RESEARCH NOTE

Evolutionary relations and population differentiation of *Acipenser gueldenstaedtii* Brandt, *Acipenser persicus* Borodin, and *Acipenser baerii* Brandt [version 1; peer review: 2 approved]

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Abstract

Russian (*Acipenser gueldenstaedtii*), Persian (*A. persicus*) and Siberian (*A. baerii*) sturgeons are closely related 'Ponto-Caspian' species. Investigation of their population structure is an important problem, the solution of which determines measures for conservation of these species. According to previous studies, 'baerii-like' mitotypes were found in the Caspian Sea among 35% of Russian sturgeon specimens, but were not found in Persian sturgeons. This confirms genetic isolation of the Persian sturgeon from the Russian sturgeon in the Caspian Sea. However, in order to clarify the relationships of these species it is necessary to analyze nuclear DNA markers. The amplified fragment length polymorphism (method) allows estimating interpopulation and interspecific genetic distances using nuclear DNA markers. In the present study, four samples were compared: Persian sturgeons from the South Caspian Sea, Russian sturgeons from the Caspian Sea and the Sea of Azov, and Siberian sturgeons from the Ob' River, which are close to these two species, but are also clearly morphologically and genetically distinct from them. For the AFLP method, eight pairs of selective primers were used. The analysis revealed that the Siberian sturgeon has formed a separate branch from the overall Persian-Russian sturgeons cluster, which was an expected result. In addition, the results showed that the Caspian Russian sturgeon is closer to the Persian sturgeon from the Caspian Sea than to the Russian Sturgeon from the Sea of Azov. The present DNA marker data confirm that despite the genetic isolation of the Persian sturgeon from the Russian sturgeon in the Caspian Sea, the Persian sturgeon is a young species.

Keywords

Russian Sturgeon, Persian Sturgeon, Siberian sturgeon, AFLP

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01 Dec 2016	view	view

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Any reports and responses or comments on the article can be found at the end of the article.

This article is included in the **Phylogenetics** collection.

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Introduction

Three closely related species, the Russian (*Acipenser gueldenstaedtii*), Persian (*A. persicus*), and Siberian (*A. baerii*) sturgeons belong to a polychromosomal group of sturgeon species ($2n = 240-260$; Vasil'ev, 1985). They form the Ponto-Caspian clade of sturgeons (Birstein & DeSalle, 1998). *A. persicus* inhabits the Caspian Sea, and *A. gueldenstaedtii* inhabits the Caspian Sea and the Azov Sea (Berg, 1961). *A. baerii* is geographically isolated from the other two species, and it inhabits Siberian Rivers. Presumably, its Ponto-Caspian ancestors migrated to Siberia (Birstein & DeSalle, 1998).

These species are closely related, which has caused some difficulties with their molecular genetic identification and clarification of their phylogenetic relations. *A. persicus* was described as a species by Borodin in 1897 (Borodin, 1897). Later, Berg called it a morphologically distinguishable subspecies of *A. gueldenstaedtii* (Berg, 1961). Following Berg researchers considered the Persian sturgeon as a subspecies of the Russian sturgeon *Acipenser gueldenstaedtii persicus* (Legeza, 1975), *Acipenser gueldenstaedtii persicus natio kurensis* (Abdurakhmanov, 1962; Legeza & Voinova, 1967). Research of the antigenic components of sturgeon blood serum proteins, carried out in 1974, revealed that the Persian sturgeon is a valid sympatric species (Lukyanenko *et al.*, 1974a; Lukyanenko *et al.*, 1974b).

The taxonomic rank of *A. persicus* is still disputed. Some researchers point to a distinct morphological differences between Russian and Persian sturgeons (Artyukhin, 2008; Vasil'eva, 2004). Others find these differences weak and point to the impossibility to distinguish single specimens of Russian, Persian and Adriatic (*A. naccarii*) sturgeons by mitochondrial DNA markers (Birstein *et al.*, 2005; Ruban *et al.*, 2008).

The Siberian sturgeon is geographically isolated from the Russian and Persian sturgeons and morphologically is easily distinguishable from them. However, approximately 30% of the Russian sturgeon specimens from the Caspian Sea have mitochondrial DNA that is similar to mitochondrial DNA of *A. baerii* (Jennekens *et al.*, 2000). It was shown that a 'baerii-like' mitotype of *A. gueldenstaedtii* is similar, but not identical, to mitochondrial DNA of *A. baerii* (Muge *et al.*, 2008). In total, 2% of Russian sturgeons in the Azov Sea also have a 'baerii-like' mitotype (Timoshkina *et al.*, 2009), whereas this has not been found in Persian sturgeons (Muge *et al.*, 2008). It is assumed that the 'baerii-like' mitochondrial DNA found in some Russian sturgeons from the Caspian Sea is a result of an introgression event during the Pleistocene glaciation (Muge *et al.*, 2008; Rastorguev *et al.*, 2013).

In order to clarify the phylogenetic relations and population structure of the species within the Ponto-Caspian sturgeon clade, some authors point out the necessity to explore nuclear DNA markers (Krieger *et al.*, 2008; Muge *et al.*, 2008). It should be noted that currently researchers have the opportunity to work with single nucleotide polymorphism (SNP) markers, which have been discovered for Ponto-Caspian sturgeons (Ogden *et al.*, 2013; Rastorguev *et al.*, 2013).

Moreover, to estimate genetic distances within the Ponto-Caspian sturgeon species group, the amplified fragment length polymorphism (AFLP) method is also applicable, as the AFLP technique allows to obtain a high number of dominant nuclear DNA markers (Congiu *et al.*, 2002).

This report presents the results of a molecular genetic study of interpopulation and interspecific genetic distances of the Ponto-Caspian sturgeon clade carried out with the AFLP method.

Materials and methods

For this research, sturgeon tissue samples (ethanol fixed fin fragments) were obtained from the Russian Federal Reference Collection of Genetic Materials (maintained by the Russian Federal Research Institute of Fisheries and Oceanography, Moscow, Russia). The sample included 24 specimens of *A. gueldenstaedtii* from the Azov Sea (catalog number GUE2906,2908-2930), 24 specimens of *A. gueldenstaedtii* from the Caspian Sea (catalog number GUE2812-2835), 24 specimens of *A. persicus* from the Southern Caspian Sea (catalog number PER0120-143) and 24 specimens of *A. baerii* from the Ob' River (catalog number BAE0325-348).

DNA was extracted and purified with the Wizard SV Genomic DNA Purification System (Promega). For genetic analysis, the AFLP method was used (Vos *et al.*, 1995). Briefly, genomic DNA was incubated with the MspI and EcoR enzyme combination (Fermentas). Next, DNA fragments were ligated with oligonucleotide adapters and used for pre-selective and selective PCR with eight combinations of fluorescent primers (Table 1):

- 1) EcoFAM_AAG - Msp_pr_AAC, 2) EcoFAM_ATT - Msp_pr_AAG, 3) EcoFAM_ACA - Msp_pr_AAT, 4) EcoFAM_AAG - Msp_pr_ACA, 5) EcoFAM_ACA - Msp_pr_ACC, 6) EcoFAM_ATT - Msp_pr_ACC, 7) EcoFAM_AAG - Msp_pr_ACT, 8) EcoFAM_AAG - Msp_pr_ATC.

Table 1. The list of AFLP primers (Syntol).

PCR	AFLP primers	5'-3' sequences of oligonucleotides
Pre-selective PCR	ERpr_A	gactgcgtaccaattcA
	Msp_pr_A	gatgagtcctgagcggA
Selective PCR	EcoFAM_AAG	FAM gactgcgtaccaattcAag
	EcoFAM_ATT	FAM gactgcgtaccaattcAtt
	EcoFAM_ACA	FAM gactgcgtaccaattcAca
	Msp_pr_AAC	gatgagtcctgagcggAac
	Msp_pr_AAG	gatgagtcctgagcggAag
	Msp_pr_AAT	gatgagtcctgagcggAat
	Msp_pr_ACA	gatgagtcctgagcggAca
	Msp_pr_ACC	gatgagtcctgagcggAcc
	Msp_pr_ACT	gatgagtcctgagcggAct
	Msp_pr_ATC	gatgagtcctgagcggAtc

Pre-selective PCR was performed for 20 cycles with the following cycle profile: a 30 sec DNA denaturation step at 94°C, a 1 min annealing step at 56°C, and a 1 min extension step at 72°C. Selective PCR was performed for 36 cycles with the following cycle profile: a 30 sec DNA denaturation step at 94°C, a 30 sec annealing step, and a 1 min extension step at 72°C. The annealing temperature in the first cycle was 65°C, was subsequently reduced each cycle by 0.7°C for the next 12 cycles, and was continued at 56°C for the remaining 23 cycles. All steps were carried out with the PTC-225 Peltier Thermal Cycler (MJ Research).

Capillary electrophoresis was carried out with the ABI Prism Genetic Analyzer 3100 (Applied Biosystems).

Analysis of the obtained AFLP-profiles was performed using Phoretix 1D Advanced v. 5.20 software (Nonlinear Dynamics). The resulting binary matrix was created for further statistical analysis with the program Tools for Population Genetic Analysis v 1.3 (TFPGA). To estimate the allele frequencies of the dominant markers, we used the approach of Lynch & Milligan (1994), which allows work with tetraploid species (Rodzen & May, 2002). With TFPGA and the unweighted-pair group method with arithmetic

means (UPGMA) method, we obtained the matrix of genetic distances (Nei, 1978) between investigated samples and constructed a dendrogram.

Results

Using eight combinations of primers, we obtained AFLP profiles (Figure 1) with 588 markers (molecular length from 100 to 380 bp). In total, 79.59% of the loci were polymorphic. A total of 4 loci were species-specific and monomorphic in the AFLP profiles of *A. baerii*. The differentiation between Russian and Persian sturgeons was observed only in the marker frequencies.

Using the TFPGA software, genetic distances (Nei, 1978) were estimated between four sturgeon samples: (1) *A. gueldenstaedtii* from the Caspian Sea; (2) *A. gueldenstaedtii* from the Azov Sea; (3) *A. baerii* from the Ob' River; and (4) *A. persicus* from the Southern Caspian Sea (Table 2). We considered the sample size, the amount of obtained markers and used unbiased statistical estimation. The UPGMA dendrogram was constructed with a bootstrap support (1000 permutations) for each node to validate the resulting topology (Figure 2).

Figure 1. AFLP profile fragments from four sturgeon samples (90-190 bp, EcoFAM_ACA - Msp_pr_ACC primer combination). (1–8) *A. gueldenstaedtii* from the Caspian Sea; (9–16) *A. gueldenstaedtii* from the Azov Sea; (17–24) *A. baerii* from the Ob' River; and (25–32) *A. persicus* from the Southern Caspian Sea.

Table 2. The matrix of genetic original Nei distances (Nei, 1978) of four sturgeon samples. (1) *A. gueldenstaedtii* from the Caspian Sea; (2) *A. gueldenstaedtii* from the Azov Sea; (3) *A. baerii* from the Ob' River; and (4) *A. persicus* from the Southern Caspian Sea.

Sample number	1	2	3	4
1	*****			
2	0.0105	*****		
3	0.0138	0.0213	*****	
4	0.0084	0.0136	0.0224	*****

Figure 2. Dendrogram depicting genetic similarity of four sturgeon samples. (1) *A. gueldenstaedtii* from the Caspian Sea; (2) *A. gueldenstaedtii* from the Azov Sea; (3) *A. baerii* from the Ob' River; and (4) *A. persicus* from the Southern Caspian Sea. Similarities were estimated based on the UPGMA method. The values refer to bootstrap values greater than 0.7.

Discussion

The AFLP method conducted in the present study revealed that the Siberian sturgeon has formed a branch that is separate from the overall Persian-Russian sturgeon cluster. The Siberian sturgeon is geographically isolated from Persian and Russian sturgeons and is morphologically easily distinguishable from them. According to the results obtained, the Caspian Russian sturgeon is closer to the Persian sturgeon from the Caspian Sea than to the Russian Sturgeon from the Sea of Azov.

The DNA marker data confirms that, despite the genetic isolation, the Persian sturgeon is a young species. Presumably, the reproductive isolation of Persian sturgeon appeared later than the event of

geographic isolation of the Black Sea-Azov and the Caspian basins. Perhaps, there is a gene flow between populations of Persian and Russian sturgeons in the Caspian Sea, which is typical for sturgeons' natural interspecific hybridization. In this case, it should be mentioned that there is no gene flow from the Russian sturgeon to the Persian sturgeon, as the Persian sturgeon is completely free from the 'baeri-like' mitotype, typical for the Russian sturgeon in the Caspian Sea (Muge *et al.*, 2008).

The results of this study show the special status of the Russian sturgeon of the Azov Sea, which is geographically and genetically isolated from the Russian sturgeon of the Caspian Sea. This differentiation was shown in previous studies with morphology, mtDNA and STR markers of the Russian sturgeon from the Black Sea-Azov and the Caspian basins (Timoshkina *et al.*, 2009). The present study has now confirmed these results using the AFLP method.

On the dendrogram, we can observe high bootstrap support values (Salemi & Vandamme, 2003). However, the obtained genetic distances are unusually small for river spawning species. This can be explained by a slower molecular evolution rate of sturgeons (Krieger & Fuerst, 2002). Further studies applying SNP and micro-satellite analysis approaches are needed in order to confirm results of this study.

Data availability

The raw data is available from Zenodo: (<https://zenodo.org/record/167463#.WC8wTtWLTcs>) DOI, 10.5281/zenodo.167463 (Sergeev, 2016).

Dataset 1 includes AFLP chromatograms (ABI Prism Genetic Analyzer 3100, Applied Biosystems). **Dataset 2** includes AFLP profiles for Phoretix 1D Advanced v. 5.20 software (Nonlinear Dynamics). **Dataset 3** includes TFPGA files (Tools for Population Genetic Analysis v 1.3) with genetic distances and trees.

Author contributions

AS carried out all work relating to this study.

Competing interests

No competing interests were disclosed.

Grant information

The author(s) declared that no grants were involved in supporting this work.

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Reviewer Report 08 December 2016

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Dmitri D. Pervouchine

Bioinformatics and Genomics Programme, Centre for Genomic Regulation (CRG), Barcelona
Institute of Science and Technology (BIST), Barcelona, Spain

The manuscript entitled "Evolutionary relations and population differentiation of *Acipenser gueldenstaedtii* Brandt, *Acipenser persicus* Borodin, and *Acipenser baerii* Brandt" by Alexey Sergeev describes the relationship between mitochondrial genotypes of four closely related sturgeon species by using the amplified fragment length polymorphism (AFPL) method. As a result, an analog of a phylogenetic tree is constructed, one in which the species paradoxically cluster by the habitat (*A. gueldenstaedtii* from the Caspian Sea is more closely related to *A. persicus* from the Southern Caspian Sea rather than to *A. gueldenstaedtii* from the Azov Sea).

Generally, the experiment is carried out at a good level and the findings are sufficiently novel. My main criticism is that it is not quite clear to what extent the AFPL method reflects the actual phylogenetic distance between species and that only the mitochondrial genome was interrogated. Generally, the reader has to get an idea of the evolutionary similarity between species by looking at the similarity of patterns in the AFPL profiles. I think the paper can be accepted for indexing if the author explains the caveats and limitations related to this method somewhere in the introduction. Otherwise the readership might be misled by the dendrogram and the phylogeny reported by the author.

Currently I have very cosmetic comments which are outlined below.

Abstract:

According to previous studies, 'baerii-like' mitotypes => mitochondrial genotypes -- the readership might be unfamiliar with this term

The amplified fragment length polymorphism (method) => The amplified fragment length polymorphism method (AFLP) -- the abbreviation must cited in the abstract next to full size sentence, since it appears later without reference

Persian sturgeons from the South Caspian Sea, Russian sturgeons from the Caspian Sea and the Sea of Azov, and Siberian sturgeons from the Ob' River, which are close to these - which these? Change 'these' to 'the latter two' - two species, but are also clearly morphologically and genetically distinct from them.

Main text has to be checked carefully with a native English speaker for word usage.

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 14 Dec 2016

Alexey Sergeev, Russian Federal Research Institute of Fisheries and Oceanography, Moscow, Russian Federation

We appreciate a lot such a detailed analysis of the article and very useful comments of Dr. Pervuchine. All suggestions will be applied to the edited version of the article. Some clarifications are needed.

1. In this study the mitochondrial genome was not investigated at all. It will be mentioned in the article. In fact, mitochondrial DNA of *A. gueldenstaedtii* has only two restriction sites which EcoRI endonuclease recognizes and about fourty sites matched to MspI. In our research we analyze those fragments of AFLP pattern which meet the following conditions: 1) the AFLP band must have length between 100 – 400 bp, 2) the amplified fragment must be produced with EcoRI selective primer (due to that only EcoRI primer has fluorescent dye), 3) the bases that followed by restriction site should be complementary to the selective bases of primers at final step of amplification. This mtDNA has only two restriction fragments which could be produced but they do not meet these conditions. Previous studies showed weak ability of mitochondrial DNA markers to perform exact species identification of individuals from this sturgeon species group and to clarify their phylogenetic relations. The main goal of this work was to analyze nuclear markers as much more informative ones. The AFLP method allows obtaining a large set of anonymous nuclear marker of the genome.
2. AFLP profiles reveal patterns of nuclear DNA markers obtained from the whole genome and reflect its polymorphism. Interrogating these profiles we can estimate similarity of the sample genomes, and statistically verify significance of AFLP pattern difference which reflect similarity of nuclear genomes. By examining the differences between the populations and computing genetic distances, and taking into account former geological events, we can make suggestions on approximate time of the population and species separation (Nei, 1972).
3. Obviously, this method has some limitations. Dominant markers are applicable for polyploid genome study but less informative than co-dominant markers (G. Guillot and Carpentier-Skandalis, 2010). We obtained large marker set from nuclear DNA but these markers are anonymous. We can't distinguish which of them are selectively

neutral and more informative. We work with them in complex which can somewhat distort the whole picture. Therefore, it's not correct to make the ultimate phylogenetic conclusions based only on this data. However, these results can be very useful in comparison with the data obtained from other approaches. It will also be mentioned in the article.

Many thanks for very useful comments. They will be very helpful for improving the manuscript.

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Competing Interests: No competing interests were disclosed.

Reviewer Report 06 December 2016

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Ekaterina V. Ponomareva

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In this article, Sergeev addressed the problem having long and complex history of discussion ¹⁻³. Molecular genetic methods (namely, AFLP in Sergeev's study) open a possibility to solve some principal and controversial questions of sturgeon species systematic relationships.

It is of particular interest also, because the problem has strong practical (and international) dimension, connected with commercial use, conservation and restoration of sturgeons. In spite of significant amount of literature on the subject, there is still a deficit of studies using genomic markers.

It should be specially mentioned, that Sergeev's study supports differentiation between Russian sturgeons (*Acipenser gueldenstaedtii*) from the Caspian Sea and the Sea of Azov, previously

revealed by using other methods⁴.

I think results obtained by Sergeev are important for resolving relationships between *Acipenser* spp.

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Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 14 Dec 2016

Alexey Sergeev, Russian Federal Research Institute of Fisheries and Oceanography, Moscow, Russian Federation

We appreciate the time and effort of Dr. Ponomareva and would like to thank her for detailed analysis of the investigated theme and high appreciation of this work.

Competing Interests: No competing interests were disclosed.

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