faculty Rev):517 <https://doi.org/10.12688/f1000research.6378.1>

Introduction

Multiple sclerosis (MS) is an inflammatory, demyelinating disease of the central nervous system (CNS) and is the most common cause of non-traumatic neurologic disability in young adults. Although the etiology of MS has been debated in the past, recent data from GWAS studies provide exceptionally strong evidence that MS is an autoimmune disease^{1–3}. This view was originally based on the observation that experimental autoimmune encephalomyelitis (EAE), an animal model that recapitulates many important features of MS, is induced by the activation or adoptive transfer of self-reactive CD4⁺ T cells specific for myelin proteins⁴. Furthermore, susceptibility to MS is most strongly associated with MHC class II alleles¹. Inflammatory lesions and plaques of demyelination in the CNS are considered hallmark features of MS; however, substantial heterogeneity (with respect to clinical course, pathology, and response to therapies) is seen among patients. This review will focus on current ideas regarding the role of different pathological mechanisms in shaping these different manifestations of MS. Achieving an understanding of the specific pathogenic pathways that are relevant to individual patients with MS is critically important in order to predict which patients will respond well to current therapies, and to identify new therapeutic targets that can be tailored for patients with different types of MS.

Heterogeneity in MS

For the majority of patients with MS, disease course begins with a relapsing-remitting phase with intermittent, discrete periods of neurological symptoms that coincide with the appearance of inflammatory lesions. Over time, most patients with relapsing-remitting MS convert to a progressive stage called secondary progressive MS, characterized by a decrease in the frequency, or complete cessation, of relapses and gadolinium-enhancing MRI lesions, and the gradual accumulation of disability associated with brain and spinal cord atrophy. A small subset of patients with MS initially present with a progressive disease course that is not preceded by clinical exacerbations. This form of MS is called primary progressive MS because it is not preceded by a relapsing-remitting phase^{5,6}.

Significant heterogeneity is also observed in the structure of lesions and types of tissue injury seen in patients with MS. Lesions have been grouped into four different patterns, based on the presence or absence of antibody deposition and complement activation, differential loss of certain myelin proteins, whether lesions occur at perivenous sites, and whether oligodendrocytes are spared or die by apoptosis or necrotic cell death^{7,8}. The observation that individual patients with MS exhibit only one pattern of lesions led to the proposal that different patterns may arise from distinct pathogenic pathways. However, these patterns have not been associated with particular clinical disease courses. Differences are also seen among patients in the distribution of lesions between the brain and spinal cord. The vast majority of patients with relapsing-remitting MS exhibit numerous lesions in the cerebral white matter. However, a small subset (10–15%) of patients with MS exhibit lesions predominantly in the spinal cord with relatively sparse brain involvement.

Relapsing-remitting MS pathogenesis and therapies

Relapsing-remitting MS is the best studied form of MS, as the majority of patients initially exhibit this form of disease, and the EAE model recapitulates many aspects of the inflammatory lesions

seen in relapsing-remitting MS. Both MRI and immunohistochemical analyses of tissue sections indicate that inflammation is the key component leading to tissue injury and clinical relapses in relapsing-remitting MS patients. Perivascular lesions are comprised of inflammatory infiltrates dominated by lymphocytes and myeloid cells. Our conceptual framework for understanding how these lesions arise is based on studies in EAE. In EAE, activation of myelin-specific T cells induces expression of adhesion molecules and integrins that facilitate their extravasation across the blood brain barrier (BBB). Upon entry into the CNS, myelin-specific T cells are reactivated by the small number of antigen-presenting cells (APCs) in the healthy CNS that constitutively present myelin antigens in the perivascular and subarachnoid spaces. This reactivation triggers the T cells to produce soluble, inflammatory mediators that cause BBB permeability and recruitment of a range of inflammatory leukocytes⁴. Formation of a localized, inflammatory environment within lesions results in plaques of demyelination and axonal damage.

In EAE, CD4⁺ T cells are the predominant lymphocyte in the infiltrate, as the protocol for inducing EAE specifically primes CD4⁺ T cells. However, there is an abundance of CD8⁺ T cells in lesions in MS patients, and clonal expansion indicative of antigen-driven activation is more evident in the CD8⁺ T cell subset in the blood and cerebrospinal fluid of MS patients. Furthermore, depletion of CD4⁺ T cells resulted in limited therapeutic efficacy in MS patients⁹, although the results of this trial may not be conclusive. In contrast, treatments that deplete all leukocytes demonstrated greater efficacy in MS patients¹⁰. These observations raise key questions (Box 1) about the role for different lymphocyte subsets in CNS autoimmunity.

Box 1. Outstanding questions regarding the role of lymphocyte subsets in relapsing-remitting MS

- Do both CD4 and CD8 T cells contribute to the pathogenesis of MS? What are their relevant effector mechanisms?
- Do CD8⁺ T cells exhibit both regulatory and pathogenic activity as seen for different subsets of CD4⁺ T cells?
- How do B cells contribute to the pathogenesis of MS?

The case for CD4 T cells

An important role for CD4⁺ T cells in MS is clear from the strong association of MHC class II molecules with disease susceptibility¹. Studies of effector CD4⁺ T cells in EAE implicated both IFN- γ -producing Th1 and IL-17-producing Th17 cells as pathogenic mediators¹¹. Originally, Th1 cells were considered the main effector cells, as adoptive transfer of Th1 cells could induce EAE^{12,13}. This finding was consistent with an earlier observation that MS was exacerbated by the administration of IFN- γ ¹⁴. However, mice deficient in cytokines associated with the differentiation and function of Th1 cells developed severe EAE¹⁵. In contrast, deficiency in IL-23, which is important for Th17 cell stabilization, conferred resistance to EAE, suggesting that Th17 cells may be the true effector cells¹⁶. An increase in IL-17 transcripts was also reported in chronic MS lesions¹⁷. However, IL-17A^{-/-} and IL-17F^{-/-} mice, treated with anti-IL-17A blocking antibody, are still susceptible to EAE¹⁸.

Subsequently, GM-CSF was reported to be essential for EAE pathogenesis^{19–21}, and recent studies have suggested that T cells producing GM-CSF may represent a distinct T cell subset^{22,23}.

The studies described above were all carried out in C57BL/6 mice. Our studies of EAE in C3Heb/Fej mice have provided further insight into the complexity of EAE pathogenesis. While the incidence of EAE in C3Heb/Fej mice is strongly reduced in mice lacking both IL-17RA and IFN- γ receptors, disease incidence was only modestly reduced in mice that lacked either only IL-17 or only IFN- γ signaling. Importantly, we found that IFN- γ and IL-17 signaling had a much greater impact on the pattern of lesion localization within the CNS. IL-17 promoted inflammation in the brain *via* induction of chemokines that recruit neutrophils, and neutrophils contributed significantly to parenchymal tissue damage in the brain²⁴. In contrast, IFN- γ inhibited inflammation in the brain by inhibiting neutrophil recruitment. Surprisingly, IFN- γ exerted the opposite influence in the spinal cord by promoting both neutrophil recruitment and inflammation in this microenvironment. Despite the enhanced, IFN- γ -mediated neutrophil recruitment to the spinal cord, neutrophils contributed less to spinal cord tissue damage compared to their role in the brain²⁴. Recent work by Segal and colleagues in C57BL/6 mice also suggested that neutrophils may be more important effector cells in the brain compared to the spinal cord²⁵. Interestingly, in contrast to EAE in C57BL/6 mice, we have observed only a modest reduction in disease incidence in C3Heb/Fej mice when EAE is induced by adoptive transfer of GM-CSF-/- T cells in C3Heb/Fej mice, suggesting that the stringent requirement for GM-CSF seen in C57BL/6 mice may be a strain-specific finding (Pierson E.R., Johnson M.C., and Goverman J.M., unpublished observations). Collectively, these studies demonstrate that it is critical to study different mouse strains in order to understand the complexity of disease manifestation in MS patients.

The finding that the brain and spinal cord microenvironments in mice respond very differently to cytokines produced by infiltrating T cells suggests that patients with distinct neuroinflammatory patterns may respond quite differently to therapies that target specific cytokines. There are also other challenges in designing cytokine-based therapies for MS patients. Despite the substantial data supporting key roles for Th1 and Th17 cells in EAE, a clinical trial administering ustekinumab (an antibody that neutralizes cytokines that promote differentiation of both Th1 and Th17 cells), had no beneficial effect²⁶. It is difficult to draw conclusions from this one trial, however, especially in light of the fact that it is not known whether effector T cell differentiation occurs in the periphery or the CNS, or how important ongoing T cell differentiation is in patients with established MS. The dramatic benefit seen in patients with psoriasis following administering of an IL-17-neutralizing antibody has also not yet been reported for similar clinical trials in patients with MS. It is possible that better stratification of patients with MS, with respect to their neuroinflammatory pattern and other key disease characteristics, is needed to properly evaluate the effectiveness of therapeutic targeting of specific cytokines.

The case for CD8 T cells

CD8+ T cells often predominate in tissue sections and in CSF of MS patients, and clonal expansion is more commonly observed in

the CD8+ compared to the CD4+ T cell subset^{4,27,28}. However, the role of CD8+ T cells is still unclear, as EAE models have pointed to both pathogenic and regulatory functions. Global elimination of CD8+ T cells using either CD8-/- mice or antibody-mediated depletion of CD8+ T cells *in vivo* suggested a regulatory role for CD8+ T cells^{29,30}. The observations that Qa-1-deficient mice exhibit increased susceptibility to EAE, and that adoptive transfer of Qa-1-restricted CD8+ T cells ameliorates disease, suggested that there may be distinct regulatory subsets of CD8+ T cells^{31,32}. Other studies have reported a pathogenic role for myelin-specific CD8+ T cells in CNS autoimmunity^{33–35}, and animal models using neo-antigens expressed in the CNS and CD8+ T cells that recognize the neo-antigen support a pathogenic role for CD8+ T cells^{36–40}. We identified CD8+ T cells that recognize a MHC class I-restricted myelin basic protein (MBP) epitope and showed that these CD8+ T cells were pathogenic and produced lesions distinct from those seen in conventional EAE but similar to some lesions seen in patients with MS^{33,41,42}. We also showed that both dendritic cells and oligodendrocytes presented the MHC class I-restricted epitope of MBP within the CNS of mice with CD4+ T cell-initiated EAE⁴³. We speculate that CD8+ T cells could be pathogenic if they are triggered to produce inflammatory cytokines upon encountering dendritic cells and/or lyse oligodendrocytes, but they might ameliorate disease if they subsequently lyse dendritic cells that present antigen to both CD4+ and CD8+ T cells within the CNS. Our preliminary data suggest that recruitment of MBP-specific CD8+ T cells during disease induction can exacerbate CD4+ T cell-initiated EAE and may enhance brain inflammation (Wagner C.A. and Goverman J.M., unpublished data). However, CD8+ T cells may play different roles at different phases of disease, and it is important to identify their specific effects during each disease stage in order to therapeutically target (or harness) their activity.

The case for B cells

A pathogenic role for B cells in MS is suggested by the therapeutic benefit observed in patients treated with anti-CD20 monoclonal antibodies (rituximab or ocrelizumab) that deplete B cells^{44,45}. As anti-CD20 does not deplete antibody-producing plasma cells, the ability of B cells to present antigen to T cells may be critical in CNS autoimmunity. In animal models, B cells have been shown to promote EAE induction by acting as antigen-presenting cells for T cells in both the periphery^{46–49} and CNS⁵⁰. We found that in the healthy CNS, B cells comprise the majority of MHC class II+ cells, and that they play a role in the initial reactivation of infiltrating myelin-specific T cells, specifically the Th1 subset⁵⁰. Despite the therapeutic efficacy of B cell depletion in MS, B cells that produce IL-10 and ameliorate EAE have been described^{51,52}. If these regulatory B cells retain a stable phenotype in patients, transfer of this subset may be beneficial, in addition to depleting B cells that function only to present antigen to T cells⁵³.

Secondary progressive MS pathogenesis and therapies

Following the success of anti-inflammatory therapies in relapsing-remitting MS, clinical trials were initiated in secondary progressive MS cohorts. However, clinical trials analyzing the effects of beta-interferon found no significant treatment effect, particularly in patients that had not exhibited MRI lesions for several years^{54–57}. The lack of efficacy of anti-inflammatory therapies in secondary

progressive MS suggested that neurodegeneration proceeds independently of inflammation in this stage⁵⁸. However, further analyses of normal-appearing white matter and meninges tissue sections revealed that inflammation was still present, albeit in a different form⁵⁹. Staining, using a marker that selectively stains for leaky endothelial cells, indicated that inflammation is compartmentalized behind a less permeable BBB⁶⁰. Differences in the pathology, clinical signs and response to current therapies suggest that different mechanisms predominate in progressive MS, requiring new therapeutic approaches.

Secondary progressive MS is characterized by increasing brain atrophy and accumulation of irreversible axonal and neuronal degeneration. Gadolinium-enhancing MRI lesions subside⁶¹ and a diffuse pattern of inflammation predominates in normal-appearing white and grey matter⁶². The mechanisms underlying these changes in pathology are poorly understood. Axonal transection begins early in relapsing-remitting MS⁶³; however, the mechanisms may differ in these two stages of disease. During relapsing-remitting MS, inflammatory cells are thought to mediate demyelination *via* secretion of degradative enzymes, production of oxidative products, and increased levels of glutamate that can damage oligodendrocytes *via* excessive NMDA receptor signaling⁶⁴. Thus, transected axons are more abundant in active lesions where inflammatory cells are localized *versus* chronic lesions in relapsing-remitting MS⁶³. In contrast, demyelination and axonal injury is primarily associated with microglia activation in secondary progressive MS⁶⁵. The pattern of diffuse parenchymal inflammation, as well as the presence of T and B cells in the meninges, may contribute to microglial activation; however the role of adaptive immune cells in facilitating, *versus* responding to, microglial activation remains to be established. A major consequence of microglial activation is the production of reactive oxygen species. Mitochondria and mitochondrial DNA are very susceptible to oxidative injury^{66,67}, and axons are extremely susceptible to mitochondrial dysfunction due to their high demand for ATP production to propagate action potentials⁶⁸. Failure of remyelination in progressive disease is another key difference between relapsing-remitting MS and secondary progressive MS. The state of chronic demyelination leads to diffusion of Na⁺ channels away from nodes of Ranvier and a subsequent influx of sodium and increased ATP consumption. The resulting energy imbalance ultimately leads to axonal degeneration and tissue damage⁶⁴. Thus, remyelination may be a critical therapeutic target in secondary progressive MS, together with strategies to dampen microglial activation. Potential contributions of adaptive immune cells may also need to be addressed in secondary progressive MS.

An additional challenge in designing effective therapies for secondary progressive MS is our lack of understanding of mechanisms that lead to subpial demyelination. Therapies that prevent or resolve subpial lesions may be very important in treating secondary progressive MS patients as increased progression in disability is associated with cortical atrophy⁶⁹, and subpial lesions are represented to a greater extent than leukocortical lesions in the total cortical lesion load^{62,70}. However, subpial lesions typically lack peripheral inflammatory cells and the mechanism of demyelination that produces these lesions, while unknown, may be distinct from mechanisms of demyelination in white matter. Additionally, meningeal inflammatory aggregates are present in some patients with secondary progressive

MS and some studies have identified lymph follicle-like structures. Post-mortem analyses of brain and spinal cord tissue revealed proliferating B cells and follicular dendritic cells in the follicles, suggesting germinal center formation⁷¹. The presence of these follicles correlates with disease severity and the extent of demyelination⁷². However, the contributions of these follicles to disease pathogenesis are still unclear.

Concluding remarks

In this review, we have highlighted potential differences in pathogenic mechanisms between relapsing-remitting MS and secondary progressive MS, and how these differences may require distinct therapeutic approaches. Lesions in relapsing-remitting MS patients arise from a complex interplay of both CD4⁺ and CD8⁺ T cells, with B cells and innate immune cells playing critical roles in orchestrating T cell responses. The efficacy of therapies such as Natalizumab⁷³ and fingolimod^{74,75} that reduce T cell entry into the CNS highlight the key role played by T cells in relapsing-remitting MS. While current therapies are relatively effective at reducing new MRI lesion formation and relapse rates, they are broadly anti-inflammatory and often associated with side effects. New therapies capable of targeting inflammation relevant only to the CNS are needed. In addition, patients with relapsing-remitting MS exhibit heterogeneity in CNS lesions, including the distribution of lesions within the CNS, reinforcing the need for treatments tailored to the individual patient. Finally, the long-term impact of reducing inflammatory lesions and clinical relapses during relapsing-remitting MS on the progression of neurological disability remains to be firmly established.

The distinct pathology seen in secondary progressive MS patients suggests that targeting different disease mechanisms may be important. Specifically, therapies that promote remyelination and prevent microglia activation, mitochondria dysfunction, and oxidative damage, while beneficial in relapsing-remitting MS, appear particularly crucial in treating patients with secondary progressive MS. It is also important to determine the exact role of inflammation during this phase to prevent relapse after tissue damage has been resolved. Because the BBB is more intact in secondary progressive MS, developing therapeutic agents capable of crossing the BBB is an additional challenge. A key to developing therapies for secondary progressive MS is the generation of new animal models that better reproduce the key features seen in this disease. While we have focused on secondary progressive MS, similarities in the pathology seen between secondary progressive MS and primary progressive MS support the hope that the same therapies will be beneficial in both types of disease.

Competing interests

No competing interests were disclosed.

Grant information

This work was supported by grants to J.M. Goverman from NIAID (R37 AI107494-01) and the National Multiple Sclerosis Society (RG 4792A6/1), and by a grant from NIAID supporting C.A. Wagner (T32 AI106677).

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

References

1. **F** International Multiple Sclerosis Genetics Consortium, Hafler DA, Compston A, *et al.*: **Risk alleles for multiple sclerosis identified by a genome-wide study.** *N Engl J Med.* 2007; **357**(9): 851–862.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
2. **F** International Multiple Sclerosis Genetics Consortium, Wellcome Trust Case Control Consortium, Sawcer S, *et al.*: **Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis.** *Nature.* 2011; **476**(7359): 214–219.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
3. **F** Bryneda B, Duvefelt K, Jonasdottir G, *et al.*: **HLA-A confers an HLA-DRB1 independent influence on the risk of multiple sclerosis.** *PLoS One.* 2007; **2**(7): e664.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
4. Goverman J: **Autoimmune T cell responses in the central nervous system.** *Nat Rev Immunol.* 2009; **9**(6): 393–407.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
5. **F** Aktas O, Kieseier B, Hartung HP: **Neuroprotection, regeneration and immunomodulation: broadening the therapeutic repertoire in multiple sclerosis.** *Trends Neurosci.* 2010; **33**(3): 140–152.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
6. Lublin FD, Reingold SC: **Defining the clinical course of multiple sclerosis: results of an international survey. National Multiple Sclerosis Society (USA) Advisory Committee on Clinical Trials of New Agents in Multiple Sclerosis.** *Neurology.* 1996; **46**(4): 907–911.
[PubMed Abstract](#) | [Publisher Full Text](#)
7. Lucchinetti C, Brück W, Parisi J, *et al.*: **Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination.** *Ann Neurol.* 2000; **47**(6): 707–717.
[PubMed Abstract](#) | [Publisher Full Text](#)
8. **F** Metz I, Weigand SD, Popescu BF, *et al.*: **Pathologic heterogeneity persists in early active multiple sclerosis lesions.** *Ann Neurol.* 2014; **75**(5): 728–738.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
9. van Oosten BW, Lai M, Hodgkinson S, *et al.*: **Treatment of multiple sclerosis with the monoclonal anti-CD4 antibody cM-T412: results of a randomized, double-blind, placebo-controlled, MR-monitored phase II trial.** *Neurology.* 1997; **49**(2): 351–357.
[PubMed Abstract](#) | [Publisher Full Text](#)
10. **F** CAMMS223 Trial Investigators, Coles AJ, Compston DA, *et al.*: **Alemtizumab vs. interferon beta-1a in early multiple sclerosis.** *N Engl J Med.* 2008; **359**(17): 1786–1801.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
11. **F** Kroenke MA, Carlson TJ, Andjelkovic AV, *et al.*: **IL-12- and IL-23-modulated T cells induce distinct types of EAE based on histology, CNS chemokine profile, and response to cytokine inhibition.** *J Exp Med.* 2008; **205**(7): 1535–1541.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
12. Baron JL, Madri JA, Ruddle NH, *et al.*: **Surface expression of alpha 4 integrin by CD4 T cells is required for their entry into brain parenchyma.** *J Exp Med.* 1993; **177**(1): 57–68.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
13. Segal BM, Shevach EM: **IL-12 unmasks latent autoimmune disease in resistant mice.** *J Exp Med.* 1996; **184**(2): 771–775.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
14. Panitch HS, Hirsch RL, Schindler J, *et al.*: **Treatment of multiple sclerosis with gamma interferon: exacerbations associated with activation of the immune system.** *Neurology.* 1987; **37**(7): 1097–1102.
[PubMed Abstract](#) | [Publisher Full Text](#)
15. **F** Steinman L: **A brief history of T_H17, the first major revision in the T_H1/T_H2 hypothesis of T cell-mediated tissue damage.** *Nat Med.* 2007; **13**(2): 139–145.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
16. **F** Cua DJ, Sherlock J, Chen Y, *et al.*: **Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain.** *Nature.* 2003; **421**(6924): 744–748.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
17. **F** Lock C, Hermans G, Pedotti R, *et al.*: **Gene-microarray analysis of multiple sclerosis lesions yields new targets validated in autoimmune encephalomyelitis.** *Nat Med.* 2002; **8**(5): 500–508.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
18. **F** Haak S, Croxford AL, Kreyenborg K, *et al.*: **IL-17A and IL-17F do not contribute vitally to autoimmune neuro-inflammation in mice.** *J Clin Invest.* 2009; **119**(1): 61–69.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
19. **F** Codarri L, Györfi G, Tosevski V, *et al.*: **RORγt drives production of the cytokine GM-CSF in helper T cells, which is essential for the effector phase of autoimmune neuroinflammation.** *Nat Immunol.* 2011; **12**(6): 560–567.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
20. **F** El-Behi M, Ciric B, Dai H, *et al.*: **The encephalitogenicity of T_H17 cells is dependent on IL-1- and IL-23-induced production of the cytokine GM-CSF.** *Nat Immunol.* 2011; **12**(6): 568–575.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
21. **F** Ponomarev ED, Shriver LP, Maresz K, *et al.*: **GM-CSF production by autoreactive T cells is required for the activation of microglial cells and the onset of experimental autoimmune encephalomyelitis.** *J Immunol.* 2007; **178**(1): 39–48.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
22. **F** Sheng W, Yang F, Zhou Y, *et al.*: **STAT5 programs a distinct subset of GM-CSF-producing T helper cells that is essential for autoimmune neuroinflammation.** *Cell Res.* 2014; **24**(12): 1387–1402.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
23. **F** Noster R, Riedel R, Mashreghi MF, *et al.*: **IL-17 and GM-CSF expression are antagonistically regulated by human T helper cells.** *Sci Transl Med.* 2014; **6**(241): 241ra280.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
24. Simmons SB, Liggitt D, Goverman JM: **Cytokine-regulated neutrophil recruitment is required for brain but not spinal cord inflammation during experimental autoimmune encephalomyelitis.** *J Immunol.* 2014; **193**(2): 555–563.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
25. **F** Stoolman JS, Duncker PC, Huber AK, *et al.*: **Site-specific chemokine expression regulates central nervous system inflammation and determines clinical phenotype in autoimmune encephalomyelitis.** *J Immunol.* 2014; **193**(2): 564–570.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
26. **F** Segal BM, Constantinescu CS, Raychaudhuri A, *et al.*: **Repeated subcutaneous injections of IL12/23 p40 neutralising antibody, ustekinumab, in patients with relapsing-remitting multiple sclerosis: a phase II, double-blind, placebo-controlled, randomised, dose-ranging study.** *Lancet Neurol.* 2008; **7**(9): 796–804.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
27. Friese MA, Fugger L: **Pathogenic CD8⁺ T cells in multiple sclerosis.** *Ann Neurol.* 2009; **66**(2): 132–141.
[PubMed Abstract](#) | [Publisher Full Text](#)
28. Mars LT, Saikali P, Liblau RS, *et al.*: **Contribution of CD8 T lymphocytes to the immuno-pathogenesis of multiple sclerosis and its animal models.** *Biochim Biophys Acta.* 2011; **1812**(2): 151–161.
[PubMed Abstract](#) | [Publisher Full Text](#)
29. Jiang H, Zhang SI, Pernis B: **Role of CD8⁺ T cells in murine experimental allergic encephalomyelitis.** *Science.* 1992; **256**(5060): 1213–1215.
[PubMed Abstract](#) | [Publisher Full Text](#)
30. Koh DR, Fung-Leung WP, Ho A, *et al.*: **Less mortality but more relapses in experimental allergic encephalomyelitis in CD8⁺ mice.** *Science.* 1992; **256**(5060): 1210–1213.
[PubMed Abstract](#) | [Publisher Full Text](#)
31. **F** Hu D, Ikizawa K, Lu L, *et al.*: **Analysis of regulatory CD8 T cells in Qa-1-deficient mice.** *Nat Immunol.* 2004; **5**(5): 516–523.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
32. **F** Tang X, Maricic I, Purohit N, *et al.*: **Regulation of immunity by a novel population of Qa-1-restricted CD8αphaαphaTCRαphabeta⁺ T cells.** *J Immunol.* 2006; **177**(11): 7645–7655.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
33. Huseby ES, Liggitt D, Brabb T, *et al.*: **A pathogenic role for myelin-specific CD8⁺ T cells in a model for multiple sclerosis.** *J Exp Med.* 2001; **194**(5): 669–676.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
34. Sun D, Whitaker JN, Huang Z, *et al.*: **Myelin antigen-specific CD8⁺ T cells are encephalitogenic and produce severe disease in C57BL/6 mice.** *J Immunol.* 2001; **166**(12): 7579–7587.
[PubMed Abstract](#) | [Publisher Full Text](#)
35. **F** Friese MA, Jakobsen KB, Friis L, *et al.*: **Opposing effects of HLA class I molecules in tuning autoreactive CD8⁺ T cells in multiple sclerosis.** *Nat Med.* 2008; **14**(11): 1227–1235.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
36. Morgan DJ, Liblau R, Scott B, *et al.*: **CD8(+) T cell-mediated spontaneous diabetes in neonatal mice.** *J Immunol.* 1996; **157**(3): 978–983.
[PubMed Abstract](#)
37. **F** Saxena A, Bauer J, Scheikl T, *et al.*: **Cutting edge: Multiple sclerosis-like lesions induced by effector CD8 T cells recognizing a sequestered antigen on oligodendrocytes.** *J Immunol.* 2008; **181**(3): 1617–1621.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
38. **F** Na SY, Cao Y, Toben C, *et al.*: **Naive CD8 T-cells initiate spontaneous autoimmunity to a sequestered model antigen of the central nervous system.** *Brain.* 2008; **131**(Pt 9): 2353–2365.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
39. **F** Sobottka B, Harrer MD, Ziegler U, *et al.*: **Collateral bystander damage by myelin-directed CD8⁺ T cells causes axonal loss.** *Am J Pathol.* 2009; **175**(3): 1160–1166.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
40. **F** Na SY, Eujen H, Göbel K, *et al.*: **Antigen-specific blockade of lethal CD8 T-cell mediated autoimmunity in a mouse model of multiple sclerosis.** *J Immunol.* 2009; **182**(10): 6569–6575.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)

41. **F** Perchellet A, Stromnes I, Pang JM, *et al.*: **CD8⁺ T cells maintain tolerance to myelin basic protein by 'epitope theft'.** *Nat Immunol.* 2004; 5(6): 606–614. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
42. **F** Ji Q, Perchellet A, Goverman JM: **Viral infection triggers central nervous system autoimmunity via activation of CD8⁺ T cells expressing dual TCRs.** *Nat Immunol.* 2010; 11(7): 628–634. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
43. Ji Q, Castelli L, Goverman JM: **MHC class I-restricted myelin epitopes are cross-presented by Tip-DCs that promote determinant spreading to CD8⁺ T cells.** *Nat Immunol.* 2013; 14(3): 254–61. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
44. **F** Hauser SL, Waubant E, Arnold DL, *et al.*: **B-cell depletion with rituximab in relapsing-remitting multiple sclerosis.** *N Engl J Med.* 2008; 358(7): 676–688. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
45. **F** Bar-Or A, Calabresi PA, Arnold D, *et al.*: **Rituximab in relapsing-remitting multiple sclerosis: a 72-week, open-label, phase I trial.** *Ann Neurol.* 2008; 63(3): 395–400. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
46. **F** Weber MS, Prod'homme T, Patarroyo JC, *et al.*: **B-cell activation influences T-cell polarization and outcome of anti-CD20 B-cell depletion in central nervous system autoimmunity.** *Ann Neurol.* 2010; 68(3): 369–383. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
47. **F** Bettelli E, Baeten D, Jäger A, *et al.*: **Myelin oligodendrocyte glycoprotein-specific T and B cells cooperate to induce a Devic-like disease in mice.** *J Clin Invest.* 2006; 116(9): 2393–2402. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
48. **F** Krishnamoorthy G, Lassmann H, Wekerle H, *et al.*: **Spontaneous opticospinal encephalomyelitis in a double-transgenic mouse model of autoimmune T cell/B cell cooperation.** *J Clin Invest.* 2006; 116(9): 2385–2392. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
49. **F** Pöllinger B, Krishnamoorthy G, Berer K, *et al.*: **Spontaneous relapsing-remitting EAE in the SJL/J mouse: MOG-reactive transgenic T cells recruit endogenous MOG-specific B cells.** *J Exp Med.* 2009; 206(6): 1303–1316. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
50. **F** Pierson ER, Stromnes IM, Goverman JM: **B cells promote induction of experimental autoimmune encephalomyelitis by facilitating reactivation of T cells in the central nervous system.** *J Immunol.* 2014; 192(3): 929–939. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
51. **F** Matsushita T, Yanaba K, Bouaziz JD, *et al.*: **Regulatory B cells inhibit EAE initiation in mice while other B cells promote disease progression.** *J Clin Invest.* 2008; 118(10): 3420–3430. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
52. **F** Fillatreau S, Sweeney CH, McGeachy MJ, *et al.*: **B cells regulate autoimmunity by provision of IL-10.** *Nat Immunol.* 2002; 3(10): 944–950. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
53. **F** Tedder TF: **B10 cells: a functionally defined regulatory B cell subset.** *J Immunol.* 2015; 194(4): 1395–1401. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
54. **Placebo-controlled multicentre randomised trial of interferon beta-1b in treatment of secondary progressive multiple sclerosis. European Study Group on interferon beta-1b in secondary progressive MS.** *Lancet.* 1998; 352(9139): 1491–1497. [PubMed Abstract](#) | [Publisher Full Text](#)
55. Kappos L, Weinshenker B, Pozzilli C, *et al.*: **Interferon beta-1b in secondary progressive MS: a combined analysis of the two trials.** *Neurology.* 2004; 63(10): 1779–1787. [PubMed Abstract](#) | [Publisher Full Text](#)
56. Panitch H, Miller A, Paty D, *et al.*: **Interferon beta-1b in secondary progressive MS: results from a 3-year controlled study.** *Neurology.* 2004; 63(10): 1788–1795. [PubMed Abstract](#) | [Publisher Full Text](#)
57. **Secondary Progressive Efficacy Clinical Trial of Recombinant Interferon-Beta-1a in MS (SPECTRIMS) Study Group: Randomized controlled trial of interferon-beta-1a in secondary progressive MS: Clinical results.** *Neurology.* 2001; 56(11): 1496–1504. [PubMed Abstract](#) | [Publisher Full Text](#)
58. Trapp BD, Nave KA: **Multiple sclerosis: an immune or neurodegenerative disorder?** *Annu Rev Neurosci.* 2008; 31: 247–269. [PubMed Abstract](#) | [Publisher Full Text](#)
59. **F** Frischer JM, Bramow S, Dal-Bianco A, *et al.*: **The relation between inflammation and neurodegeneration in multiple sclerosis brains.** *Brain.* 2009; 132(Pt 5): 1175–1189. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
60. **F** Hochmeister S, Grundtner R, Bauer J, *et al.*: **Dysferlin is a new marker for leaky brain blood vessels in multiple sclerosis.** *J Neuropathol Exp Neurol.* 2006; 65(9): 855–865. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
61. Filippi M, Rossi P, Campi A, *et al.*: **Serial contrast-enhanced MR in patients with multiple sclerosis and varying levels of disability.** *AJNR Am J Neuroradiol.* 1997; 18(8): 1549–1556. [PubMed Abstract](#) | [Publisher Full Text](#)
62. **F** Kutzelnigg A, Lucchinetti CF, Stadelmann C, *et al.*: **Cortical demyelination and diffuse white matter injury in multiple sclerosis.** *Brain.* 2005; 128(Pt 11): 2705–2712. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
63. Trapp BD, Peterson J, Ransohoff RM, *et al.*: **Axonal transection in the lesions of multiple sclerosis.** *N Engl J Med.* 1998; 338(5): 278–285. [PubMed Abstract](#) | [Publisher Full Text](#)
64. Dutta R, Trapp BD: **Relapsing and progressive forms of multiple sclerosis: insights from pathology.** *Curr Opin Neurol.* 2014; 27(3): 271–278. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
65. **F** Fischer MT, Wimmer I, Höftberger R, *et al.*: **Disease-specific molecular events in cortical multiple sclerosis lesions.** *Brain.* 2013; 136(Pt 6): 1799–1815. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
66. Lin MT, Beal MF: **Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases.** *Nature.* 2006; 443(7113): 787–795. [PubMed Abstract](#) | [Publisher Full Text](#)
67. **F** Fischer MT, Sharma R, Lim JL, *et al.*: **NADPH oxidase expression in active multiple sclerosis lesions in relation to oxidative tissue damage and mitochondrial injury.** *Brain.* 2012; 135(Pt 3): 886–899. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
68. Mahad DH, Trapp BD, Lassmann H: **Pathological mechanisms in progressive multiple sclerosis.** *Lancet Neurol.* 2015; 14(2): 183–193. [PubMed Abstract](#) | [Publisher Full Text](#)
69. **F** Geurts JJ, Calabrese M, Fisher E, *et al.*: **Measurement and clinical effect of grey matter pathology in multiple sclerosis.** *Lancet Neurol.* 2012; 11(12): 1082–1092. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
70. **F** Bø L, Vedeler CA, Nyland HI, *et al.*: **Subpial demyelination in the cerebral cortex of multiple sclerosis patients.** *J Neuropathol Exp Neurol.* 2003; 62(7): 723–732. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
71. Serafini B, Rosicarelli B, Magliozzi R, *et al.*: **Detection of ectopic B-cell follicles with germinal centers in the meninges of patients with secondary progressive multiple sclerosis.** *Brain Pathol.* 2004; 14(2): 164–174. [PubMed Abstract](#) | [Publisher Full Text](#)
72. **F** Magliozzi R, Howell O, Vora A, *et al.*: **Meningeal B-cell follicles in secondary progressive multiple sclerosis associate with early onset of disease and severe cortical pathology.** *Brain.* 2007; 130(Pt 4): 1089–1104. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
73. **F** Ransohoff RM: **Natalizumab for multiple sclerosis.** *N Engl J Med.* 2007; 356(25): 2622–2629. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
74. **F** Cohen JA, Barkhof F, Comi G, *et al.*: **Oral fingolimod or intramuscular interferon for relapsing multiple sclerosis.** *N Engl J Med.* 2010; 362(5): 402–415. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
75. **F** Kappos L, Radue EW, O'Connor P, *et al.*: **A placebo-controlled trial of oral fingolimod in relapsing multiple sclerosis.** *N Engl J Med.* 2010; 362(5): 387–401. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)

Open Peer Review

Current Peer Review Status:

Version 1

Reviewer Report 07 August 2015

<https://doi.org/10.5256/f1000research.6839.r9922>

© 2015 Segal B. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Benjamin M. Segal

Neurology Department, University of Michigan, Ann Arbor, MI, 48109, USA

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 07 August 2015

<https://doi.org/10.5256/f1000research.6839.r9926>

© 2015 Brown M. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Melissa A. Brown

Department of Microbiology-Immunology, Northwestern University Feinberg School of Medicine, Chicago, IL, 60611, USA

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 07 August 2015

<https://doi.org/10.5256/f1000research.6839.r9920>

© 2015 Fugger L. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Lars Fugger

Nuffield Department of Clinical Neurosciences, University of Oxford, Oxford, OX3 9DU, UK

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

F1000Research