RESEARCH ARTICLE

REVISED Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with difference sources of activated charcoal [version 2; peer review: 2 approved]

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Abstract

Background: The giant trevally, *Caranx ignobilis*, is a commercially important marine fish in Indonesia. This species was initially cultured in Aceh Province. Previous reports showed that charcoal has a positive effect on survival and feed utilization of the giant trevally. However, the effects of adding charcoal to the diet on gut and intestine biometrics has, to our knowledge, never been described.

Methods: Four activated charcoal sources were tested in this study using a completely randomized experimental design; coconut shell charcoal, mangrove wood charcoal, rice husk charcoal, and kernel palm shell charcoal. All treatments were performed with four replications. Juvenile giant trevally (average body weight, 16.52 ± 3.12 g; and average total length, 10.26 ± 0.64 cm) were stocked into the experimental tank at a density of 15 fish per tank. The fish were fed an experimental diet twice daily at 7 AM and 5 PM *ad satiation* for 42 days.

Results: Analysis of variance showed that adding charcoal to the diet had significant effects on the length and width of the foveola gastrica and villous intestine ($P < 0.05$). The greatest length and width of the foveola gastrica was recorded in fish fed an experimental diet of rice husk charcoal with average values of 311.811 ± 9.869 μ m and 241.786 ± 10.394 μ m, respectively. The greatest length of intestinal villous was found in fish fed the mangrove wood charcoal diet, with a value of 135.012 ± 5.147 μ m, but this length was not significantly different to

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that in fish fed rice charcoal and kernel palm shell charcoal. However, the greatest width of intestinal villous was recorded in fish fed the control diet (without charcoal; $P < 0.05$).

Conclusion: The optimal sizes of the foveola gastrica and villous intestine were found in fish fed an experimental diet with rice husk charcoal.

Keywords

Foveola gastrica, villous intestine, coconut shell, mangrove wood, rice husk, and kernel palm shell

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REVISED Amendments from Version 1

In response to comments from the reviewers, the following is our amendments:

Abstract

- We changed: 'This species was...' to be 'The species was...'
- Four activated charcoal sources were tested in this study not include control, in the Methods has been explained with five different charcoal sources including control (no active charcoal source)
- All mistake numbers in the Abstract have been adjusted based on reviewer comments

Methods and Experimental

- All the mistake numbers in the Methods have been adjusted based on the reviewer comments including type mistake in the Tables and Figures

Discussion

- Some sentences have been modified to make easy to understand such as "Hence, a longer *foveola gastrica* and larger intestinal villi were able to provide more nutrients to be absorbed due to a larger surface area of digestive organs" and also the contain amount of rice husk charcoal has been revised. Some words have been added to make easier to understand.

Any further responses from the reviewers can be found at the end of the article

Introduction

Trevally fish are a commercially important group of marine fish in the family Carangidae. A total of 146 species of trevally have been recorded worldwide¹. These fish are distributed in tropical, subtropical, and temperate waters²⁻⁷. In Indonesia, trevally fish are found in the Aceh waters^{8,9}, East Borneo¹⁰, Papua and Wester Nusa Tenggara^{11,12}, and Java¹³.

Giant trevally, *Caranx ignobilis*, is among the most popular trevally fish in Indonesia. The population of this species has declined over the years due to overfishing^{7,14-16}. Culture of this fish has been initiated in Aceh Province, Indonesia. However, farmers are faced with a feeding obstacle. Giant trevally in culture systems are currently fed waste fish and a commercial diet (Hi-Pro-Vite, Central Proteina Prima Company). The commercial diet is costly and difficult to obtain in remote areas, and the waste fish supply is very seasonal. Trash fish are limited in nutrients, particularly the essential amino acid composition¹⁷. Therefore, it is crucial to formulate a diet for giant trevally using local raw materials with higher protein, that is inexpensive, easy to find, and digestible.

Activated charcoal is commonly added to the diet to increase digestibility and trigger growth in fish. For example, Jahan *et al.*¹⁸ successfully used activated charcoal to increase the digestibility and growth performance of river catfish, *Pangasiasodon* sp. Other researchers have used charcoal in the diets of fish species, such as Nile tilapia, *Oreochromis niloticus*¹⁹⁻²¹, tiger pufferfish, *Takifugu rubripes*²², Japanese flounder, *Paralichthys olivaceus*²³, African catfish, *Clarias gariepinus*^{24,25}, gilthead

seabream, *Sparus aurata*²⁶, and sturgeon, *Huso huso*²⁷. Firdus *et al.*²⁸ added rice husk charcoal to the diet of giant trevally. However, the effect of charcoal on the morphology of the gut and intestine has not been reported.

Organogenesis of the digestive system occurs as fish age, and this process is strongly dependent on the quantity and quality of food²⁹⁻³², which is related to the development of mucosal cells, amplification of apical plasma membranes, and formation of the foveola gastrica and intestinal villi^{33,34}. It has been hypothesized that adding activated charcoal to the diet triggers the digestive organogenesis system process^{35,36}. In this study, we tested four charcoal sources in the diet to evaluate the morphology of the gut and intestine of giant trevally. Information on the gut and intestinal morphology is important to understand the absorption mechanism of nutrients from the diet.

Methods**Time and site**

The study was conducted at the Center for Brackish Water Aquaculture, Ujung Batee, Aceh, Indonesia from February to July 2018. The activated charcoal was characterized at the Integrated Laboratory of Calibration, Universitas Gajah Mada, Yogyakarta, Indonesia. Histological samples were prepared at the Laboratory of Histology, Faculty of Mathematics and Natural Sciences, Universitas Syiah Kuala, Banda Aceh, Indonesia.

Experimental design

A completely randomized experimental design with five treatments consisting of control and four different charcoal sources was used in this study. The experimental groups were: (A) the experimental diet without charcoal, (B) the experimental diet with 2% charcoal from coconut shell, (C) the experimental diet with 2% charcoal from mangrove wood, (D) the experimental diet with 2% charcoal from rice husk, and (E) the experimental diet with 2% charcoal from kernel palm shell. All treatments were performed with four replications.

Experimental fish

A total of 300 giant trevally juveniles of mixed sex (average body weight, 16.52 ± 3.12 g; total length, 10.28 ± 0.64 cm) were purchased from a local farmer in Lancang Barat Village, Aceh Utara District, Aceh, Indonesia. The fish were acclimatized in ponds (ponds size 2 m x 1.8 m and temperature of around 29°C) at the Center for Brackish Water Aquaculture, Ujung Batee for 2 weeks. The fish were fed an experimental diet containing 50% crude protein twice daily at 7 AM and 5 PM at 3% of body weight per day (Table 1).

Charcoal preparation and activation

The raw coconut shells, mangrove wood, rice husks, and kernel palm shells were chopped and ground. Approximately 500 g of the ground materials were placed on aluminum foil and heated in a furnace at 400°C for 1 hour. Nitrogen gas was flowed into the furnace to remove the oxygen. Then, the temperature was decreased to 30°C gradually and held for 1 hour. After 1 hour, the charcoal was removed from the furnace, sieved through a No. 40 mesh, and held in a jar before

Table 1. The composition of raw materials in the experimental diet (g kg⁻¹) with 50% crude protein.

Raw materials	Crude protein (%)	Composition (g kg ⁻¹)	
		Diet without charcoal (Diet A, Control)	Diet with charcoal (Diet B, C, D, E)
Ebi-shrimp meal	58.80	50	50
Fish meal	59.00	660	660
Rice flour	7.26	180	160
Soybean meal	45.06	20	20
Bloodmeal	71.00	20	20
Corn flour	6.48	10	10
Coconut oil	0	5	5
CaCO ₃	0	5	5
Isoleucine	100	10	10
L-Tryptophan	100	17.5	17.5
DL-Methionine	100	17.5	17.5
Mineral mix	0	5	5
Active charcoal	0	-	20
Total material		1000	1000
Total crude protein		50%	50%

Note: (A) diet without charcoal, (B) diet with charcoal from coconut shells, (C) diet with charcoal from mangrove wood, (D) diet with charcoal from rice husks, (E) diet with charcoal from kernel palm shells.

activating. A total of 100 g of sieved charcoal was taken and mixed with 400 ml of 0.2 M citric acid. The solution was stirred for 24 hours. After 24 hours, the solution was filtered through filter paper. The filtered charcoal was washed with distilled water and dried in an oven at 110°C for 24 hours.

Diet preparation

The experimental diet was formulated from both plant and animal-based protein sources, such as Ebi-shrimp meal, fish meal, blood meal, soybean meal, rice flour, and corn flour. All raw materials were subjected to a proximate analysis before use in the formulation. Three types of amino acids i.e. isoleucine, L-tryptophan, and DL-methionine were also added (Table 1). A total of 2% of the tested charcoal sources was added to the formulation (Table 1). The formulated diets were subjected to a proximate analysis before use in the experiment.

Stocking and feeding

The fish was captured randomly, measured for body weight and total length, and then distributed into 20 plastic containers (48 × 43 × 70 cm) at a stocking density of 15 fish per container. The water volume in the container was 75 L. The fish were fed an experimental diet twice daily at 7 AM and 5 PM to satiation for 42 days.

Histological sample preparation

Gastric and intestinal samples were collected at the end of the study. Three fish from each treatment were taken randomly from the experimental tanks. The fish were anesthetized with 30 mg L⁻¹ clove oil³⁷, and the abdomen of the fish was gently dissected following the procedure of Purushothaman *et al.*³⁸. The stomach and intestines were removed with scalpel scissors and preserved in 4% formalin for 1 week. Histological sampling was carried using the paraffin method based on Osman and Caceci³⁹. The samples were dehydrated through an alcohol series and cleared in xylol. Subsequently, the gut and intestine samples were embedded in paraffin. The paraffin block was sectioned to 6 µm, and the sections were stained with hematoxylin and eosin. The size (height and width) of villi was determined using a binocular microscope (Zeiss Primo Star, Carl Zeiss Suzhou Co., Ltd., Suzhou, China) which was connected to a CCD camera and computer monitor¹⁹. All efforts were made to lessen harm to the animals by complying to the guidelines of ethics animal use in research of Syiah Kuala University.

Data analysis

The qualitative gut and intestinal morphology data were subjected to one-way analysis of variance followed by Duncan's multiple range test. The analysis was performed using SPSS

ver. 18.0 software. The qualitative (histological) gut and intestinal data were analyzed descriptively. A P -value < 0.05 was considered significant.

Results

Adding activated charcoal to the diet significantly affected the length and width of the foveola gastrica and intestinal villi ($P < 0.05$). In general, fish fed the activated charcoal diets produced better results than those not fed the charcoal (Figure 1 and Figure 2). The best foveola gastrica morphology was obtained with the rice husk charcoal and the mean length and width of the foveola gastrica were 311.811 μm and 241.786 μm , respectively; followed by coconut shell charcoal (257.040 μm and 183.816 μm), kernel palm charcoal (229.969 μm and 169.131 μm), and mangrove wood charcoal (229.595 μm and 166.509 μm).

The greatest length of the villous intestine was recorded in fish fed a diet with activated charcoal than those not fed the activated charcoal (Figure 3). The greatest growth of intestinal villi was determined in the mangrove active charcoal (mean, 135.012 μm) group, but this value was not significantly different from the rice husk or kernel palm shell charcoals (Figure 4). However, the greatest intestinal villi width was obtained in the treatment without activated charcoal (38.341 μm), and this value was significantly different from the other treatments.

Raw biometric data, in addition to unprocessed imaged, are available as *Underlying data*^{40–42}.

Discussion

The results show that adding activated charcoal to the diet of *C. ignobilis* significantly affected foveola gastrica and intestinal villi biometrics. According to Pirarat *et al.*¹⁹, activated charcoal plays a significant role stimulating the development of epithelial cells of the digestive organs. Activated charcoal in the diet functions as a decontaminating agent to eliminate pathogenic organisms and toxic compounds, such as mycotoxins²⁰. Hence, a longer foveola gastrica and larger intestinal villi were able to provide more nutrients to be absorbed due to a larger surface area of digestive organs⁴³. Optimal development of the alimentary tract was recorded in giant trevally juveniles fed the experimental diet containing rice husk charcoal. This was presumably due to the high hemicellulose, cellulose, and lignin contents in the rice husk charcoal. A previous report indicated that rice husk charcoal contains 29.3% hemicellulose, 34.4% cellulose, and 19.2% lignin⁴⁴, while mangrove wood charcoal has 30% hemicellulose, 36% cellulose, and 28% lignin⁴⁵, coconut shell charcoal has 19.27% hemicellulose, 33.61% cellulose, and 36.51% lignin⁴⁶, and kernel palm shell charcoal has 26.27% cellulose, 12.61% hemicellulose, and 42.96% lignin⁴⁷. Maria and Banu⁴⁸ and Jamilatun *et al.*⁴⁹ reported that the concentration and quality of charcoal depend on the composition of hemicellulose, cellulose, and lignin. The quality of the activated charcoal is higher

Figure 1. The average of length and width of the foveola gastrica. (A) Diet without charcoal, (B) diet with coconut shell charcoal, (C) diet with mangrove wood charcoal, (D) diet with rice husk charcoal, (E) diet with kernel palm shell charcoal.

Figure 2. Histology of *foveola gastrica* from a juvenile giant trevally. (A) Diet without charcoal, (B) diet with coconut shell charcoal, (C) diet with mangrove wood charcoal, (D) diet with rice husk charcoal, (E) diet with kernel palm shell charcoal. M, tunica mucosa; SM, tunica submucosa; Mc, tunica muscularis; Le, lamina epithelialis; Lp, lamina propria; m, muscle; Lm, longitudinal muscle fibers; Cm, circular muscle fibers (Cm).

Figure 3. The average length and width of intestine villi from juvenile giant trevally. (A) Diet without charcoal, (B) diet with coconut shell charcoal, (C) diet with mangrove wood charcoal, (D) diet with rice husk charcoal, (E) diet with kernel palm shell charcoal.

Figure 4. Histology of intestinal villi from a giant trevally juvenile. (A) Diet without charcoal, (B) diet with coconut shell charcoal, (C) diet with mangrove wood charcoal, (D) diet with rice husk charcoal, (E) diet with kernel palm shell charcoal. M, tunica mucosa; SM, tunica submucosa; Mc, tunica muscularis.

when these three components increase. According to Jasman⁵⁰, rice husk contains 85–95% activated charcoal, while mangrove wood has 76% activated charcoal⁵¹, kernel palm shell 65% activated charcoal⁴⁷, and coconut shell has 60% activated charcoal⁴⁶.

The microscopic observations showed that the intestinal villi of the fish fed the diet with activated rice husk charcoal had a more pointed shape compared to other treatments, in which the villi tended to be round and blunt. According to Guo *et al.*⁵², blunt or rounded villi probably occur due to inflammation in the intestinal mucosa, which is characterized by infiltration of neutrophils into the lamina propria. An increase of intestinal villus size is related to nutrient absorption capacity. According to Nafis *et al.*⁵³, long mucosal folds increase nutrient absorption and reduce food flow movement due to reduced peristaltic contractions, which provides sufficient time to optimally absorb nutrients. The increase in intestinal villi size is strongly related to the activities of digestive enzymes, such as lactase, sucrase, alkaline phosphatase, and disaccharidase^{54–57}.

The morphology of the intestinal villi of fish fed a diet without activated charcoal was wider and shorter than that of fish fed the diets with activated charcoal. This was probably due to impaired intestinal mucosal integrity, causing interference in nutrient absorption. According to Choct⁵⁸, shortening of the

intestinal villi is related to the accumulation of intestinal pathogenic bacteria, resulting in increased susceptibility to infection in the intestinal mucosal layer. This causes the digestive organs to form more secretory cells than absorbent cells, which reduces nutrient uptake^{59,60}. The active charcoal likely acts as an adsorbent of metabolic pathogens in the intestine in the form of endotoxins and ammonia, therefore, it was able to improve intestinal function⁶¹.

Conclusions

The application of activated charcoal in the diet significantly affected the length and width of the foveola gastrica and intestinal villi of giant trevally, *C. ignobilis*. The optimal biometrics of the foveola gastrica and intestinal villi were observed in fish fed the experimental diet with activated rice husk charcoal.

Data availability

Figshare: Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with difference sources of activated charcoal. <https://doi.org/10.6084/m9.figshare.12203525.v2>⁴⁰.

This project contains the following underlying data:

- DATA BIOMETRIC GUT OF GIANT TREVALLY *Caranx ignobilis*_Edited (XLSX). (Raw biometric data for the foveola gastrica of all fish examined in this study.)

- DATA BIOMETRIC OF INTESTINE OF GIANT TREVALLY *Caranx ignobilis*_edited (XLSX). (Raw biometric data for the intestinal villi of all fish examined in this study.)

Figshare: Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with difference sources of activated charcoal. <https://doi.org/10.6084/m9.figshare.12301124.v2>⁴¹.

This project contains uncropped, unprocessed images of the intestinal villi of giant trevally.

Figshare: Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with difference

sources of activated charcoal. <https://doi.org/10.6084/m9.figshare.12269606.v2>⁴².

This project contains uncropped, unprocessed images of the foveola gastrica of the giant trevally.

Data are available under the terms of the [Creative Commons Attribution 4.0 International license](#) (CC-BY 4.0).

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References

1. Fishbase: **Family Carangidae - Jacks and pompanos**. 2020. (Accessed on 9 April 2020). [Reference Source](#)
2. Wetherbee BM, Holland KN, Meyer CG, *et al.*: **Use of a marine reserve in Kaneohe Bay, Hawaii by the giant trevally, *Caranx ignobilis***. *Fish Res.* 2004; **67**(3): 253–263. [Publisher Full Text](#)
3. Ho JS, Lin CL: **Three species of *Caligus* Müller, 1785 (Copepoda: Caligidae) parasitic on *Caranx* spp. (Teleostei: Carangidae) off Taiwan**. *Syst Parasitol.* 2007; **68**(1): 33–43. [PubMed Abstract](#) | [Publisher Full Text](#)
4. Meyer CG, Holland KN, Papastamatiou YP: **Seasonal and diel movements of giant trevally *Caranx ignobilis* at remote Hawaiian atolls: implications for the design of marine protected areas**. *Mar Ecol Prog Ser.* 2007; **333**: 13–25. [Publisher Full Text](#)
5. Amarasinghe US, Wickramaratne IU, Wijeyaratne MJS: **Hook selectivity of giant trevally (*Caranx ignobilis*) and nakedbreast trevally (*Carangoides gymnotethus*) (Carangidae) caught in the hook-and-line fishery off Negombo, Sri Lanka**. *J Lanka J Aquat Sci.* 2011; **16**: 11–26. [Publisher Full Text](#)
6. Acuña-Marrero D, Salinas-De-León P: **New record of two Indo-Pacific reef fish, *Caranx ignobilis* and *Naso annulatus*, from the Galapagos Islands**. *Mar Biodivers Rec.* 2013; **6**: 1–5. [Publisher Full Text](#)
7. Neethiselvan N, Karthy A, Mol CB: **Gillnet selectivity on the yellow fin Trevally (*Caranx ignobilis*, Forsskal, 1775) along Thoothukudi coast, Southeast coast of India**. *J Exp Zool India.* 2015; **18**(1): 29–37. [Reference Source](#)
8. Timorya Y, Abdullah A, Batubara AS, *et al.*: **Conservation and economic status fishes in the Krueng Sabee River, Aceh Jaya District, Aceh Province, Indonesia**. *IOP Conf Ser: Earth Environ Sci.* 2018; **216**: 012044. [Publisher Full Text](#)
9. Batubara AS, Muchlisin ZA, Thamren MY, *et al.*: **Check list of marine fishes from Simeulue Island waters, Aceh Province, Indonesia**. *Aceh J Anim Sci.* 2017; **2**(2): 77–84. [Publisher Full Text](#)
10. Abdusysyahid S, Anggoro S, Bambang AN: **The distribution of capture fisheries based small pelagic-mackerel fish species in Balikpapan waters, East Kalimantan**. *Int J Eng Sci.* 2014; **6**(2): 149–153. [Publisher Full Text](#)
11. Purba GY, Haryono E, Manan J, *et al.*: **Jellyfish Lakes at Misool Islands, Raja Ampat, West Papua, Indonesia**. *Biodiversitas.* 2018; **19**: 172–182. [Publisher Full Text](#)
12. Wahyudewantoro G: **The fish diversity of mangrove waters in Lombok Island, West Nusa Tenggara, Indonesia**. *Biodiversitas.* 2018; **19**: 71–76. [Publisher Full Text](#)
13. Piranti AS, Setyaningrum N, Retna UD, *et al.*: **Fish conservation status in eastern part of segara anak-anak Cilacap Indonesia**. *IOP Conf Ser: Earth Environ Sci.* 2019; **406**: 012029. [Publisher Full Text](#)
14. Abdussamad EM, Kasim HM, Balasubramanian TS: **Distribution, biology and behaviour of the giant trevally, *Caranx ignobilis*-a candidate species for mariculture**. *Bangladesh J Fish Res.* 2008; **12**: 89–94. [Reference Source](#)
15. Santos SR, Xiang Y, Tagawa AW: **Population structure and comparative phylogeography of jack species (*Caranx ignobilis* and *C. melampygus*) in the high Hawaiian Islands**. *J Hered.* 2011; **102**(1): 47–54. [PubMed Abstract](#) | [Publisher Full Text](#)
16. Daly R, Filmlalter JD, Daly CA, *et al.*: **Acoustic telemetry reveals multi-seasonal spatiotemporal dynamics of a giant trevally *Caranx ignobilis* aggregation**. *Mar Ecol Prog Ser.* 2019; **621**: 185–197. [Publisher Full Text](#)
17. Muhammadar AA, Mazlan AG, Samat A, *et al.*: **Crude protein and amino acids content in some common feeds of tiger grouper (*Epinephelus fuscoguttatus*) juvenile**. *AACL Bioflux.* 2011; **4**(4): 449–504. [Reference Source](#)
18. Jahan R, Abdul MQ, Jahan N, *et al.*: **Dietary added bamboo charcoal can evoke *pangasianodon* growth and can reduce ammonia from culture medium**. *Int J Fish Aquac.* 2014; **6**(7): 87–93. [Publisher Full Text](#)
19. Pirarat N, Surinton B, Laddawan K, *et al.*: **Effect of activated charcoal-supplemented diet on growth performance and intestinal morphology of Nile tilapia (*Oreochromis niloticus*)**. *Thai J Vet Med.* 2015; **45**(1): 113–119. [Reference Source](#)
20. Boonanuntanasarn S, Khaomek P, Pitaksong T, *et al.*: **The effects of the supplementation of activated charcoal on the growth, health status and fillet composition-odor of Nile tilapia (*Oreochromis niloticus*) before harvesting**. *Aquacult Int.* 2014; **22**(4): 1417–1436. [Publisher Full Text](#)
21. Michael FR, Saleh NE, Shalaby SM, *et al.*: **Effect of different dietary levels of commercial wood charcoal on growth, body composition and environmental loading of red tilapia hybrid**. *Aquac Nutr.* 2017; **23**(1): 210–216. [Publisher Full Text](#)
22. Thu M, Koshio S, Ishikawa M, *et al.*: **Effects of dietary bamboo charcoal on growth parameters, apparent digestibility and ammonia nitrogen excretion of tiger puffer fish, *Takifugu rubripes***. *Aquac Sci.* 2009; **57**(1): 53–60. [Publisher Full Text](#)
23. Thu M, Koshio S, Ishikawa M, *et al.*: **Effects of supplementation of dietary bamboo charcoal on growth performance and body composition of juvenile Japanese flounder, *Paralichthys olivaceus***. *World Aquac Soc.* 2010; **41**(s2): 255–262. [Publisher Full Text](#)
24. Alonge TO, Lawal MO, Aderolu AZ, *et al.*: **Evaluation of soybean meal replacement with sesame seed meal using activated charcoal as an additive in the diet of African catfish juveniles, *Clarias gariepinus***. *Int J Aquat*

- Biol. 2016; **4**(1): 43–50.
[Publisher Full Text](#)
25. Aderolu AZ, Lawal MO, Adesola TT: **Effects of graded activated charcoal in rice husk diets for mud catfish, *Clarias gariepinus* juveniles (Teleostei: Clariidae).** *Iran J Ichthyol.* 2016; **3**(3): 203–209.
[Reference Source](#)
 26. Michael FR, Helal AM: **Rule of dietary activated wood charcoal on the growth and biochemical composition of Gilthead Seabream (*Sparus aurata*) reared under different stocking densities.** *Life Sci J.* 2018; **15**(4): 79–86.
[Publisher Full Text](#)
 27. Samadaii S, Bahrekazemi M: **The effect of diets containing different levels of active charcoal on growth performance, body composition, haematological parameters and possibility of heavy metals detoxification in big sturgeon (*Huso huso*).** *Aquac Res.* 2020; **51**(1): 91–101.
[Publisher Full Text](#)
 28. Firdus F, Muhammadar AA, Samadi S, *et al.*: **The effect of feeding with the addition of activated charcoal on feed conversion and survival of juvenile Giant Trevally (*Caranx ignobilis*).** *IOP Conf Ser: Earth Environ Sci.* 2020; **425**: 012051.
[Publisher Full Text](#)
 29. Infante JZ, Cahu C: **Development and response to a diet change of some digestive enzymes in sea bass (*Dicentrarchus labrax*) larvae.** *Fish Physiol Biochem.* 1994; **12**(5): 399–408.
[PubMed Abstract](#) | [Publisher Full Text](#)
 30. Pryor GS, Royes JB, Chapman FA, *et al.*: **Mannanoligosaccharides in fish nutrition: effects of dietary supplementation on growth and gastrointestinal villi structure in Gulf of Mexico sturgeon.** *N Am J Aquac.* 2003; **65**(2): 106–111.
[Publisher Full Text](#)
 31. Liu H, Guo X, Gooneratne R, *et al.*: **The gut microbiome and degradation enzyme activity of wild freshwater fishes influenced by their trophic levels.** *Sci Rep.* 2016; **6**: 24340.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 32. Özel OT, Coşkun I, Çakmak E: **Intestine villi morphology of black sea trout (*Salmo labrax*, Pallas, 1814).** *Limnol Fish.* 2018; **4**(1): 42–46.
[Publisher Full Text](#)
 33. Infante JZ, Cahu CL: **Ontogeny of the gastrointestinal tract of marine fish larvae.** *Comp Biochem Physiol C Toxicol Pharmacol.* 2001; **130**(4): 477–487.
[PubMed Abstract](#) | [Publisher Full Text](#)
 34. Wilson JM, Castro LFC: **Morphological diversity of the gastrointestinal tract in fishes.** *Fish physiology.* 2010; **30**: 1–55.
[Publisher Full Text](#)
 35. Burr G, Gatlin D III, Ricke S: **Microbial ecology of the gastrointestinal tract of fish and the potential application of prebiotics and probiotics in finfish aquaculture.** *World Aquaculture society.* 2005; **36**(4): 425–436.
[Publisher Full Text](#)
 36. Hoseinifar SH, Roosta Z, Hajimoradloo A, *et al.*: **The effects of *Lactobacillus acidophilus* as feed supplement on skin mucosal immune parameters, intestinal microbiota, stress resistance and growth performance of black swordtail (*Xiphophorus helleri*).** *Fish Shellfish Immunol.* 2015; **42**(2): 533–538.
[PubMed Abstract](#) | [Publisher Full Text](#)
 37. Javahery S, Nekoubin H, Moradlu AH: **Effect of anaesthesia with clove oil in fish (review).** *Fish Physiol Biochem.* 2012; **38**(6): 1545–1552.
[PubMed Abstract](#) | [Publisher Full Text](#)
 38. Purushothaman K, Lau D, Saju JM, *et al.*: **Morpho-histological characterisation of the alimentary canal of an important food fish, Asian seabass (*Lates calcarifer*).** *PeerJ.* 2016; **4**: e2377.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
 39. Osman AHK, Caceci T: **Histology of the stomach of *Tilapia nilotica* (Linnaeus, 1758) from the River Nile.** *J Fish Biol.* 1991; **38**(2): 211–223.
[Publisher Full Text](#)
 40. Muchlisin ZA, Firdus F, Samadi S, *et al.*: **Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with difference sources of activated charcoal.** *figshare.* Dataset. 2020.
<http://www.doi.org/10.6084/m9.figshare.12203525.v2>
 41. Muchlisin ZA, Firdus F, Samadi S, *et al.*: **Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with difference sources of activated charcoal.** *figshare.* Figure. 2020.
<http://www.doi.org/10.6084/m9.figshare.12301124.v2>
 42. Muchlisin ZA, Firdus F, Samadi S, *et al.*: **Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with difference sources of activated charcoal.** *figshare.* Figure. 2020.
<http://www.doi.org/10.6084/m9.figshare.12269606.v2>
 43. Quaiyum MA, Jahan R, Jahan N, *et al.*: **Effects of bamboo charcoal added feed on reduction of ammonia and growth of *Pangasius hypophthalmus*.** *J Aquac Res Dev.* 2014; **5**(6): 1–5.
[Publisher Full Text](#)
 44. Mahvi AH, Maleki A, Elslami A: **Potential of rice husk and rice husk ash for phenol removal from aqueous systems.** *Am J Appl Sci.* 2004; **1**(4): 321–326.
[Reference Source](#)
 45. Verheyden A, Roggeman M, Bouillon S, *et al.*: **Comparison between $\delta^{13}\text{C}$ of α -cellulose and bulk wood in the mangrove tree *Rhizophora mucronata*: implications for dendrochemistry.** *Chemical Geology.* 2005; **219**(1–4): 275–282.
[Publisher Full Text](#)
 46. Maryono M, Sudding S, Rahmawati R: **Preparation and quality analysis of coconut shell charcoal briquette observed by starch concentration.** *Jurnal Chemica.* 2013; **14**: 74–83.
 47. Sunardi S, Nurliani N: **Pemanfaatan arang aktif sekam padi dengan aktivator natrium karbonat (Na_2CO_3) 5% untuk mengurangi kadar besi (Fe) dalam air ledeng.** *Jurnal Teknologi Hasil Hutan.* 2008; **23**: 99–104.
[Reference Source](#)
 48. Maria NSL, Banu ASS: **Activated carbon from rice husk for treating dye waste water.** *Internasional Journal of Green Chemistry.* 2015; **1**(1): 1–9.
[Reference Source](#)
 49. Jamilatun S, Setyawan M, Salamah S, *et al.*: **Pembuatan arang aktif dari tempurung kelapa dengan aktivasi sebelum dan sesudah pirlisis.** 2015.
[Reference Source](#)
 50. Jasman J: **Uji coba arang aktif sekam padi sebagai media filtrasi dalam menurunkan kadar Fe pada air sumur bor di Asrama Jurusan Kesehatan Lingkungan Manado.** *Jurnal Kesehatan Lingkungan.* 2011; **1**(1): 49–53.
[Reference Source](#)
 51. Jauhari A: **Penanggulungan kadar besi (Fe) air sumur menggunakan arang aktif kayu bakau (*Rhizophora mucronata*. Lamck) dengan aktivator natrium karbonat.** *Jurnal Hutan Tropis Borneo.* 2009; **28**: 321–331.
 52. Guo J, Luo Y, Lua AC, *et al.*: **Adsorption of hydrogen sulphide (H_2S) by activated carbons derived from oil-palm shell.** *Carbon.* 2007; **45**(2): 330–336.
[Publisher Full Text](#)
 53. Nafis M, Zainuddin Z, Masyitha D: **Gambaran histologi saluran pencernaan ikan gabus (*Channa striata*).** *Jimvet.* 2017; **1**(2): 196–202.
[Reference Source](#)
 54. Samanya M, Yamauchi K: **Morphological changes of the intestinal villi in chickens fed the dietary charcoal powder including wood vinegar compounds.** *J Poult Sci.* 2001; **38**(4): 289–301.
[Publisher Full Text](#)
 55. Gilmore MS, Ferretti JJ: **The thin line between gut commensal and pathogen.** *Science.* 2003; **299**(5615): 1999–2002.
[Publisher Full Text](#)
 56. Yamauchi K: **Review of a histological intestinal approach to assessing the intestinal function in chickens and pigs.** *Anim Sci J.* 2007; **78**(4): 356–370.
[Publisher Full Text](#)
 57. Šabatková J, Kumprecht I, Zobač P, *et al.*: **The probiotic BioPlus 2B as an alternative to antibiotics in diets for broiler chickens.** *Acta Veterinaria Brno.* 2008; **77**: 569–574.
[Publisher Full Text](#)
 58. Choct M: **Managing gut health through nutrition.** *Br Poult Sci.* 2009; **50**(1): 9–15.
[Publisher Full Text](#)
 59. McLean E, Donaldson EM: **Absorption of bioactive proteins by the gastrointestinal tract of fish: a review.** *J Aquat Anim Health.* 1990; **2**(1): 1–11.
[Publisher Full Text](#)
 60. Ringø E, Olsen RE, Mayhew TM, *et al.*: **Electron microscopy of the intestinal microflora of fish.** *Aquaculture.* 2003; **227**(1–4): 395–415.
[Publisher Full Text](#)
 61. Mulyono P, Wibisono W: **Kinetika adsorpsi amonia dalam air dengan karbon aktif.** *Jurnal Media Teknik.* 2007; **2**: 26–42.
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Abdus Salam

Department of Aquaculture, Faculty of Fisheries, Bangladesh Agricultural University, Mymensingh, Bangladesh

Gut and intestinal biometrics of the giant trevally, *Caranx ignobilis*, fed an experimental diet with different sources of activated charcoal.

The title should be different not difference. Others are fine.

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Aquaculture, Aquaponics, fish nutrition

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 1

Reviewer Report 10 July 2020

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Abdus Salam

Department of Aquaculture, Faculty of Fisheries, Bangladesh Agricultural University, Mymensingh,

Bangladesh

- It seems the manuscript has potentials in the scientific world, and work was done and formatted with care. The abstract is concise but well written.
- The introduction has supported with up to date literature and aim and objectives are well described.
- Results supported with details and tables and figures were provided.
- The discussion was also well described and literature supported.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Aquaculture

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 15 June 2020

<https://doi.org/10.5256/f1000research.26249.r63939>

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Srikanta Samanta

Principal Scientist & Head (Acting) Riverine Ecology and Fisheries Division, ICAR-Central Inland Fisheries Research Institute, Barrackpore, India

The paper has communicated a short but interesting study on the beneficial effects of adding different types of charcoal as a dietary component of giant trevally. The addition of charcoal has significantly improved the gastro-intestinal microstructures including foveola gastrica & villous intestine and thereby, has the potentiality to improve the fish nutrition and health. The rice husk was recorded as the best source of charcoal with respect to 3 other sources including coconut shell, mangrove wood, and kernel palm shell. However, some deficiencies have been noticed which needs rectification.

1. In Abstract : This species was---: Better to use The species was---
2. Four activated charcoal sources were tested in this study ----but in Experimental Design mentioned ----A completely randomized experimental design with five different charcoal sources----
3. In Abstract : average total length, 10.26 ± 0.64 cm, In Experimental fish : total length, 10.28 ± 0.64 cm
4. ponds size 2 x 1.8 m. No unit given after 2. It may be 2 m X 1.8 m
5. (see Table 1) : see may be deleted
6. Table 1 : CaCo3 : CaCO₃
7. In Abstract : and 241.786 μ m --- In Figure 1 241.768
8. Figure 2 and 4 : Histological sample ---- may be replaced by ---- Histology
9. Sentence to be modified : Hence, a longer foveola gastrica and larger intestinal villi provide a larger surface area to absorb nutrients⁴³.
10. Sentence to be modified : A previous report indicated that rice husk charcoal contains 39% hemicellulose, 44% cellulose, and 30% lignin⁴⁴ : Since the addition of 3 parameters are giving values >100%, you may mention "up to" otherwise people will not understand.
11. Sentence to be modified/changed : The active charcoal likely acts as an adsorbent of metabolic pathogens in the intestine in the form of endotoxins and ammonia to improve intestinal function⁶¹. The sentence to be rewritten to increase clarity.
12. It has been noticed that the SE values have gone out of the bar area. Please check about it and reason behind. For the paper, the fish growth parameters may be given as supplements for better clarity.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Aquatic Chemistry

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 22 Jul 2020

Samadi Samadi, Syiah Kuala University, Banda Aceh, Indonesia

Dear Prof. Srikanta Samanta,
Thank you very much for your comment and we will revise the article based on your suggestions.
Best regards,
Samadi

Competing Interests: No competing interests were disclosed.

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