REVIEW

Diagnostic and therapeutic options in recurrent implantation failure [version 1; peer review: 2 approved]

Sarah Moustafa , Steven L. Young

Division of Reproductive Endocrinology, Department of Obstetrics and Gynecology, University of North Carolina at Chapel Hill, Chapel Hill, NC, USA

V1 First published: 25 Mar 2020, 9(F1000 Faculty Rev):208
<https://doi.org/10.12688/f1000research.22403.1>

Latest published: 25 Mar 2020, 9(F1000 Faculty Rev):208
<https://doi.org/10.12688/f1000research.22403.1>

Abstract

Recurrent implantation failure (RIF) is an uncommon, imprecisely defined clinical disorder characterized by failure to achieve pregnancy after repeated embryo transfers. The diverse etiologies and incomplete understanding of RIF provide significant diagnostic and therapeutic challenges to patients and providers. Careful clinical evaluation prior to assisted reproduction can uncover many treatable causes, including thyroid dysfunction, submucosal myomas, and tobacco use. The more-subtle causes often require a more-targeted assessment. Undetected, small polyps or small areas of intrauterine synechiae are relatively common and easily treated contributors to RIF. Molecular and cellular abnormalities pose a greater therapeutic challenge. Putative causes of RIF, including progesterone resistance, shifted window of receptivity, decreased integrin expression, and immunologic disturbances, should be considered in the evaluation of a patient with otherwise unexplained RIF. It may also be true that a more complex and standardized definition of RIF would be helpful in these cases. In this paper, we review the diagnostic and therapeutic approaches to RIF, with emphasis on disorders of endometrial receptivity.

Keywords

Recurrent implantation failure, Assisted reproductive technology, in vitro fertilization, Embryo implantation, implantation, pregnancy, endometrium,

Open Peer Review

Approval Status

	1	2
version 1 25 Mar 2020		

Faculty Reviews are review articles written by the prestigious Members of **Faculty Opinions**. The articles are commissioned and peer reviewed before publication to ensure that the final, published version is comprehensive and accessible. The reviewers who approved the final version are listed with their names and affiliations.

1. **Raoul Orvieto**, Chaim Sheba Medical Center at Tel Hashomer, Tel-Hashomer, Israel
2. **Hugh Taylor**, Yale University School of Medicine, New Haven, USA

Any comments on the article can be found at the end of the article.

Corresponding author: Sarah Moustafa (sarah_moustafa@med.unc.edu)

Author roles: Moustafa S: Writing – Original Draft Preparation; Young SL: Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: Steven L. Young is co-inventor of the technology on which Receptiva (BCL6) testing is based. Sarah M. Moustafa declares that she has no competing interests.

Grant information: The author(s) declared that no grants were involved in supporting this work.

Copyright: © 2020 Moustafa S and Young SL. This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Moustafa S and Young SL. **Diagnostic and therapeutic options in recurrent implantation failure [version 1; peer review: 2 approved]** F1000Research 2020, 9(F1000 Faculty Rev):208 <https://doi.org/10.12688/f1000research.22403.1>

First published: 25 Mar 2020, 9(F1000 Faculty Rev):208 <https://doi.org/10.12688/f1000research.22403.1>

Introduction

Embryo implantation is a delicately coordinated event, relying on multicomponent, bidirectional signaling between the embryo and endometrium^{1,2}. While embryonic euploidy is one of the greatest determinants of successful conception, human studies suggest that less than 60% euploid embryos result in an ongoing pregnancy³. A number of different lines of evidence demonstrate that normal endometrium is an important part of the implantation process^{4,5}. This further supports an independent and critical endometrial component for success.

In cases when a good-quality embryo fails to result in pregnancy, an endometrial cause is often suspected but difficult to ascertain in the clinical environment. When good-quality embryos repeatedly fail to implant, a condition known as recurrent implantation failure (RIF), endometrial pathologies are often present. Although there is no universally agreed upon definition, RIF is often defined as the failure to achieve clinical pregnancy after the transfer of four or more good-quality embryos. Many propose alternate cutoffs taking into account maternal age or known euploidy, with consideration for applying the definition to two or more failed embryo transfers in appropriate populations⁶. A study by Koot *et al.* surveying 118 patients with RIF, defined by failure to conceive after three embryo transfers, found that 49% of respondents ultimately achieved live birth in a 5.5-year follow up period⁷. While this represents a promising prognosis overall, approximately 50% of RIF patients may ultimately not complete their family goals. A better understanding of the underlying issues is necessary to improve outcomes.

The establishment of endometrial receptivity is primarily coordinated by estrogen and progesterone, by way of direct and indirect transcriptional and translational regulation, leading to changes in function of all endometrial cell types, including glandular and luminal epithelium, stroma, resident immune cells, and endothelium⁸. Implantation potential is directed by both embryonic factors (euploidy, expression of critical adhesion molecules, trophoblast differentiation, and adequate invasion)⁹ and endometrial receptivity.

Estrogen stimulates endometrial proliferation, with resulting thickness directly correlated to success in assisted reproduction¹⁰. Estrogen also induces an increase in progesterone receptor expression, enabling the necessary actions of progesterone for the establishment of the “window of receptivity”. These actions include inducing the production of key molecules that promote embryo attachment, such as endometrial integrins^{11,12}. Disruptions of estrogen and progesterone action have repeatedly demonstrated impacts on pregnancy rates in assisted reproduction. The duration of progesterone exposure is responsible for timing the opening (and closing) of the window of receptivity¹³. The window of receptivity itself is correlated with a molecular signature and tightly controlled inflammatory response, responsible for coordinating the attachment of an embryo, a permissive state for invasion of this semi-foreign entity, and establishment of adequate vascular supply necessary to nurture a healthy pregnancy¹⁴.

Structural components such as polyps, submucosal myomas, intrauterine synechiae, and uterine septums have demonstrated detrimental effects on successful implantation by poorly defined mechanisms, very likely involving these processes¹⁵. Generalized conditions, such as endometriosis, have demonstrated effects on various individual components of implantation. Key studies^{16–18} demonstrated endometriosis-driven progesterone resistance, disrupting events downstream of progesterone signaling. Meanwhile, other studies have highlighted the impacts of endometriosis-driven inflammatory markers disrupting critical pathways¹⁹ for decidualization²⁰, tolerance²¹, and successful implantation²².

Other systemic disruptions may have a more occult impact on the endometrium. Obesity, for instance, has been associated with altered endometrial gene expression and reduced pregnancy rates in a dose-dependent fashion^{23,24}. While some of these pathologies may be obvious in a patient with RIF, many patients do not demonstrate clear etiology on routine testing. We intend to review additional evaluations that may be indicated, specifically with regard to the endometrium, and proposed therapies in the literature.

Evaluations to consider in a patient with recurrent implantation failure

Patients undergoing *in vitro* fertilization (IVF) have ongoing ultrasound evaluation of the endometrium. As noted above, an endometrium that does not sufficiently thicken predicts a lower chance of embryo implantation. It may also suggest the possibility of additional pathology, such as intrauterine synechiae. However, many patients with RIF exhibit normal endometrial thickness and may have other occult processes implicated. Complete evaluation would require both hysteroscopy and endometrial sampling, as discussed below.

Diagnostic hysteroscopy

Most, if not all, centers utilize at least one form of uterine cavity evaluation prior to embryo transfer. Hysterosalpingogram (HSG), saline-infusion sonography (SIS), and hysteroscopy are all accepted tools for cavity assessment. While hysteroscopy is widely regarded as the gold standard, it is not often deployed as a first-line method because of higher costs and equipment needs, even if performed in an ambulatory setting. However, the incidence of uterine pathology in women undergoing IVF has been reported to be as high as 40%²⁵. Additionally, the use of hysteroscopy in patients with prior failed transfers was associated with an increase in clinical pregnancy, whether or not pathology was detected, perhaps owing to a benefit of endometrial injury and repair²⁶. One small study showed that approximately 43% of patients with a previously normal uterine cavity evaluation were found to have abnormality on hysteroscopy²⁷. While previously missed pathology may be minor in size, studies have suggested significant fecundability improvements with removal of polyps or intrauterine adhesive disease¹⁵ and that molecular changes may impair implantation, even if size or location does not²⁸. Furthermore, hysteroscopy may raise suspicion of one culprit of RIF: chronic endometritis (CE, see below). Features such as micropolyps and hyperemic

or edematous endometrium may be identified on hysteroscopy, and these may raise suspicion for otherwise subclinical CE. Targeted biopsy allows confident CE diagnosis (with caveats), and studies suggest successful identification and treatment may improve pregnancy rates²⁹.

Endometrial sampling

Targeted endometrial sampling at the time of hysteroscopy or more general sampling in the office allows the identification of subclinical or clinically suspected CE. While some studies have shown the incidence of CE is low in a general infertility population³⁰, multiple studies have demonstrated a high incidence of CE (30%) patients with RIF^{29,31}, and it must be considered in the evaluation of unexplained RIF. Cicinelli *et al.* demonstrated that adequate treatment of CE in this population resulted in significantly higher pregnancy rates than in patients who had persistent CE after therapy. Additionally, endometrial biopsy may provide value in the diagnosis of other endometrial pathology such as polyps (whether or not this is performed with hysteroscopy). Unfortunately, diagnostic criteria for endometritis have remained a subject of debate for decades, limiting the precision of this diagnosis^{32,33}. Despite the uncertainty, CE is correlated with reproductive failure, and the relatively benign treatment with antibiotics is likely beneficial to identified cases²⁹. Polyp identification on an undirected biopsy can lead to hysteroscopic removal, and a portion of the same endometrial sample can be used for specialized endometrial testing (see below). For these reasons, we strongly support the use of endometrial sampling as part of the evaluation of all RIF patients.

BCL6 testing

Increased BCL6 expression in the endometrium has been implicated in patients with unexplained or endometriosis-associated infertility^{34,35}. While the vast majority of patients with endometriosis exhibited aberrant BCL6 expression, it cannot be ruled out that abnormal expression may occur in the absence of evident endometriosis. Regardless of fertility diagnosis, over-expression of this transcriptional gene repressor has been linked to progesterone resistance by disrupting early and critical P4 signaling¹⁶. A prospective study in patients with unexplained infertility by Almquist *et al.* showed a live birth rate following IVF of 11.5% versus 58% in patients with and without elevated BCL6, respectively³⁵. Furthermore, Likes *et al.* found that treating these patients with GnRH agonist or surgical management of endometriosis lesions significantly improved pregnancy rates³⁶. As such, it is reasonable to consider BCL6 testing in patients with otherwise unexplained RIF.

Endometrial receptivity analysis and related assays

Many studies have demonstrated different genomic signatures between pre-receptive and receptive endometrium^{37,38}. Similarly, an altered endometrial transcriptome has been implicated in patients with RIF, suggesting that these patients may fail to achieve the necessary molecular signature for receptive endometrium³⁹. These approaches are based on the premise that the window of receptivity is present for a highly specific period of time defined by hormone exposures. In 2011, Diaz-Gimeno *et al.* published results of the Endometrial Receptivity Analysis

(ERA), including 238 genes, which could accurately determine endometrial dating⁴⁰. Alonso *et al.*⁴¹ utilized this analysis to show that approximately 25% of patients with RIF had an altered window of implantation (WOI), with the correct gene signature occurring before or after the expected time of 5.5 days of progesterone exposure. These results argued for personalized embryo transfer (pET) timing. While utility has not yet been fully demonstrated in a generalized population, the altered genomic signature in patients with RIF^{42,43} may be a therapeutic opportunity. The earlier works of Alonso *et al.* have been validated in a recent study by Tan *et al.*, supporting a higher incidence of an altered WOI in RIF⁴⁴. However, owing to limited size and follow up, both studies were unable to conclude clear and significant benefit from ERA and pET in RIF. Observational studies suggest a prevalence of abnormal ERA testing in about 25% of patients with RIF (as noted above) but also in about 12–15% of control groups^{41,45}. Relative abnormal test rate in the control group suggests the existence of false positive ERA testing. In the largest study examining the utility of ERA, performed in patients undergoing their first IVF cycle, the authors showed preliminary results with pregnancy rate improving in pET, but not live birth rates⁴⁶. While the premise and data derived from ERA testing are compelling, further studies are needed to understand if there is a refined clinical application that would result in consistent outcome improvements.

Currently, the ERA test is available from a single diagnostic company, but competitors are beginning to emerge and will need to be validated and compared to existing technology⁴⁷. An earlier alternative to the ERA which may be considered is immunohistochemistry evaluation of cyclin E and p27, which exhibit progesterone exposure-dependent alterations in localization⁴⁸. Head-to-head comparisons of commercially available methods have not been performed.

Management options for unexplained recurrent implantation failure

So far, we have reviewed testing for additional pathologies possibly implicated in RIF, with known management options. However, a significant portion of RIF patients will not yield definitive etiology. Many studies have examined experimental therapies in these “unexplained” RIF patients.

Diagnostic and therapeutic laparoscopy

Once a standard component of an infertility evaluation, diagnostic laparoscopy is now seldom performed for the sole purpose of diagnosing causes of infertility. Given the increased success of IVF after prolonged pituitary downregulation in women with endometriosis⁴⁹ or surgery⁵⁰, with a likely benefit to embryo implantation, the use of laparoscopy might be considered to identify and treat otherwise unrecognized endometriosis. However, many have argued that the number needed to treat is prohibitive to routine laparoscopy, with the American Society for Reproductive Medicine (ASRM) citing, “the number of laparoscopies that need to be performed to gain one additional pregnancy is actually 40”⁵¹. However, a valid argument can be made that women with significant dysmenorrhea or with abnormal BCL6³⁶ or miRNA⁵² testing have a dramatically

increased chance of endometriosis, leading to a much more favorable number needed to treat for surgical management and/or prolonged pituitary downregulation.

Intrauterine human chorionic gonadotropin

Human chorionic gonadotropin (hCG) is produced by the trophoblast and aids in embryonic invasion⁵³. The hCG receptor is highly expressed in the endometrium and has been found to coordinate cytokine secretion during the receptive period⁵⁴. As a result, instillation of intracavitary hCG immediately preceding embryo transfer has been proposed and attempted in order to improve pregnancy rates in assisted reproduction. Strug *et al.* found increased expression of estrogen and progesterone receptors in the endometrium following hCG instillation, administered after stimulation in oocyte donors, as well as increase in other targets with roles in implantation, such as C3 and NOTCH1⁵⁵. Notably, a favorable shift in ERA testing was not appreciated.

A 2018 Cochrane review of 17 randomized controlled trials (RCTs) concluded that while exogenous hCG may provide value in the transfer of cleavage-stage embryos, no benefit was seen at the blastocyst stage⁵⁶. However, the preponderance of RCTs included in the Cochrane review were studies of exogenous hCG during exclusively fresh or fresh and frozen transfer cycles. This population would have received systemic hCG 7 days prior and is not applicable to a frozen transfer cycle, wherein exposure to hCG would be limited to the study intervention and the local production of hCG by the implanting embryo. Furthermore, it is worth noting that the majority of studies did not limit the study population to patients with prior implantation failure. Interestingly, several studies that specifically examined the role of this intervention in RIF patients found significant improvement in pregnancy rates^{57,58}. A recent systematic review concluded that hCG improved clinical pregnancy rates and live birth rates while reducing miscarriage⁵⁹. These studies encompass a small number of subjects, but existing data suggest that hCG use merits further investigation in RIF patients.

Endometrial injury or “scratch”

The idea of endometrial injury to improve implantation has been used as early as 2003, with Barash *et al.* showing benefit in patients with prior failed transfers⁶⁰. The concept is based on inciting an acute inflammatory reaction, followed by repair, resulting in the release of cytokines and growth factors known to promote implantation⁶¹. Optimal timing for positive effects of the procedure was the cycle preceding transfer, with same-cycle scratch possibly detrimental to pregnancy rate⁶². However, contrary to prior studies investigating impact on day of retrieval, a more recent study did find improved pregnancy rates with endometrial injury performed during menstruation of the same cycle (64% versus 48%, $P = 0.023$)⁶³. Also, in contrast to previous studies, Tang *et al.* examined a RIF population rather than scratch effect on primary cycles. It is unclear if the difference in outcome is driven by the population or timing of endometrial scratch. The quality of overall evidence and global applicability of this method have been questioned, particularly because of the heterogeneity of RCTs^{64,65}. Potdar *et al.*⁶⁶ and Vitagliano *et al.*⁶¹ published comprehensive reviews

and meta-analyses concluding that this intervention does impart significant benefit for women with prior implantation failure. However, a large RCT of 1,364 women published in 2019 showed no advantage to endometrial injury in the generalized population or in a subgroup of patients who had two or more failed transfers⁶⁷. Discussion of this paper has yielded a variety of polarized conclusions⁶⁸. At this time, the low-risk procedure of endometrial injury via endometrial biopsy and/or hysteroscopy is useful for diagnostic purposes, while a clearer understanding of the pathophysiology of RIF and effects of endometrial injury will be necessary to better understand a possible therapeutic role and possibly refined patient selection in RIF.

Granulocyte colony-stimulating factor

Granulocyte colony-stimulating factor (G-CSF) has been proposed to improve endometrial thickness as well as treat RIF. The data on the use of G-CSF to improve endometrial thickness have been conflicting. While several groups have produced evidence supporting improved endometrial thickness and pregnancy rates^{69,70}, these findings are not uniform⁷¹ and the hypothesis that G-CSF promotes endometrial thickness and pregnancy rates in women with thin endometrium remains to be evaluated in a blinded, randomized fashion. Without further data, we interpret the evidence as not supporting the use of G-CSF to improve endometrial thickness.

The use of G-CSF for RIF in patients, independent of endometrial thickness, has the support of a single unblinded RCT that demonstrates significantly improved implantation and live birth rates⁷². In this study, 112 patients were randomized to receive subcutaneous G-CSF 1 hour prior to day 3 embryo transfer or no intervention, resulting in an adjusted odds ratio of 2.63 (1.09–6.96) for implantation rate. The findings are consistent with at least one other recent retrospective cohort study in an RIF population⁷³. While repeated demonstration in subsequent RCTs would support more widespread use of this intervention, existing evidence suggests minimal harm and potential benefit to this therapy in indicated patients with unexplained RIF.

Platelet-rich plasma

Platelet-rich plasma (PRP) is an experimental intervention, initially geared at addressing poor proliferation and suboptimal endometrial thickness by promoting tissue regeneration. Several small, observational studies have demonstrated a positive impact of intrauterine infusion of PRP on endometrial thickness and pregnancy rates^{74,75}. The largest available RCT was performed by Nazari *et al.* in 138 RIF patients and supported a possible benefit⁷⁶. Another study investigated the use of PRP in patients with normal or optimal endometrial thickness and did not note a statistically significant improvement in pregnancy rates⁷⁷. Most existing studies are limited by small numbers, lack of randomization, and suboptimal control groups. Further research is warranted, but we interpret the evidence in support of the use of PRP for RIF as too limited for generalized application.

Letrozole

As mentioned in previous sections, an important etiology to consider for RIF is altered hormone exposure or receptor

expression in the endometrium, with consequently altered expression of key molecules for receptivity, such as $\alpha v\beta 3$ integrin and leukemia inhibitory factor (LIF). Occult endometriosis is considered a possible etiology for RIF and is associated with decreased integrin expression and increased aromatase expression^{78,79}. One proposed solution to this is the use of an aromatase inhibitor such as letrozole to restore integrin expression. A retrospective study demonstrated an increase in integrin expression as well as ongoing pregnancy rate in patients who were initially integrin negative and treated with letrozole compared to those who were integrin negative and did not receive the intervention⁸⁰. These results were validated in a more recent retrospective cohort study utilizing both a GnRH agonist and letrozole, with improved live birth rates in RIF patients receiving letrozole (superior to GnRH alone)⁸¹. While RCTs are needed to further support the use of this intervention, existing basic and clinical evidence combined with low cost and no evidence of harm support a consideration of letrozole in RIF patients.

Growth hormone

Growth hormone (GH) expression has been demonstrated in secretory-phase endometrium⁸² and shown to promote the expression of critical factors for receptivity such as VEGF, LIF, and $\beta 3$ integrin subunit^{83,84}. In one RCT, Altmae *et al.* showed an improvement in endometrial thickness, pregnancy, and live birth rates in oocyte donor recipients with RIF treated with GH compared to those who were not⁸⁵. The data are encouraging, and GH is low risk (other than expense), and thus the data may prompt use in some cases of RIF. Additional studies would be helpful in better understanding the role of this intervention.

Glucocorticoids

As previously mentioned, cytokines and uterine natural killer cells have important roles in successful implantation⁸⁶, but excessive and altered inflammatory signaling has long been suspected in implantation failure and recurrent pregnancy loss. This conceptual paradigm led to widespread use of glucocorticoids based on the biologic plausibility of restoring a normal immunologic response in the endometrium to promote healthy embryo implantation. Despite this general rationale, many small RCTs in general IVF populations have shown no clinical improvements with glucocorticoid treatment. Larger reviews and meta-analyses have similarly failed to show a benefit. As a result, ASRM guidelines⁸⁷ currently recommend against the routine use of glucocorticoids to improve implantation rates. It is important to note, however, that data specific to RIF are lacking. Some data suggest that immune-modulation in patients without auto-antibodies or exaggerated cytotoxic activity may provide benefit^{88,89}. Any benefit of glucocorticoids remains unclear, and there are rare, but clinically significant, side effects, relegating their use to research or, possibly, as a last resort in select cases with clinical and histological evidence of excessive inflammation without chronic infection. Similarly, other immune-modulators, such as IVIG and intralipids, may also be considered as a last resort in patients with punitive immunologic disorders^{90,91}. However, these data are significantly limited, and treatments are expensive.

Summary

RIF is a clinical problem, often with poorly defined criteria and underlying etiology. Despite the lack of full consensus, studies strongly implicate problems affecting the endometrial cavity, many of which are amenable to treatment.

References

- Fazleabas AT, Kim JJ, Srinivasan S, *et al.*: **Implantation in the baboon: endometrial responses.** *Semin Reprod Endocrinol.* 1999; 17(3): 257–65.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Nimbkar-Joshi S, Rosario G, Katkam RR, *et al.*: **Embryo-induced alterations in the molecular phenotype of primate endometrium.** *J Reprod Immunol.* 2009; 83(1–2): 65–71.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Scott RT Jr, Ferry K, Su J, *et al.*: **Comprehensive chromosome screening is highly predictive of the reproductive potential of human embryos: a prospective, blinded, nonselection study.** *Fertil Steril.* 2012; 97(4): 870–5.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Kovacs P, Matyas S, Boda K, *et al.*: **The effect of endometrial thickness on IVF/ICSI outcome.** *Hum Reprod.* 2003; 18(11): 2337–41.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Zhao J, Zhang Q, Wang Y, *et al.*: **Endometrial pattern, thickness and growth in predicting pregnancy outcome following 3319 IVF cycle.** *Reprod Biomed Online.* 2014; 29(3): 291–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Tarlatzis BC, Bosdou JK, Kolibianakis EM: **What is recurrent implantation failure?** In: *Recurrent Implantation Failure.* 2019.
[Publisher Full Text](#)
- Koot YEM, Hviid Saxtorph M, Goddijn M, *et al.*: **What is the prognosis for a live birth after unexplained recurrent implantation failure following IVF/ICSI?** *Hum Reprod.* 2019; 34(10): 2044–52.
[PubMed Abstract](#) | [Publisher Full Text](#) | **F1000 Recommendation**
- Lessey BA, Young SL: **What exactly is endometrial receptivity?** *Fertil Steril.* 2019; 111(4): 611–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Vitiello D, Patrizio P: **Implantation and early embryonic development: implications for pregnancy.** *Semin Perinatol.* 2007; 31(4): 204–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Liu KE, Hartman M, Hartman A, *et al.*: **The impact of a thin endometrial lining on fresh and frozen-thaw IVF outcomes: an analysis of over 40 000 embryo transfers.** *Hum Reprod.* 2018; 33(10): 1883–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
- Lessey BA: **Endometrial integrins and the establishment of uterine receptivity.** *Hum Reprod.* 1998; 13 Suppl 3: 247–58; discussion 259–61.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Lessey BA, Yeh I, Castelbaum AJ, *et al.*: **Endometrial progesterone receptors and markers of uterine receptivity in the window of implantation.** *Fertil Steril.* 1996; 65(3): 477–83.
[PubMed Abstract](#)
- Prapas Y, Prapas N, Jones EE, *et al.*: **The window for embryo transfer in oocyte donation cycles depends on the duration of progesterone therapy.** *Hum Reprod.* 1998; 13(3): 720–3.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Cha J, Sun X, Dey SK: **Mechanisms of implantation: strategies for successful pregnancy.** *Nat Med.* 2012; 18(12): 1754–67.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Tulandi T, Marzal A: **Redefining reproductive surgery.** *J Minim Invasive Gynecol.*

- 2012; 19(3): 296–306.
[PubMed Abstract](#) | [Publisher Full Text](#)
16. Yoo JY, Kim TH, Fazleabas AT, *et al.*: **KRAS Activation and over-expression of SIRT1/BCL6 Contributes to the Pathogenesis of Endometriosis and Progesterone Resistance.** *Sci Rep.* 2017; 7(1): 6765.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 17. **F** Pei T, Liu C, Liu T, *et al.*: **miR-194-3p Represses the Progesterone Receptor and Decidualization in Eutopic Endometrium From Women With Endometriosis.** *Endocrinology.* 2018; 159(7): 2554–62.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 18. Bulun S, Cheng YH, Pavone M, *et al.*: **Estrogen receptor-beta, estrogen receptor-alpha, and progesterone resistance in endometriosis.** *Semin Reprod Med.* 2010; 28(1): 36–43.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 19. **F** Miller JE, Ahn SH, Monsanto SP, *et al.*: **Implications of immune dysfunction on endometriosis associated infertility.** *Oncotarget.* 2017; 8(4): 7138–7147.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 20. Inoue T, Kanzaki H, Iwai M, *et al.*: **Tumour necrosis factor alpha inhibits in-vitro decidualization of human endometrial stromal cells.** *Hum Reprod.* 1994; 9(12): 2411–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
 21. Thiruchelvam U, Wingfield M, O'Farrelly C: **Increased uNK Progenitor Cells in Women With Endometriosis and Infertility are Associated With Low Levels of Endometrial Stem Cell Factor.** *Am J Reprod Immunol.* 2016; 75(4): 493–502.
[PubMed Abstract](#) | [Publisher Full Text](#)
 22. Du H, Taylor HS: **The Role of Hox Genes in Female Reproductive Tract Development, Adult Function, and Fertility.** *Cold Spring Harb Perspect Med.* 2015; 6(1): a023002.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 23. Bellver J, Pellicer A, García-Velasco JA, *et al.*: **Obesity reduces uterine receptivity: clinical experience from 9,587 first cycles of ovum donation with normal weight donors.** *Fertil Steril.* 2013; 100(4): 1050–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
 24. Bellver J, Martínez-Conejero JA, Labarta E, *et al.*: **Endometrial gene expression in the window of implantation is altered in obese women especially in association with polycystic ovary syndrome.** *Fertil Steril.* 2011; 95(7): 2335–2341.e8.
[PubMed Abstract](#) | [Publisher Full Text](#)
 25. Doldi N, Persico P, Di Sebastiano F, *et al.*: **Pathologic findings in hysteroscopy before in vitro fertilization-embryo transfer (IVF-ET).** *Gynecol Endocrinol.* 2009; 21(4): 235–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
 26. Demiroglu A, Gurgan T: **Effect of treatment of intrauterine pathologies with office hysteroscopy in patients with recurrent IVF failure.** *Reprod Biomed Online.* 2004; 8(5): 590–4.
[PubMed Abstract](#) | [Publisher Full Text](#)
 27. Shamma FN, Lee G, Gutmann JN, *et al.*: **The role of office hysteroscopy in in vitro fertilization.** *Fertil Steril.* 1992; 58(6): 1237–9.
[PubMed Abstract](#)
 28. Rackow BW, Jorgensen E, Taylor HS: **Endometrial polyps affect uterine receptivity.** *Fertil Steril.* 2011; 95(8): 2690–2.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 29. Cicinelli E, Matteo M, Tinelli R, *et al.*: **Prevalence of chronic endometritis in repeated unexplained implantation failure and the IVF success rate after antibiotic therapy.** *Hum Reprod.* 2015; 30(2): 323–30.
[PubMed Abstract](#) | [Publisher Full Text](#)
 30. Kasius JC, Fatemi HM, Bourgain C, *et al.*: **The impact of chronic endometritis on reproductive outcome.** *Fertil Steril.* 2011; 96(6): 1451–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
 31. Johnston-MacAnanny EB, Hartnett J, Engmann LL, *et al.*: **Chronic endometritis is a frequent finding in women with recurrent implantation failure after in vitro fertilization.** *Fertil Steril.* 2010; 93(2): 437–41.
[PubMed Abstract](#) | [Publisher Full Text](#)
 32. Vicetti Miguel RD, Chivukula M, Krishnamurti U, *et al.*: **Limitations of the criteria used to diagnose histologic endometritis in epidemiologic pelvic inflammatory disease research.** *Pathol Res Pract.* 2011; 207(11): 680–5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 33. **F** Liu Y, Chen X, Huang J, *et al.*: **Comparison of the prevalence of chronic endometritis as determined by means of different diagnostic methods in women with and without reproductive failure.** *Fertil Steril.* 2018; 109(5): 832–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 34. Evans-Hoeker E, Lessey BA, Jeong JW, *et al.*: **Endometrial BCL6 Overexpression in Eutopic Endometrium of Women With Endometriosis.** *Reprod Sci.* 2016; 23(9): 1234–41.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 35. Almquist LD, Likes CE, Stone B, *et al.*: **Endometrial BCL6 testing for the prediction of in vitro fertilization outcomes: a cohort study.** *Fertil Steril.* 2017; 108(6): 1063–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 36. Likes CE, Cooper LJ, Eifird J, *et al.*: **Medical or surgical treatment before embryo transfer improves outcomes in women with abnormal endometrial BCL6 expression.** *J Assist Reprod Genet.* 2019; 36(3): 483–90.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 37. Haouzi D, Assou S, Mahmoud K, *et al.*: **Gene expression profile of human endometrial receptivity: comparison between natural and stimulated cycles for the same patients.** *Hum Reprod.* 2009; 24(6): 1436–45.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 38. Riesenwijk A, Martin J, van Os R, *et al.*: **Gene expression profiling of human endometrial receptivity on days LH+2 versus LH+7 by microarray technology.** *Mol Hum Reprod.* 2003; 9(5): 253–64.
[PubMed Abstract](#) | [Publisher Full Text](#)
 39. Tapia A, Gangi LM, Zegers-Hochschild F, *et al.*: **Differences in the endometrial transcript profile during the receptive period between women who were refractory to implantation and those who achieved pregnancy.** *Hum Reprod.* 2007; 22(3): 340–51.
[PubMed Abstract](#) | [Publisher Full Text](#)
 40. Díaz-Gimeno P, Horcajadas JA, Martínez-Conejero JA, *et al.*: **A genomic diagnostic tool for human endometrial receptivity based on the transcriptomic signature.** *Fertil Steril.* 2011; 95(1): 50–60.e15.
[PubMed Abstract](#) | [Publisher Full Text](#)
 41. Ruiz-Alonso M, Blesa D, Díaz-Gimeno P, *et al.*: **The endometrial receptivity array for diagnosis and personalized embryo transfer as a treatment for patients with repeated implantation failure.** *Fertil Steril.* 2013; 100(3): 818–24.
[PubMed Abstract](#) | [Publisher Full Text](#)
 42. **F** Macklon N: **Recurrent implantation failure is a pathology with a specific transcriptomic signature.** *Fertil Steril.* 2017; 108(1): 9–14.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 43. Koot YE, van Hooff SR, Boomsma CM, *et al.*: **An endometrial gene expression signature accurately predicts recurrent implantation failure after IVF.** *Sci Rep.* 2016; 6: 19411.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 44. Tan J, Kan A, Hitkari J, *et al.*: **The role of the endometrial receptivity array (ERA) in patients who have failed euploid embryo transfers.** *J Assist Reprod Genet.* 2018; 35(4): 683–92.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 45. Mahajan N: **Endometrial receptivity array: Clinical application.** *J Hum Reprod Sci.* 2015; 8(3): 121–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 46. Simon C, Vladimirov IK, Castillon Cortes G, *et al.*: **Prospective, randomized study of the endometrial receptivity analysis (ERA) test in the infertility work-up to guide personalized embryo transfer versus fresh transfer or deferred embryo transfer.** *Fertil Steril.* 2016; 106(3): e46–e47.
[Publisher Full Text](#)
 47. Enciso M, Carrascosa JP, Sarasa J, *et al.*: **Development of a new comprehensive and reliable endometrial receptivity map (ER Map/ER Grade) based on RT-qPCR gene expression analysis.** *Hum Reprod.* 2018; 33(2): 220–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
 48. Dubowy RL, Feinberg RF, Keefe DL, *et al.*: **Improved endometrial assessment using cyclin E and p27.** *Fertil Steril.* 2003; 80(1): 146–56.
[PubMed Abstract](#) | [Publisher Full Text](#)
 49. Surrey ES, Silverberg KM, Surrey MW, *et al.*: **Effect of prolonged gonadotropin-releasing hormone agonist therapy on the outcome of in vitro fertilization-embryo transfer in patients with endometriosis.** *Fertil Steril.* 2002; 78(4): 699–704.
[PubMed Abstract](#) | [Publisher Full Text](#)
 50. Duffy JM, Arambage K, Correa FJ, *et al.*: **Laparoscopic surgery for endometriosis.** *Cochrane Database Syst Rev.* 2014; (4): CD011031.
[PubMed Abstract](#) | [Publisher Full Text](#)
 51. Practice Committee of the American Society for Reproductive Medicine: **Endometriosis and infertility: a committee opinion.** *Fertil Steril.* 2012; 98(3): 591–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
 52. Cosar E, Mamillapalli R, Ersoy GS, *et al.*: **Serum microRNAs as diagnostic markers of endometriosis: a comprehensive array-based analysis.** *Fertil Steril.* 2016; 106(2): 402–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
 53. **F** Bashiri A, Halper KI, Orvieto R: **Recurrent Implantation Failure-update overview on etiology, diagnosis, treatment and future directions.** *Reprod Biol Endocrinol.* 2018; 16(1): 121.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 54. Srivastava A, Sengupta J, Kriplani A, *et al.*: **Profiles of cytokines secreted by isolated human endometrial cells under the influence of chorionic gonadotropin during the window of embryo implantation.** *Reprod Biol Endocrinol.* 2013; 11: 116.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 55. Strug MR, Su R, Young JE, *et al.*: **Intrauterine human chorionic gonadotropin infusion in oocyte donors promotes endometrial synchrony and induction of early decidual markers for stromal survival: a randomized clinical trial.** *Hum Reprod.* 2016; 31(7): 1552–61.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 56. **F** Craciunas L, Tsampras N, Raine-Fenning N, *et al.*: **Intrauterine administration of human chorionic gonadotropin (hCG) for subfertile women undergoing assisted reproduction.** *Cochrane Database Syst Rev.* 2018; 10(5): CD011537.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 57. **F** Huang P, Wei L, Li X: **A study of intrauterine infusion of human chorionic**

- gonadotropin (hCG) before frozen-thawed embryo transfer after two or more implantation failures. *Gynecol Endocrinol.* 2017; **33**(1): 67–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
58. **F** Xie H, Zeng H, He D, *et al.*: Effect of intrauterine perfusion of human chorionic gonadotropin before embryo transfer after two or more implantation failures: A systematic review and meta-analysis. *Eur J Obstet Gynecol Reprod Biol.* 2019; **243**: 133–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 59. **F** Gao M, Jiang X, Li B, *et al.*: Intrauterine injection of human chorionic gonadotropin before embryo transfer can improve *in vitro* fertilization-embryo transfer outcomes: a meta-analysis of randomized controlled trials. *Fertil Steril.* 2019; **112**(1): 89–97.e1.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 60. Barash A, Dekel N, Fieldust S, *et al.*: Local injury to the endometrium doubles the incidence of successful pregnancies in patients undergoing *in vitro* fertilization. *Fertil Steril.* 2003; **79**(6): 1317–22.
[PubMed Abstract](#) | [Publisher Full Text](#)
 61. **F** Vitagliano A, Di Spiezo Sardo A, Saccone G, *et al.*: Endometrial scratch injury for women with one or more previous failed embryo transfers: A systematic review and meta-analysis of randomized controlled trials. *Fertil Steril.* 2018; **110**(4): 687–702.e2.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 62. Karimzade MA, Oskoui H, Ahmadi S, *et al.*: Local injury to the endometrium on the day of oocyte retrieval has a negative impact on implantation in assisted reproductive cycles: a randomized controlled trial. *Arch Gynecol Obstet.* 2010; **281**(3): 499–503.
[PubMed Abstract](#) | [Publisher Full Text](#)
 63. **F** Tang Z, Hong M, He F, *et al.*: Effect of endometrial injury during menstruation on clinical outcomes in frozen-thawed embryo transfer cycles: A randomized control trial. *J Obstet Gynaecol Res.* 2020; **46**(3): 451–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 64. Simón C, Bellver J: Scratching beneath 'The Scratching Case': Systematic reviews and meta-analyses, the back door for evidence-based medicine. *Hum Reprod.* 2014; **29**(8): 1618–21.
[PubMed Abstract](#) | [Publisher Full Text](#)
 65. Nastri CO, Lensen SF, Gibreel A, *et al.*: Endometrial injury in women undergoing assisted reproductive techniques. *Cochrane Database Syst Rev.* 2015; (3): CD009517.
[PubMed Abstract](#) | [Publisher Full Text](#)
 66. Potdar N, Gelbaya T, Nardo LG: Endometrial injury to overcome recurrent embryo implantation failure: a systematic review and meta-analysis. *Reprod Biomed Online.* 2012; **25**(6): 561–71.
[PubMed Abstract](#) | [Publisher Full Text](#)
 67. **F** Lensen S, Osavlyuk D, Armstrong S, *et al.*: A Randomized Trial of Endometrial Scratching before In Vitro Fertilization. *N Engl J Med.* 2019; **380**(4): 325–34.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 68. Lensen S, Venetis C, Ng EHY, *et al.*: Should we stop offering endometrial scratching prior to *in vitro* fertilization? *Fertil Steril.* 2019; **111**(6): 1094–101.
[PubMed Abstract](#) | [Publisher Full Text](#)
 69. **F** Gleicher N, Kim A, Michaeli T, *et al.*: A pilot cohort study of granulocyte colony-stimulating factor in the treatment of unresponsive thin endometrium resistant to standard therapies. *Hum Reprod.* 2013; **28**(1): 172–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 70. Xu B, Zhang Q, Hao J, *et al.*: Two protocols to treat thin endometrium with granulocyte colony-stimulating factor during frozen embryo transfer cycles. *Reprod Biomed Online.* 2015; **30**(4): 349–58.
[PubMed Abstract](#) | [Publisher Full Text](#)
 71. **F** Barad DH, Yu Y, Kushnir VA, *et al.*: A randomized clinical trial of endometrial perfusion with granulocyte colony-stimulating factor in *in vitro* fertilization cycles: impact on endometrial thickness and clinical pregnancy rates. *Fertil Steril.* 2014; **101**(3): 710–5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 72. Aleyasin A, Abediasl Z, Nazari A, *et al.*: Granulocyte colony-stimulating factor in repeated IVF failure, a randomized trial. *Reproduction.* 2016; **151**(6): 637–42.
[PubMed Abstract](#) | [Publisher Full Text](#)
 73. Zeyneloglu HB, Tohma YA, Onalan G, *et al.*: Granulocyte colony-stimulating factor for intracytoplasmic sperm injection patients with repeated implantation failure: which route is best? *J Obstet Gynaecol.* 2019; 1–5.
[PubMed Abstract](#) | [Publisher Full Text](#)
 74. Kim H, Shin JE, Koo HS, *et al.*: Effect of Autologous Platelet-Rich Plasma Treatment on Refractory Thin Endometrium During the Frozen Embryo Transfer Cycle: A Pilot Study. *Front Endocrinol (Lausanne).* 2019; **10**: 61.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 75. Coksuer H, Akdemir Y, Ulas Barut M: Improved *in vitro* fertilization success and pregnancy outcome with autologous platelet-rich plasma treatment in unexplained infertility patients that had repeated implantation failure history. *Gynecol Endocrinol.* 2019; **35**(9): 815–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
 76. Nazari L, Salehpour S, Hosseini MS, *et al.*: The effects of autologous platelet-rich plasma in repeated implantation failure: a randomized controlled trial. *Hum Fertil (Camb).* 2019; **8**: 1–5.
[PubMed Abstract](#) | [Publisher Full Text](#)
 77. Aghajanzadeh F, Esmailzadeh S, Basirat Z, *et al.*: Using autologous intrauterine platelet-rich plasma to improve the reproductive outcomes of women with recurrent implantation failure. *JBRA Assist Reprod.* 2020; **24**(1): 30–3.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 78. Lessey BA, Castelbaum AJ, Sawin SW, *et al.*: Aberrant integrin expression in the endometrium of women with endometriosis. *J Clin Endocrinol Metab.* 1994; **79**(2): 643–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
 79. Noble LS, Simpson ER, Johns A, *et al.*: Aromatase expression in endometriosis. *J Clin Endocrinol Metab.* 1996; **81**(1): 174–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
 80. Miller PB, Parnell BA, Bushnell G, *et al.*: Endometrial receptivity defects during IVF cycles with and without letrozole. *Hum Reprod.* 2012; **27**(3): 881–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 81. **F** Steiner N, Shrem G, Tannus S, *et al.*: Effect of GnRH agonist and letrozole treatment in women with recurrent implantation failure. *Fertil Steril.* 2019; **112**(1): 98–104.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 82. Sbracia M, Scarpellini F, Poverini R, *et al.*: Immunohistochemical localization of the growth hormone in human endometrium and decidua. *Am J Reprod Immunol.* 2004; **51**(2): 112–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
 83. **F** Altmäe S, Aghajanova L: Growth Hormone and Endometrial Receptivity. *Front Endocrinol (Lausanne).* 2019; **10**: 653.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 84. **F** Cui N, Li AM, Luo ZY, *et al.*: Effects of growth hormone on pregnancy rates of patients with thin endometrium. *J Endocrinol Invest.* 2019; **42**(1): 27–35.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 85. **F** Altmäe S, Mendoza-Tesarik R, Mendoza C, *et al.*: Effect of Growth Hormone on Uterine Receptivity in Women With Repeated Implantation Failure in an Oocyte Donation Program: A Randomized Controlled Trial. *J Endocr Soc.* 2018; **2**(1): 96–105.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
 86. Lédée-Bataille N, Dubanchet S, Coulomb-L'hermine A, *et al.*: A new role for natural killer cells, interleukin (IL)-12, and IL-18 in repeated implantation failure after *in vitro* fertilization. *Fertil Steril.* 2004; **81**(1): 59–65.
[PubMed Abstract](#) | [Publisher Full Text](#)
 87. **F** Practice Committee of the American Society for Reproductive Medicine: The role of immunotherapy in *in vitro* fertilization: a guideline. *Fertil Steril.* 2018; **110**(3): 387–400.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
 88. Litwicka K, Arrivi C, Varricchio MT, *et al.*: In women with thyroid autoimmunity, does low-dose prednisolone administration, compared with no adjuvant therapy, improve *in vitro* fertilization clinical results? *J Obstet Gynaecol Res.* 2015; **41**(5): 722–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
 89. Taniguchi F: Results of prednisolone given to improve the outcome of *in vitro* fertilization-embryo transfer in women with antinuclear antibodies. *J Reprod Med.* 2005; **50**(6): 383–8.
[PubMed Abstract](#)
 90. Elram T, Simon A, Israel S, *et al.*: Treatment of recurrent IVF failure and human leukocyte antigen similarity by intravenous immunoglobulin. *Reprod Biomed Online.* 2005; **11**(6): 745–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
 91. Shreeve N, Sadek K: Intralipid therapy for recurrent implantation failure: new hope or false dawn? *J Reprod Immunol.* 2012; **93**(1): 38–40.
[PubMed Abstract](#) | [Publisher Full Text](#)

Open Peer Review

Current Peer Review Status:

Editorial Note on the Review Process

Faculty Reviews are review articles written by the prestigious Members of **Faculty Opinions**. The articles are commissioned and peer reviewed before publication to ensure that the final, published version is comprehensive and accessible. The reviewers who approved the final version are listed with their names and affiliations.

The reviewers who approved this article are:

Version 1

1. **Hugh Taylor**

Department of Obstetrics, Gynecology & Reproductive Sciences, Yale University School of Medicine, New Haven, CT, USA

Competing Interests: No competing interests were disclosed.

2. **Raoul Orvieto**

Department for Obstetrics and Gynecology, Chaim Sheba Medical Center at Tel Hashomer, Tel-Hashomer, 52621, Israel

Competing Interests: No competing interests were disclosed.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

F1000Research